

Package ‘nCal’

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Title Nonlinear Calibration

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Depends R (>= 3.3.0), drc, gdata, gWidgets, kyotil

Suggests knitr, RUnit, rjags, gWidgetstcltk, nlme, R.rsp

VignetteBuilder R.rsp

Description Performs nonlinear calibration and curve fitting for data from Luminex, RT-PCR, ELISA, RPPA etc. Its precursor is Ruminex. This package is described in nCal: a R package for nonlinear calibration. Bioinformatics, <DOI:10.1093/bioinformatics/btt456>.

License GPL (>= 2)

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bcrm	<i>Bayesian Concentration-Response Model</i>
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Description

bcrm fit concentration-response curves with a Bayesian random effects model using JAGS

Usage

```
bcrm (formula, data,
      parameterization=c("gh", "classical"),
      error.model=c("norm", "t4", "mixnorm", "mix2", "replicate_re", "tar1", "lar1"),
      prior=c("cytokine", "BAMA", "RT-PCR", "ELISA", "default"),
      prior.sensitivity=c("none", "1", "2", "3", "4"),
      mean.model=c("5PL", "4PL"),
      n.iter=1e5, jags.seed=1, n.thin=NULL, T.init=NULL,
      keep.jags.samples=FALSE, standards.only=TRUE, n.adapt=1e3,
      t.unk.truth=NULL, params.true=NULL, # for simulation study use
      verbose=FALSE
)

## S3 method for class 'bcrm'
plot(x,
     assay_id=NULL, fit.2=NULL, fit.3=NULL,
     points.only=FALSE, all.lines.only=FALSE,
     same.ylim=FALSE, lty3=NULL, lcol2=NULL, lcol3=NULL,
     lcol=1, lwd=.1, lty=1, # for lines
     t=NULL, log="x", col.outliers=TRUE, pch.outliers=TRUE,
     use.dif.pch.for.replicate=FALSE, main=NULL,
     additional.plot.func=NULL, add=FALSE, ...
)

## S3 method for class 'bcrm'
print(x, ...)

## S3 method for class 'bcrm'
coef(object, type="gh", ...)

## S3 method for class 'bcrm'
vcov(object, type="gh", ...)

## S3 method for class 'bcrm'
getVarComponent(object, ...)
```

```
get.single.fit (fit, assay_id)
```

Arguments

formula	formula. Gives the response column and concentration column.
data	a data frame. Each row represents the measurement from one well/bead_type. See details
parameterization	string.
error.model	string.
prior	string.
mean.model	mean model
pch.outliers	pch for outliers
n.iter	a number indicating the number of iterations to run.
jags.seed	a number to seed the random number generator used within jags.
n.thin	a number specifying the thinning factor applied to the jags samples.
keep.jags.samples	boolean. If TRUE, the fit object being returned has an element named "jags.samples". coef samples will always be saved in "coef.samples".
t.unk.truth	True unknown concentrations, for simulation study use only.
params.true	True curve parameters, for simulation study use only.
T.init	a integer vector. Initial values for mixture indicators.
prior.sensitivity	integer. A number between 1 and 4. Change priors.
standards.only	boolean. If TRUE, data is subset to standard samples only.
n.adapt	integer. Passed to jags.model. If 0, then no adaptation happens and reproducible results can be obtained from jags.model.
verbose	boolean. If TRUE, debug messages are printed.
x	bcrm fit object.
object	bcrm fit object.
type	string. 5PL parameterization.
fit	bcrm fit object.
...	...
assay_id	string. Label for the assay run.
add	Boolean. If TRUE, adding to an existing plot.
lcol	integer. Line color.
fit.2	a bcrm object. A second fit object to be plotted together with x.
lwd	numeric. Line width.
points.only	Boolean. If TRUE, only the data points are plotted and not the fitted curves

<code>all.lines.only</code>	Boolean. If TRUE, only the fitted curves are plotted.
<code>t</code>	a numeric vector. The log concentrations.
<code>same.ylim</code>	Boolean. If TRUE, all fitted curves are plotted with the same ylim.
<code>lty3</code>	integer. lty for plotting fit.3.
<code>fit.3</code>	bcrm object. A third optional bcrm object to be plotted.
<code>lcol2</code>	integer. Line color for plotting fit.2.
<code>lcol3</code>	integer. Line color for plotting fit.3.
<code>col.outliers</code>	Boolean. If TRUE, outliers are colored differently.
<code>main</code>	string.
<code>lty</code>	integer. Line type for plotting x.
<code>log</code>	string. If it is "x", then the x axis is labeled on the scale of concentration; otherwise, it is labeled on the scale of log concentration.
<code>additional.plot.func</code>	function, to be called after plotting each fit
<code>use.dif.pch.for.replicate</code>	boolean

Details

data is expected to contain one to many plates with the same analyte.

