Package 'netmeta'

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Title Network Meta-Analysis using Frequentist Methods

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Depends R (>= 4.0.0), meta (>= 5.5-0)

Imports magic, MASS, ggplot2 (>= 3.0.0), metafor

Suggests colorspace, rgl, hasseDiagram (>= 0.1.3), grid, mvtnorm, gridExtra, igraph (>= 1.0.1), tictoc, writexl

URL https://github.com/guido-s/netmeta

https://link.springer.com/book/10.1007/978-3-319-21416-0

Description A comprehensive set of functions providing frequentist methods for network meta-analysis and supporting Schwarzer et al. (2015) <DOI:10.1007/978-3-319-21416-0>, Chapter 8 ``Network Meta-Analysis'':
- frequentist network meta-analysis following Rücker (2012) <DOI:10.1002/jrsm.1058>;
- net heat plot and design-based decomposition of Cochran's Q according to Krahn et al. (2013) <DOI:10.1186/1471-2288-13-35>;
- measures characterizing the flow of evidence between two treatments by König et al. (2013) <DOI:10.1002/sim.6001>;
- ranking of treatments (frequentist analogue of SUCRA) according to Rücker & Schwarzer (2015) <DOI:10.1186/s12874-015-0060-8>;
- partial order of treatment rankings ('poset') and Hasse diagram for 'poset' (Carlsen & Bruggemann, 2014) <DOI:10.1002/cem.2569>; (Rücker & Schwarzer, 2017) <DOI:10.1002/jrsm.1270>;
- split direct and indirect evidence to check consistency (Dias et al., 2010) <DOI:10.1002/sim.3767>, (Efthimiou et al., 2019) <DOI:10.1002/sim.8158>;
- league table with network meta-analysis results;
- additive network meta-analysis for combinations of treatments (Rücker et al., 2020) <DOI:10.1002/bimj.201800167>;
- network meta-analysis of binary data using the Mantel-Haenszel or non-central hypergeometric distribution method (Efthimiou et al., 2019) <DOI:10.1002/sim.8158>;
- 'comparison-adjusted' funnel plot (Chaimani & Salanti, 2012) <DOI:10.1002/jrsm.57>;
- automated drawing of network graphs described in Rücker & Schwarzer (2016) <DOI:10.1002/jrsm.1143>;
- rankograms and ranking by SUCRA;

- contribution matrix as described in Papakonstantinou et al. (2018) <DOI:10.12688/f1000research.14770.3> and Davies et al. (2021) <arXiv:2107.02886>.

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RoxygenNote 7.2.1

NeedsCompilation no

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R topics documented:

netmeta-package *netmeta: Brief overview of methods and general hints*

Description

R package **netmeta** provides frequentist methods for network meta-analysis and supports Schwarzer et al. (2015), Chapter 8 on network meta-analysis https://link.springer.com/book/10.1007/978-3-319-21416-0.

Details

R package **netmeta** is an add-on package for **meta** providing the following meta-analysis methods:

- frequentist network meta-analysis (function netmeta) based on Rücker (2012) and Rücker & Schwarzer (2014);
- net heat plot (netheat) and design-based decomposition of Cochran's Q (decomp.design) described in Krahn et al. (2013);
- measures characterizing the flow of evidence between two treatments (netmeasures) described in König et al. (2013);
- ranking of treatments (netrank) based on P-scores (Rücker & Schwarzer, 2015) or SUCRAs (Salanti et al., 2011);
- rankograms (rankogram) (Salanti et al., 2011);
- partial order of treatment rankings (netposet, plot.netposet) and Hasse diagram (hasse) according to Carlsen & Bruggemann (2014) and Rücker & Schwarzer (2017);
- split direct and indirect evidence (netsplit) to check for consistency (Dias et al., 2010; Efthimiou et al., 2019);
- contribution of direct comparisons to network estimates (netcontrib) (Papakonstantinou et al., 2018; Davies et al., 2021);
- league table with network meta-analysis results (netleague);
- table with network, direct and indirect estimates from one or more network meta-analyses (nettable);
- additive network meta-analysis for combinations of treatments (netcomb, discomb for disconnected networks) (Rücker et al., 2020);
- calculate comparison effects of two arbitrary complex interventions in component network meta-analysis (netcomparison);
- calculate effect of arbitrary complex interventions in component network meta-analysis (netcomplex);
- network meta-analysis of binary data (netmetabin) using the Mantel-Haenszel or non-central hypergeometric distribution method (Efthimiou et al., 2019);
- 'comparison-adjusted' funnel plot (funnel.netmeta) to assess funnel plot asymmetry in network meta-analysis (Chaimani & Salanti, 2012);
- conduct pairwise meta-analyses for all comparisons with direct evidence in a network meta-analysis (netpairwise); netbind to show these results in a forest plot;

- automated drawing of network graphs (netgraph.netmeta) described in Rücker & Schwarzer (2016);
- results of several network meta-analyses can be combined with netbind to show these results in a forest plot.