- `well_role` Defines the role of a well. This should be from Standard, Unknown, Standard wells are used to generate standard curves, and concentrations of the substance in the Unknown well will be estimated
- `assay_id` Identifies an assay, which is defined to be a collection of Standard and non-Standard wells. Measured fi from the Standard wells are used to create a set of standard curves, one of each bead type. Based on the standard curves and the measured fi from the non-Standard wells, concentrations of the substance in the non-Standard wells will be estimated. An assay can be a plate, if every plate has Standard wells; or it can be multiple plates run by one technician at one time, if there are only one set of Standard wells on these plates
- `dilution` Standard samples are often prepared by starting with one sample and doing serial dilutions. Unknown samples may be measured at several dilutions so that one of the dilutions may fall into the more reliable region of the standard curve
- `replicate` Index of technical replicates for a sample. Typical values are 1 or 2. May be used in plotting. Optional
- `expected_conc` Standard samples have expected concentrations. If this column is present, the `dilution` and `starting_conc` are optional and will not be used. This column does not apply to non-Standard samples

Main error.model supported: `drc`, `classical_norm`, `classical_t4`, `classical_mixnorm`, `classical_lar1`, `gh_norm`, `gh_mixnorm`, `gh_lar1` Also support: `classical_replicate_re`, `gh_replicate_re`, `gh_tar1`

Only two replicates are supported for now for the correlated noise models.

Sometimes `jags.model` fails with one `.RNG.seed`. The function will increase it by 1 and try again. The function tries three times before giving up.

Value

An object of type bcrm.

Author(s)

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References

Fong, Y., Wakefield, J., DeRosa, S., Frahm, N. (2012) A robust Bayesian random effects model for nonlinear calibration problems, *Biometrics*, 68:1103-1112.

Examples

```

set.seed(1)
log.conc=log(1e4)-log(3)*9:0
n.replicate=2
fi=simulate1curve (p.eotaxin[1,], rep(log.conc,each=n.replicate), sd.e=0.3)
dat.std=data.frame(fi, expected_conc=exp(rep(log.conc,each=n.replicate)), analyte="test",
  assay_id="assay1", sample_id=NA, well_role="Standard", dilution=rep(3**(9:0), each=n.replicate))
# add unknown
dat.unk=rbind(
  data.frame(fi=exp(6.75), expected_conc=NA, analyte="test", assay_id="assay1",
    sample_id=1, well_role="Unknown", dilution=1)
  , data.frame(fi=exp(6.70), expected_conc=NA, analyte="test", assay_id="assay1",
    sample_id=2, well_role="Unknown", dilution=1)
  , data.frame(fi=exp(3), expected_conc=NA, analyte="test", assay_id="assay1",
    sample_id=3, well_role="Unknown", dilution=1)
  , data.frame(fi=exp(4.4), expected_conc=NA, analyte="test", assay_id="assay1",
    sample_id=4, well_role="Unknown", dilution=10)
)
dat=rbind(dat.std, dat.unk)
# second plate
fi=simulate1curve (p.eotaxin[2,], rep(log.conc,each=n.replicate), sd.e=0.3)
dat.std=data.frame(fi, expected_conc=exp(rep(log.conc,each=n.replicate)), analyte="test",
  assay_id="assay2", sample_id=NA, well_role="Standard", dilution=rep(3**(9:0), each=n.replicate))
# add unknown
dat.unk=rbind(
  data.frame(fi=exp(6.75), expected_conc=NA, analyte="test", assay_id="assay2",
    sample_id=1, well_role="Unknown", dilution=1)
  , data.frame(fi=exp(6.70), expected_conc=NA, analyte="test", assay_id="assay2",
    sample_id=2, well_role="Unknown", dilution=1)
  , data.frame(fi=exp(3), expected_conc=NA, analyte="test", assay_id="assay2",
    sample_id=3, well_role="Unknown", dilution=1)
  , data.frame(fi=exp(4.4), expected_conc=NA, analyte="test", assay_id="assay2",
    sample_id=4, well_role="Unknown", dilution=10)
)
dat=rbind(dat, dat.std, dat.unk)

fits = bcrm(log(fi)~expected_conc, dat, parameterization="gh", error.model="norm",