Furthermore, functions and datasets from **netmeta** are utilised in Schwarzer et al. (2015), Chapter 8 "Network Meta-Analysis", https://link.springer.com/book/10.1007/978-3-319-21416-0.

Type help(package = "netmeta") for a listing of all R functions available in **netmeta**.

Type citation("netmeta") on how to cite **netmeta** in publications.

To report problems and bugs

- type bug.report(package = "netmeta") if you do not use RStudio,
- send an email to Guido Schwarzer <sc@imbi.uni-freiburg.de> if you use RStudio.

The development version of **netmeta** is available on GitHub https://github.com/guido-s/netmeta.

Author(s)

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References

Carlsen L, Bruggemann R (2014): Partial order methodology: a valuable tool in chemometrics. *Journal of Chemometrics*, **28**, 226–34

Chaimani A & Salanti G (2012): Using network meta-analysis to evaluate the existence of small-study effects in a network of interventions. *Research Synthesis Methods*, **3**, 161–76

Davies AL, Papakonstantinou T, Nikolakopoulou A, Rücker G, Galla T (2021): Network meta-analysis and random walks. Available from: http://arxiv.org/abs/2107.02886

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Efthimiou O, Rücker G, Schwarzer G, Higgins J, Egger M, Salanti G (2019): A Mantel-Haenszel model for network meta-analysis of rare events. *Statistics in Medicine*, 1–21, https://doi.org/10.1002/sim.8158

König J, Krahn U, Binder H (2013): Visualizing the flow of evidence in network meta-analysis and characterizing mixed treatment comparisons. *Statistics in Medicine*, **32**, 5414–29

Krahn U, Binder H, König J (2013): A graphical tool for locating inconsistency in network meta-analyses. *BMC Medical Research Methodology*, **13**, 35

Papakonstantinou, T., Nikolakopoulou, A., Rücker, G., Chaimani, A., Schwarzer, G., Egger, M., Salanti, G. (2018): Estimating the contribution of studies in network meta-analysis: paths, flows and streams. *F1000Research*

Rücker G (2012): Network meta-analysis, electrical networks and graph theory. *Research Synthesis Methods*, **3**, 312–24

Rücker G, Schwarzer G (2014): Reduce dimension or reduce weights? Comparing two approaches to multi-arm studies in network meta-analysis. *Statistics in Medicine*, **33**, 4353–69

Rücker G, Schwarzer G (2015): Ranking treatments in frequentist network meta-analysis works without resampling methods. *BMC Medical Research Methodology*, **15**, 58

Rücker G, Schwarzer G (2016): Automated drawing of network plots in network meta-analysis. *Research Synthesis Methods*, **7**, 94–107

Rücker G, Schwarzer G (2017): Resolve conflicting rankings of outcomes in network meta-analysis: Partial ordering of treatments. *Research Synthesis Methods*, **8**, 526–36

Rücker G, Petropoulou M, Schwarzer G (2020): Network meta-analysis of multicomponent interventions. *Biometrical Journal*, **62**, 808–21

Salanti G, Ades AE, Ioannidis JP (2011): Graphical methods and numerical summaries for presenting results from multiple-treatment meta-analysis: an overview and tutorial. *Journal of Clinical Epidemiology*, **64**, 163–71

Schwarzer G, Carpenter JR and Rücker G (2015): *Meta-Analysis with R (Use R!)*. Springer International Publishing, Switzerland.

`as.data.frame.netconnection` *Create a data frame from an object of class netconnection*

Description

The `as.data.frame` method returns a data frame containing information on membership of studies / pairwise comparisons to a (sub)network.

Usage

```
## S3 method for class 'netconnection'
as.data.frame(x, ...)
```

Arguments

`x`	An object of class `netconnection`.
`...`	Additional arguments (ignored).

Value

A data frame is returned by the function `as.data.frame`.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

`netconnection`

Examples

```
# Artificial example with two subnetworks
#
t1 <- c("G", "B", "B", "D", "A", "F")
t2 <- c("B", "C", "E", "E", "H", "A")
#
nc2 <- netconnection(t1, t2)
print(nc2, details = TRUE)

as.data.frame(nc2)
```

---

as.data.frame.netmeta *Create a data frame from an object of class netmeta*

Description

The `as.data.frame` method returns a data frame containing information on individual studies, e.g., estimated treatment effect and its standard error.

Usage

```
## S3 method for class 'netmeta'
as.data.frame(x, row.names = NULL, optional = FALSE, details = FALSE, ...)
```

Arguments

`x`	An object of class `netmeta`.
`row.names`	`NULL` or a character vector giving the row names for the data frame.
`optional`	A logical. If `TRUE`, setting row names and converting column names (to syntactic names) is optional.
`details`	A logical. If `TRUE`, additional variables of less interest are included in data frame.
`...`	Additional arguments.

Value

A data frame is returned by the function `as.data.frame`.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netmeta

Examples

```
data(Senn2013)

# Conduct network meta-analysis
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD")

as.data.frame(net1)
as.data.frame(net1, details = TRUE)
```

Baker2009 *Network meta-analysis of pharmacologic treatments for chronic obstructive pulmonary disease*

Description

This data set comes from a systematic review of randomized controlled trials on pharmacologic treatments for chronic obstructive pulmonary disease (COPD) (Baker et al., 2009).

The primary outcome, occurrence of one or more episodes of COPD exacerbation, is binary (yes / no). For this outcome, five drug treatments (fluticasone, budesonide, salmeterol, formoterol, tiotropium) and two combinations (fluticasone + salmeterol, budesonide + formoterol) were compared to placebo. The authors considered the two combinations as separate treatments instead of evaluating the individual components.

Format

A data frame with the following columns:

study	study label
year	year of publication
id	study ID
treatment	treatment
exac	one or more episodes of COPD exacerbation
total	number of individuals in treatment arm

Source

Baker WL, Baker EL, Coleman CI (2009): Pharmacologic Treatments for Chronic Obstructive Pulmonary Disease: A Mixed-Treatment Comparison Meta-analysis. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*, **29**, 891–905

See Also

pairwise, metabin, netmetabin

Examples

```
data(Baker2009)
Baker2009

## Not run:
# Transform data from long arm-based format to contrast-based
# format. Argument 'sm' has to be used for odds ratio as summary
# measure; by default the risk ratio is used in the metabin
# function called internally.
#
p1 <- pairwise(treatment, exac, total, studlab = paste(study, year),
  data = Baker2009, sm = "OR")

# Conduct network meta-analysis
#
net1 <- netmeta(p1, ref = "plac")

# Conduct component network meta-analysis
#
cnet1 <- netcomb(net1)
cnet1

## End(Not run)
```

`decomp.design` *Design-based decomposition of Cochran's Q in network meta-analysis*

Description

This function performs a design-based decomposition of Cochran's Q for assessing the homogeneity in the whole network, the homogeneity within designs, and the homogeneity/consistency between designs. It allows also an assessment of the consistency assumption after detaching the effect of single designs.

Usage

```
decomp.design(
  x,
  tau.preset = x$tau.preset,
  warn = TRUE,
  nchar.trts = x$nchar.trts
)
```

Arguments

`x`	An object of class `netmeta`.
`tau.preset`	An optional value for the square-root of the between-study variance τ^2 (see Details).

warn	A logical indicating whether warnings should be printed.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.

Details

In the context of network meta-analysis and the assessment of the homogeneity and consistency assumption, a generalized Cochran's Q statistic for multivariate meta-analysis can be used as shown in Krahn et al. (2013). This Q statistic can be decomposed in a sum of within-design Q statistics and one between-designs Q statistic that incorporates the concept of design inconsistency, see Higgins et al. (2012).