```

```

      prior="cytokine", n.iter=6e3)
par(mfrow=c(1,2))
plot(fits)

## Not run:
# takes longer

# Example from Fong et al. (2012)
fits.t4 = bcrm (log(fi)~expected_conc, dat.QIL3, parameterization="gh", error.model="t4",
  prior="cytokine")
par(mfrow=c(2,3))
plot(fits.t4)

fits.norm = bcrm (log(fi)~expected_conc, dat.QIL3, parameterization="gh", error.model="norm",
  prior="cytokine")
par(mfrow=c(2,3))
plot(fits.norm)

## End(Not run)

```

 crm.fit

Fit Concentration Response Model

Description

crm.fit can fit a constant or power variance function or log transform both sides.

Usage

```

crm.fit (formula, data, fit.4pl=FALSE, var.model=c("constant","power"),
  robust="mean", method=c("gls-pl","gnls","mle"), max.iter=50, reltol=1e-3,
  gof.threshold=0.2, log.both.sides=FALSE, verbose=FALSE)

```

```

## S3 method for class 'crm'
deviance(object, ...)
## S3 method for class 'crm'
print(x, ..., digits=3)
## S3 method for class 'crm'
lines(x, ...)
## S3 method for class 'crm'
coef(object, parameterization=c("cla","gh","ed50b","edb50"), ...)

```

Arguments

formula	
data	
fit.4pl	Boolean
var.model	string
robust	string
method	string
max.iter	number
digits	number
reltol	numeric
gof.threshold	numeric
verbose	Boolean
log.both.sides	Boolean, log transform both sides
object, x	crm object
parameterization	string, output parameterization
...	additional argument

Details

crm.fit implements an iterative method for estimating a model with power variance. method: gls-pl means GLS-PL (see reference) log.both.sides: transform both sides (see reference)

Value

An object of crm and drm type.

var.power estimated power parameter in the power variance function

References

Fong, Y., Yu, X. (2014) Transformation Model Choice in Nonlinear Regression Analysis of Serial Dilution Assays, submitted

Examples

```
## Not run:
dat.std=dat.QIL3[dat.QIL3$assay_id=="LMX001",]

# run 3 iter to save time for examples
fit.1=crm.fit(fi~expected_conc, dat.std, var.model="power", verbose=TRUE, max.iter=2)
fit.2=crm.fit(log(fi)~expected_conc, dat.std, verbose=TRUE)
fit.3=crm.fit(log(fi)~expected_conc, dat.std, var.model="power", verbose=TRUE, max.iter=2)

sapply(list(fit.1, fit.2, fit.3), coef)
```

```

fit.1$var.power
fit.2$var.power
fit.3$var.power

plot(fit.1, log="xy", type="all", lwd=3, pch="*")
lines(fit.2, expy=TRUE, col=2, lwd=3)
lines(fit.3, expy=TRUE, col=4, lty=2, lwd=3)

## End(Not run)

```

dat.QIL3

An example for hierarchical modeling used in Fong, Wakefield, De Rosa, Frahm (2012)

Description

An example for hierarchical modeling used in Fong, Wakefield, De Rosa, Frahm (2012)

Format

A data frame with 120 observations on the following 13 variables.

`well` a character vector. Well identifier on a microplate.

`assay_id` a character vector. Assay identifier.

`analyte` a character vector. Substance to be measured.

`well_role` a character vector. Defines the role of a well. See `?ncal` for more information.

`beadct` a numeric vector. Number of beads in the well, specific to multiplex bead array assay.

`dilution` a numeric vector. The dilution factor of a sample.

`expected_conc` a numeric vector. The expected concentrations of a sample.

`fi` a numeric vector. Fluorescence intensity readout.

`ptid` a integer vector. Participant ID.

`sample_id` a Boolean vector. All NA.

`thawdt` a character vector. All empty string

`visit` a integer vector. All NA.

`replicate` a numeric vector. Technical replicate identifier.

References

Fong, Y., Wakefield, J., DeRosa, S., Frahm, N. (2012) A robust Bayesian random effects model for nonlinear calibration problems, *Biometrics*, 68:1103-1112.

drm.fit	<i>Fit drm</i>
---------	----------------

Description

drm.fit fit concentration-response curves using drm function from drc package.