For assessing the inconsistency in a random effects model, the between-designs Q statistic can be calculated based on a full design-by-treatment interaction random effects model (see Higgins et al., 2012). This Q statistic will be automatically given in the output (τ^2 estimated by the method of moments (see Jackson et al., 2012). Alternatively, the square-root of the between-study variance can be prespecified by argument `tau.preset` to obtain a between-designs Q statistic (in `Q.inc.random`), its design-specific contributions `Q.inc.design.random.preset`) as well as residuals after detaching of single designs (`residuals.inc.detach.random.preset`).

Since an inconsistent treatment effect of one design can simultaneously inflate several residuals, Krahn et al. (2013) suggest for locating the inconsistency in a network to fit a set of extended models allowing for example for a deviating effect of each study design in turn. The recalculated between-designs Q statistics are given in list component `Q.inc.detach`. The change of the inconsistency contribution of single designs can be investigated in more detail by a net heat plot (see function netheat). Designs where only one treatment is involved in other designs of the network or where the removal of corresponding studies would lead to a splitting of the network do not contribute to the inconsistency assessment. These designs are not included in `Q.inc.detach`.

Value

Network meta-analysis with a single design: `NULL`. Otherwise, a list containing the following components:

Q.decomp	Data frame with Q statistics (variable `Q`) based on the common effects model to assess the homogeneity/consistency in the whole network, within designs, and between designs. Corresponding degrees of freedom (`df`) and p-values (`p.val`) are also given.
Q.het.design	Data frame with design-specific decomposition of the within-designs Q statistic (`Q`) of the common effects model, corresponding degrees of freedom (`df`) and p-values (`p.val`) are given.
Q.inc.detach	Data frame with between-designs Q statistics (`Q`) of the common effects model after detaching of single designs, corresponding degrees of freedom (`df`) and p-values (`p.val`) are given.
Q.inc.design	A named vector with contributions of single designs to the between design Q statistic given in `Q.decomp`.
Q.inc.random	Data frame with between-designs Q statistic (`Q`) based on a random effects model with square-root of between-study variance `tau.within` estimated embedded in a full design-by-treatment interaction model, corresponding degrees of freedom (`df`) and p-value (`p.val`).

`Q.inc.random.preset`
: Data frame with between-designs Q statistic (`Q`) based on a random effects model with prespecified square-root of between-study variance `tau.preset` in the case if argument `tau.preset` is not NULL, corresponding degrees of freedom (`df`) and p-value (`p.val`).

`Q.inc.design.random.preset`
: A named vector with contributions of single designs to the between design Q statistic based on a random effects model with prespecified square-root of between-study variance `tau.preset` in the case if argument `tau.preset` is given.

`residuals.inc.detach`
: Matrix with residuals, i.e. design-specific direct estimates minus the corresponding network estimates after detaching the design of the column.

`residuals.inc.detach.random.preset`
: Matrix with residuals analogous to `residuals.inc.detach` but based on a random effects model with prespecified square-root of between-study variance `tau.preset` in the case if argument `tau.preset` is not NULL.

`call`
: Function call.

`version`
: Version of R package netmeta used to create object.

Author(s)

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References

Higgins JPT, Jackson D, Barrett JK, Lu G, Ades AE, White IR (2012): Consistency and inconsistency in network meta-analysis: concepts and models for multi-arm studies. *Research Synthesis Methods*, **3**, 98–110

Krahn U, Binder H, König J (2013): A graphical tool for locating inconsistency in network meta-analyses. *BMC Medical Research Methodology*, **13**, 35

Jackson D, White IR and Riley RD (2012): Quantifying the impact of between-study heterogeneity in multivariate meta-analyses. *Statistics in Medicine*, **31**, 3805–20

See Also

netmeta, netheat

Examples

```
data(Senn2013)

# Only consider first five studies (to reduce runtime of example)
#
studies <- unique(Senn2013$studlab)
Senn2013.5 <- subset(Senn2013, studlab %in% studies[1:5])

# Conduct network meta-analysis with placebo as reference treatment
```

```
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013.5, sm = "MD", reference = "plac")

# Decomposition of Cochran's Q
#
decomp.design(net1)
```

`dietaryfat` *Network meta-analysis of dietary fat*

Description

Network meta-analysis comparing the effects of two diets to control on mortality.

The data are rates, given as the number of deaths and person-years. These data are used as an example in the supplemental material of Dias et al. (2013).