Usage

```
drm.fit (formula, data, robust="mean", fit.4pl=FALSE, w=NULL, gof.threshold=.2,  
        verbose=FALSE, bcVal = NULL, bcAdd = 0)
```

```
## S3 method for class 'drc'  
getVarComponent(object, ...)
```

Arguments

formula	a formula object.
data	a data frame object.
robust	a string. Passed to drm. See ?drm for more details.
fit.4pl	boolean. If TRUE, 4PL model is fitted. If FALSE, 5PL model is fitted.
gof.threshold	a threshold to determine when to try more self start functions
w	weights
bcVal	numeric, passed to drm
bcAdd	numeric, passed to drm
...	...
verbose	Boolean. If TRUE, verbose messages are printed.
object	a drm object.

Details

drm.fit differs from drc::drm in several aspects.

- (1) It tries several self start functions in order to get better fits.
- (2) It uses gof.threshold to report lack of fit.
- (3) It tried to determine whether the standard deviation of the parameter estimates can be estimated.

Value

An object of type drm.

Examples

```
# simulate a dataset
set.seed(1)
log.conc=log(1e4)-log(3)*9:0
n.replicate=2
fi=simulate1curve (p.eotaxin[1,], rep(log.conc,each=n.replicate), sd.e=0.2)
dat.std=data.frame(fi, expected_conc=exp(rep(log.conc,each=n.replicate)), analyte="Test",
  assay_id="Run 1", sample_id=NA, well_role="Standard", dilution=rep(3**(9:0), each=n.replicate),
  replicate=rep(1:n.replicate, 10))

fit = drm.fit(log(fi) ~ expected_conc, dat = dat.std)
plot(fit, log="xy")
fit
```

ED5PL

*Functions Related to 5PL Function***Description**

5PL functions.

Usage

```
FivePL.t (t,param)
FivePL.t.func (param)
FivePL.x (x,param)
FivePL.x.inv (y,param)
FivePL.x.inv.func (param)
FivePL.t.inv (y,param)
FivePL.t.inv.func (param)
FourPL.x.inv (y, param)
FourPL.x (x, param)
FourPL.t.func (param)
cla2gh (param)
gh2cla (param)
cla2ed50 (param)
cla2ed50b (param)
ed502cla (param)
ed50b2cla (param)
get.curve.param.list (param)
simulate1curve (param, t, sd.e=0.1, expy=TRUE, gamma=0)
vp11.deriv (x,param)
vp11.deriv.func (param)
vp12.deriv (x,param)
vp12.deriv.func (param)
vp13.deriv (x,param)
```

vp13.deriv.func (param)

ED5PL (param, tao)

Arguments

param	vector of numbers. Parameters of the 5PL curve.
t	numeric vector. Log concentrations.
x	numeric vector. Concentrations.
y	numeric vector. Response.
sd.e	sd.e
tao	vector of numbers or single number. Effective doses. Vectorized for this argument.
expy	Boolean. Controls whether to exponentiate y
gamma	power variance function parameter

Details

FivePL.t and other functions are vectorized for the x and the y arguments.

Four parameterizations are implemented. Classical: b,c,d,e,f gh: c,d,f,g,h ED50: c,d,f,b,tao(ED50) ED50b: c,d,f,h,tao(ED50). This is called tao-h in Cumberland et al. (2014)

Author(s)

Youyi Fong <yfong@fhcrc.org>, Xuesong Yu, William N. Cumberland

Examples

```
FivePL.t(5:6, p.eotaxin[1,])
FivePL.t.func(p.eotaxin[1,])
FivePL.x.inv(c(4,5,11), p.eotaxin[1,])
FivePL.t.inv.func(p.eotaxin[1,])
```

elisa.R.gh

ELISA prior for gh Parameterization

Description

Priors.

Format

elisa.R.gh is a 5x5 diagonal matrix. rowNames/colNames: "c" "d" "g" "logh" "logf" elisa.mean.distr.gh is a 2x5 matrix. colNames: "c" "d" "g" "logh" "logf". rowNames: mean, prec

`get.abc`*Compute a measure of distance between two curves.*

Description

abc criterion is area between curves divided by the width of t.range. S1 criterion between two curves is defined as the integrated squared distance divided by the width of t.range. S2 is relative bias divided by the width of t.range.

Usage

```
get.abc(p1, p2, t.range)
get.S1(p1, p2, t.range)
get.S2(p1, p2, t.range)
get.abs.dev(p1, p2, t.range, y.range)
```

Arguments

p1	a vector of coefficients specify the first curve
p2	a vector of coefficients specify the second curve
t.range	a vector two real numbers. The range of log standard concentrations. The first one is the minimum t and the second one is the maximum t
y.range	

Details

p1 and p2 can be in either classical or gh parameterization.

Value

a real number

Examples

```
get.abc(p.eotaxin[1,], p.eotaxin[2,], t.range=log(c(0.51,1e4)))
get.S1(p.eotaxin[1,], p.eotaxin[2,], t.range=log(c(0.51,1e4)))
get.S2(p.eotaxin[1,], p.eotaxin[2,], t.range=log(c(0.51,1e4)))
get.abs.dev(p.eotaxin[1,], p.eotaxin[2,], t.range=log(c(0.51,1e4)), y.range=c(5,6))
```

getConc	<i>Concentration Estimation</i>
---------	---------------------------------

Description

Estimate analyte concentrations based on observed outcome and a fitted curve.

Usage

```
getConc(fit, y, n.replicate = 1, check.out.of.range = 1, x.range = NULL,
        y.range = NULL, verbose = FALSE)
```

Arguments

fit	drc object or bcrm object
y	numeric vector. Observed outcome in the samples of interest. Each element corresponds to one sample
n.replicate	integer. Number of replicates that are averaged to generate y
check.out.of.range	integer.
x.range	vector of 2 numbers. The minimal and maximal concentration of the standard samples
y.range	vector of 2 numbers. The minimal and maximal observed response of the standard samples
verbose	Boolean.

Details

Vectorized for y.

gnls.fit	<i>Fit with gnls function</i>
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Description

Fit with gnls function

Usage

```
gnls.fit(formula, data, fit.4pl = FALSE, startVal = NULL, varFun = nlme::varPower(),
        verbose = FALSE)
```

Arguments

formula
data
fit.4pl
startVal
varFun
verbose

Details

When startVal is given, varPower(value) does not seem to change numerical values of the fit

hier.model.ex.2

An example for hierarchical modeling used in Fong, Yu et al.

Description

An example for hierarchical modeling used in Fong, Yu et al.

Format

A data frame with 120 observations on the following 8 variables.

assay_id a character vector. Assay identifier.

analyte a character vector. Substance to be measured.

expected_conc a numeric vector. The expected concentrations of a sample.

dilution a numeric vector. The dilution factor of a sample.

fi a numeric vector. Fluoresence intensity readout.

well_role a character vector. Defines the role of a well. See ?ncal for more information.

sample_id a character vector. Sample identifier.

replicate a numeric vector. Technical replicate identifier.

References

Fong, Y., Yu, X., Sebestyen, K., Gilbert, P. and Self, S. (2013) nCal: a R package for nonlinear calibration. Submitted.

 ncal

Main function for the nCal package

Description

ncal fits standard curves and estimates unknown sample concentrations. rumi exists for backwards compatibility.

Usage

```
## S3 method for class 'formula'
ncal(formula, data,
      bcrm.fit=FALSE, bcrm.model="t4", robust="mean", bcrm.prior="default",
      force.fit=TRUE, fit.4pl=FALSE, return.fits=FALSE,
      plot=TRUE, auto.layout=TRUE, plot.se.profile=TRUE, plot.log="x", plot.unknown=TRUE,
      test.LOD=FALSE, find.LOD=FALSE, find.LOQ=FALSE, grid.len=50, lod.ci=95,
      unk.replicate=NULL, find.best.dilution=FALSE, unk.median=FALSE,
      control.jags=list(n.iter=1e5, jags.seed=1, n.thin=NULL,
                       keep.jags.samples=FALSE, n.adapt=1e3),
      cex=.5, additional.plot.func=NULL, check.out.of.range=1, xlab=NULL, ylab=NULL,
      main=NULL,
      var.model=c("constant", "power"), log.both.sides=FALSE,
      control.crm.fit=list(max.iter=20),
      verbose=FALSE,
      ...)

## S3 method for class 'character'
ncal(file, is.luminex.xls, formula, bcrm.fit, verbose=FALSE, ...)

rumi(data, ...)

ncalGUI (verbose=FALSE)
```

Arguments

formula	a formula. If the first argument is formula, bcrm.formula is called.
data	a data frame. If the first argument is dat, bcrm.data.frame is called. Each row of the data frame represents the measurement from one well/bead_type
bcrm.fit	Boolean. If TRUE, a Bayesian random effects model is fitted, else a drm model is fitted.
bcrm.model	string. Noise model used in bcrm. See error.model in the help file for bcrm for more information.
bcrm.prior	string.