Format

A data frame with the following columns:

treat1	treatment 1
treat2	treatment 2
treat3	treatment 3
years1	person years arm 1
years2	person years arm 2
years3	person years arm 3
d1	events (deaths) arm 1
d2	events (deaths) arm 2
d3	events (deaths) arm 3
ID	study ID

Source

Dias S, Sutton AJ, Ades AE and Welton NJ (2013): Evidence synthesis for decision making 2: A generalized linear modeling framework for pairwise and network meta-analysis of randomized controlled trials. *Medical Decision Making*, **33**, 607–17

See Also

`pairwise`, `metainc`, `netmeta`, `netgraph.netmeta`

Examples

```
data(dietaryfat)

# Transform data from arm-based format to contrast-based format
```

```
# Using incidence rate ratios (sm = "IRR") as effect measure.
# Note, the argument 'sm' is not necessary as this is the default
# in R function metainc() called internally
#
p1 <- pairwise(list(treat1, treat2, treat3),
  list(d1, d2, d3), time = list(years1, years2, years3),
  studlab = ID, data = dietaryfat, sm = "IRR")
p1

# Conduct network meta-analysis
#
net1 <- netmeta(p1)
net1

# Conduct network meta-analysis using incidence rate differences
# (sm = "IRD")
#
p2 <- pairwise(list(treat1, treat2, treat3),
  list(d1, d2, d3), time = list(years1, years2, years3),
  studlab = ID, data = dietaryfat, sm = "IRD")
net2 <- netmeta(p2)
net2

# Draw network graph
#
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.25)

netgraph(net1, points = TRUE, cex.points = 3, cex = 1.25,
  labels = c("Control","Diet", "Diet 2"))
```

`discomb` *Additive network meta-analysis for combinations of treatments (disconnected networks)*

Description

Some treatments in a network meta-analysis may be combinations of other treatments or have common components. The influence of individual components can be evaluated in an additive network meta-analysis model assuming that the effect of treatment combinations is the sum of the effects of its components. This function implements this additive model in a frequentist way and is particularly intended for disconnected networks.

Usage

```
discomb(
  TE,
  seTE,
  treat1,
  treat2,
```

```
  studlab,
  data = NULL,
  subset = NULL,
  inactive = NULL,
  sep.comps = "+",
  C.matrix,
  sm,
  level = gs("level"),
  level.ma = gs("level.ma"),
  common = gs("common"),
  random = gs("random") | !is.null(tau.preset),
  reference.group,
  baseline.reference = TRUE,
  seq = NULL,
  tau.preset = NULL,
  tol.multiarm = 0.001,
  tol.multiarm.se = NULL,
  details.chkmultiarm = FALSE,
  details.chkident = FALSE,
  sep.trts = ":",
  nchar.comps = 666,
  func.inverse = invmat,
  backtransf = gs("backtransf"),
  title = "",
  warn = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  nchar.trts = nchar.comps,
  ...
)
```

Arguments

TE	Estimate of treatment effect, i.e. difference between first and second treatment (e.g. log odds ratio, mean difference, or log hazard ratio). Or an R object created with pairwise.
seTE	Standard error of treatment estimate.
treat1	Label/Number for first treatment.
treat2	Label/Number for second treatment.
studlab	An optional - but important! - vector with study labels (see netmeta).
data	An optional data frame containing the study information.
subset	An optional vector specifying a subset of studies to be used.
inactive	A character string defining the inactive treatment component (see Details).
sep.comps	A single character to define separator between treatment components.
C.matrix	C matrix (see Details).
sm	A character string indicating underlying summary measure, e.g., "RD", "RR", "OR", "ASD", "HR", "MD", "SMD", or "ROM".

`level` The level used to calculate confidence intervals for individual comparisons.

`level.ma` The level used to calculate confidence intervals for network estimates.

`common` A logical indicating whether a common effects / common effects network meta-analysis should be conducted.

`random` A logical indicating whether a random effects network meta-analysis should be conducted.

`reference.group`
Reference treatment (first treatment is used if argument is missing).

`baseline.reference`
A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa. This argument is only considered if `reference.group` has been specified.

`seq` A character or numerical vector specifying the sequence of treatments in printouts.

`tau.preset` An optional value for the square-root of the between-study variance τ^2.

`tol.multiarm` A numeric for the tolerance for consistency of treatment estimates in multi-arm studies which are consistent by design.

`tol.multiarm.se`
A numeric for the tolerance for consistency of standard errors in multi-arm studies which are consistent by design. This check is not conducted if the argument is `NULL`.