<code>var.model</code>	constant: constant variance. power: power variance function
<code>robust</code>	string. Passed to <code>drm</code> .
<code>plot</code>	a Boolean, default FALSE. This controls whether or not to make plots of the fitted curves and the standard error profiles
<code>plot.unknown</code>	a Boolean. This controls whether or not to make plots of the fitted curves with unknown sample concentrations layered on top
<code>auto.layout</code>	a Boolean, default TRUE This controls whether or not to let <code>bcrm</code> controls the layout of the plots
<code>test.LOD</code>	a Boolean, default TRUE This controls whether or not to test for limits of detection
<code>plot.se.profile</code>	a Boolean, default FALSE. This controls whether or not to make plots of standard error profile, this doubles as <code>find.LOQ</code>
<code>force.fit</code>	a Boolean, default FALSE. If FALSE, return NULL when the goodness of fit is bad; otherwise, always return the fit
<code>find.LOD</code>	Boolean. Controls whether or not to compute limits of detection.
<code>find.LOQ</code>	Boolean. Controls whether or not to compute limits of quantification.
<code>find.best.dilution</code>	a Boolean, default FALSE. When there are more than one dilutions for a given non-standard sample, this controls whether or not to find the best dilution
<code>return.fits</code>	a Boolean, default FALSE. This controls whether or not to return as an attribute the curve fits
<code>grid.len</code>	an integer, default 500 This determines the resolution of the standard error profile plot and limits of quantification
<code>unk.replicate</code>	an integer, default NULL. This is the number of replicates for an unknown sample (at a single dilution if multiple dilutions are available)
<code>unk.median</code>	a Boolean, default FALSE. If TRUE, median of unknown replicates are used to estimate conc, instead of mean
<code>fit.4pl</code>	a Boolean, default FALSE
<code>lod.ci</code>	an integer, default 95. If default, one sided 50 percent CI is used to determine LOD_i .
<code>plot.log</code>	a string, default "x". Used to populate the log argument of the plot function.
<code>control.jags</code>	list. Parameters controlling the behavior of posterior sampling by JAGS.
<code>is.luminex.xls</code>	Boolean. Indicates whether the file is in the Luminex Excel file format outputted by the Bio-Plex software.
<code>cex</code>	numeric. Passed to plot functions.
<code>main</code>	plotting parameter
<code>xlab</code>	plotting parameter
<code>ylab</code>	plotting parameter
<code>additional.plot.func</code>	function. Will be called after the first panel is drawn.

check.out.of.range	integer. If 1, an estimated concentration is set to half of the smallest standard concentration, or the largest standard if it is outside the range of standard concentration. If 2, do this only when the estimated concentration is 0 or Inf
verbose	a Boolean, default FALSE. This controls whether or not to print detailed messages
log.both.sides	Boolean, whether to log transform both sides of the formula
control.crm.fit	list. Parameters controlling the behavior of crm.fit
file	string. Name of the data file.
...	passed to the function plotting fitted curves

Details

Certain columns are expected of the input data frame:

- **analyte** Identifies a type of bead. Beads can be named after the substance that is recognized by the antibody that coats the bead, or they can be named by the protein that coats the bead. This is also known as analyte or antigen in different contexts. To be retro-compatible, this column can also be named `analyte`
- **well_role** Defines the role of a well. This should be from `Standard`, `Unknown`, `Standard` wells are used to generate standard curves, and concentrations of the substance in the `Unknown` well will be estimated
- **assay_id** Identifies an assay, which is defined to be a collection of `Standard` and non-`Standard` wells. Measured `fi` from the `Standard` wells are used to create a set of standard curves, one of each bead type. Based on the standard curves and the measured `fi` from the non-`Standard` wells, concentrations of the substance in the non-`Standard` wells will be estimated. An assay can be a plate, if every plate has `Standard` wells; or it can be multiple plates run by one technician at one time, if there are only one set of `Standard` wells on these plates
- **dilution** `Standard` samples are often prepared by starting with one sample and doing serial dilutions. `Unknown` samples may be measured at several dilutions so that one of the dilutions may fall into the more reliable region of the standard curve
- **replicate** Index of technical replicates for a sample. Typical values are 1 or 2. May be used in plotting. Optional
- **sample_id** Identifies a non-`Standard` sample. One sample may be measured in replicates and/or in several dilutions. Should be defined meaningfully for `Unknowns`

If no formula is specified, we also expect

- **fi** Fluorescence intensity
- **starting_conc** `Standard` samples are often prepared by starting with one sample and doing serial dilutions. This column does not apply to non-`Standard` samples
- **expected_conc** `Standard` samples have expected concentrations. If this column is present, the `dilution` and `starting_conc` are optional and will not be used. This column does not apply to non-`Standard` samples

The program processes each assay separately. For each assay, each bead_type is processed separately. First `fit.drc()` is called to fit dose-response curves using information from the Standard wells. A plot of the fitted curve is generated. Then for each non-Standard sample, the number of dilutions is determined. For each dilution, $\log(\text{fi})$ from replicate wells are averaged and used to compute the estimated concentration and the standard error of the estimate.

Bad fits can happen for three reasons. 1) An error occurs in `drm`. 2) Estimated variance for some curve parameter is negative. 3) A goodness of fit statistics is above a pre-set threshold. When a bad fit happens, by default we plot the observed `fi`, not the fitted curve, and do not proceed to estimation of concentration. However, when `force.fit` is set to `TRUE`, in the latter two cases of bad fits, we will proceed to plot the fitted curve and use it to estimate concentrations. In the second case, the standard errors of the estimated concentrations are set to `NA`. In the third case, standard errors will be computed. The default value for `force.fit` is set to `FALSE` to promote caution.

An important factor affecting the success of the fitting procedure is the choice of self start function. In addition to trying the default self start function in the most current `drc` package, we also try a self start function that is based on four parameter log-logistic model, and the self start function in version 1.5.2 of the `drc` package.

Standard errors convey the uncertainty we have about estimated concentrations. Standard error profiles show the relationship between standard errors and estimated concentrations. A sequence of hypothetical `fi` between the expected `fi` for the smallest and largest concentrations on the Standard curve are generated. The number of hypothetical `fi` is controlled by the variable `grid.len`. For each hypothetical `fi`, the estimated concentration and associated standard error are computed. There are two sources of uncertainty in an estimated concentration. One part of the uncertainty, we call replication-sensitive, comes from the fact that the measured `fi` has measurement error in it. It can be reduced by doing replicates of the non-Standard samples. The number of replicates used in computing standard error profile is controlled by the variable `rep.se.profile`. The other part, we call replication-insensitive, comes from the fact that there are uncertainty about the Standard curve as well. The total uncertainty is plotted in black, the replication-sensitive in red, and the replication-insensitive in blue. The two parts of uncertainty add up to the total not on the standard error scale, but on the variance scale, which is the square of standard errors. We choose to plot on the standard error scale because this scale is more comparable to the estimated concentration.

When `unk.replicate` is `NULL` (default), the program sets it to the number of replicates of the first non-Standard sample it encounters; if there is no non-Standard sample, it is set to 1. This value is only used in the computation of LOD. It does not affect `se` of unknown sample conc estimate.

When `find.best.dilution` is `false`, the estimated concentrations for all dilutions (after adjusting for dilutions) are returned; otherwise, the best dilution, determined as the dilution having the smallest standard error, is returned.

When `plot` is `FALSE`, no plot is made. When `plot` is `TRUE`, but `plot.rep.profile` is `false`, only fitted curves are plotted.

When `auto.layout` is `TRUE`, `bcrn` will choose a layout that makes sense for showing one analyte per page. For example, if `plot.se.profile` is `TRUE`, it will be 2x2.

When `test.LOD` is `true`, estimated concentration will be tested against the null hypothesis that it is not different from the extremem data points of the standard samples

The difference between `Inf` and `NA` for `se`: `Inf` is if the `fi` is outside certain ranges, `NA` is if the `s.e.` is bad for some reason.

`find.LOD`, return an attribute "LOD", that is the lowest and highest concentration detectable, defined in the sense that ... `plot.se.profile` also leads to the return of an attribute "LOQ", that is the lowest

and highest concentration at which percent cv is at 20

drm requires that weights are evaluated in the global env. `drm.weights (drm.fit.R)` is our answer to that.

Value

A data frame, each row contains one estimated concentration. All columns of the input data frame are preserved, in addition two new columns are added: `est.log.conc` and `se.est.log.conc`. `est.log.conc` contains estimated log concentration, while `se` is the standard error of the `est.log.conc`.

When `return.fits` is `TRUE`, the returned data frame has an attribute "fits", that is a list of the fitted curves.

Standard curves are plotted. When `plot.se.profile` is `TRUE`, error profiles are also plotted. Two error profile are plotted, one is the standard error of the estimated log concentration versus estimated log concentration. The other is $100 \times$ (standard error of estimated concentration / estimated concentration) versus log estimated concentration.

Author(s)

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References

Fong, Y., Yu, X., Sebestyen, K., Gilbert, P. and Self, S. (2013) nCal: a R package for nonlinear calibration. *Bioinformatics*

Examples

```
#begin=Sys.time()

# basic example

# simulate a dataset
set.seed(1)
log.conc=log(1e4)-log(3)*9:0
n.replicate=2
fi=simulate1curve (p.eotaxin[1,], rep(log.conc,each=n.replicate), sd.e=0.2)
dat.std=data.frame(fi, expected_conc=exp(rep(log.conc,each=n.replicate)), analyte="Test",
  assay_id="Run 1", sample_id=NA, well_role="Standard", dilution=rep(3**(9:0), each=n.replicate),
  replicate=rep(1:n.replicate, 10))
# add unknown
dat.unk=rbind(
  data.frame(fi=exp(6.75), expected_conc=NA, analyte="Test", assay_id="Run 1", sample_id=1,
    well_role="Unknown", dilution=1, replicate=1)
  , data.frame(fi=exp(6.70), expected_conc=NA, analyte="Test", assay_id="Run 1", sample_id=2,
    well_role="Unknown", dilution=1, replicate=1)
  , data.frame(fi=exp(3), expected_conc=NA, analyte="Test", assay_id="Run 1", sample_id=3,
    well_role="Unknown", dilution=1, replicate=1)
  , data.frame(fi=exp(4.4), expected_conc=NA, analyte="Test", assay_id="Run 1", sample_id=4,
    well_role="Unknown", dilution=10, replicate=1)
```

```

)
dat=rbind(dat.std, dat.unk)

## Not run:
# to save some time

# does drm fit
out = ncal(log(fi)~expected_conc, dat, return.fits = TRUE, plot.se.profile=TRUE)
fit=attr(out, "fits")[[1]]

# does jags fit and collect 1e5 posterior samples, it may be better to set n.iter higher
out.norm = ncal(log(fi)~expected_conc, dat, bcrm.fit=TRUE, bcrm.model="norm",
  return.fits = TRUE, plot.se.profile=TRUE,
  control.jags=list(n.iter=1e4), verbose=FALSE)
fit.norm=attr(out.norm, "fits")

# compare drm fit with bcrm fit
rbind(out, out.norm)
rbind(cloglik(coef(fit)), coef(fit.norm))
rbind(sqrt(diag(vcov(fit))), sqrt(diag(vcov(fit.norm, type="classical")))) )
sd.est = c(summary(fit)$rseMat[1], getVarComponent.bcrm(fit.norm)^.5)
sd.est

# do ncal with imported data from a raw Luminex output file
# the importing step can step a litte time
out = ncal (paste(system.file(package="nCal")[1], '/misc/02-14A22-IgA-Biotin-tiny.xls', sep=""),
  is.luminex.xls=TRUE, formula=log(fi)~expected_conc, bcrm.fit=FALSE, return.fits=TRUE)
fit=attr(out, "fits")[[1]]
getConc(fit, c(5,6))

## End(Not run)

#end=Sys.time();print(end-begin)

```

p.eotaxin

5PL parameters from an Eotaxin dataset.

Description

5PL parameters from an Eotaxin dataset.

Details

x axis is always drawn in the log scale.

Author(s)

Youyi Fong <yfong@fhcrc.org>

read.luminex.xls	<i>Read a Luminex File</i>
------------------	----------------------------

Description

Read a Luminex raw output .xls file

Usage

```
read.luminex.xls (file, verbose = FALSE, sheets = NULL, assay_id=NULL,
  na.strings = c("NA", "#DIV/0!"), ..., perl = "perl")
```

Arguments

file	string. The file name.
verbose	Boolean. If TRUE, some debug messages are printed.
sheets	vector of strings. The names of the sheets to be read. If NULL, all sheets are read.
assay_id	string. Used to name the assay that generated the data file. If not provided, a random number is generated as the name.
na.strings	vector of strings. Strings to be mapped to NA.
...	
perl	string. Command used to execute perl. If perl.exe is not in the path, the full path can be specified, e.g. "C:/Perl64/bin/perl.exe"

Examples

```
#begin=Sys.time()

# example from https://www.labkey.org/wiki/home/Documentation/page.view?name=luminexFileFormats
dat = read.luminex.xls(paste(system.file(package="nCal")[1],
  '/misc/02-14A22-IgA-Biotin-tiny.xls', sep=""), verbose=TRUE)
out = ncal(log(fi)~expected_conc, dat, return.fits = TRUE, plot.se.profile=FALSE)
out

#end=Sys.time();print(end-begin)
```

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