`details.chkmultiarm`
A logical indicating whether treatment estimates and / or variances of multi-arm studies with inconsistent results or negative multi-arm variances should be printed.

`details.chkident`
A logical indicating whether details on unidentifiable components should be printed.

`sep.trts` A character used in comparison names as separator between treatment labels.

`nchar.comps` A numeric defining the minimum number of characters used to create unique names for components (see Details).

`func.inverse` R function used to calculate the pseudoinverse of the Laplacian matrix L (see `netmeta`).

`backtransf` A logical indicating whether results should be back transformed in printouts and forest plots. If `backtransf = TRUE`, results for `sm = "OR"` are presented as odds ratios rather than log odds ratios, for example.

`title` Title of meta-analysis / systematic review.

`warn` A logical indicating whether warnings should be printed (e.g., if studies are excluded from meta-analysis due to zero standard errors).

`warn.deprecated`
A logical indicating whether warnings should be printed if deprecated arguments are used.

`nchar.trts` Deprecated argument (replaced by `nchar.comps`).

`...` Additional arguments (to catch deprecated arguments).

Details

Treatments in network meta-analysis (NMA) can be complex interventions. Some treatments may be combinations of others or have common components. The standard analysis provided by `netmeta` is a NMA where all existing (single or combined) treatments are considered as different nodes in the network. Exploiting the fact that some treatments are combinations of common components, an additive component network meta-analysis (CNMA) model can be used to evaluate the influence of individual components. This model assumes that the effect of a treatment combination is the sum of the effects of its components which implies that common components cancel out in comparisons.

This R function can be used for disconnected networks. Use `netmeta` and `netcomb` for connected networks.

The additive CNMA model has been implemented using Bayesian methods (Mills et al., 2012; Welton et al., 2013). This function implements the additive model in a frequentist way (Rücker et al., 2020).

The underlying multivariate model is given by

$$\boldsymbol{\delta} = \boldsymbol{B\theta}, \boldsymbol{\theta} = \boldsymbol{C\beta}$$

with

$\boldsymbol{\delta}$ vector of true treatment effects (differences) from individual studies,

$\boldsymbol{B}$ design matrix describing the structure of the network,

$\boldsymbol{\theta}$ parameter vector that represents the existing combined treatments,

$\boldsymbol{C}$ matrix describing how the treatments are composed,

$\boldsymbol{\beta}$ parameter vector representing the treatment components.

All parameters are estimated using weighted least squares regression.

Argument `inactive` can be used to specify a single component that does not have any therapeutic value. Accordingly, it is assumed that the treatment effect of the combination of this component with an additional treatment component is equal to the treatment effect of the additional component alone.

Argument `sep.comps` can be used to specify the separator between individual components. By default, the matrix **C** is calculated internally from treatment names. However, it is possible to specify a different matrix using argument `C.matrix`.

By default, component names are not abbreviated in printouts. However, in order to get more concise printouts, argument `nchar.comps` can be used to define the minimum number of characters for abbreviated component names (see `abbreviate`, argument `minlength`). R function `treats` is utilised internally to create abbreviated component names.

Value

An object of classes `discomb` and `netcomb` with corresponding `print`, `summary`, and `forest` functions. The object is a list containing the following components:

`studlab`	Study labels.
`treat1`	Label/Number for first treatment.

`treat2` Label/Number for second treatment.

`TE` Estimate of treatment effect, i.e. difference between first and second treatment.

`seTE` Standard error of treatment estimate.

`seTE.adj.common`, `seTE.adj.random`
Standard error of treatment estimate, adjusted for multi-arm studies.

`event1` Number of events in first treatment group.

`event2` Number of events in second treatment group.

`n1` Number of observations in first treatment group.

`n2` Number of observations in second treatment group.

`k` Total number of studies.

`m` Total number of pairwise comparisons.

`n` Total number of treatments.

`d` Total number of designs (corresponding to the unique set of treatments compared within studies).

`c` Total number of components.

`trts` Treatments included in network meta-analysis.

`comps` Unique list of components present in the network.

`TE.cnma.common`, `TE.cnma.random`
A vector of length *m* of consistent treatment effects estimated by the additive (common and random effects) model.

`seTE.cnma.common`, `seTE.cnma.random`
A vector of length *m* with standard errors estimated by the additive (common and random effects) model.

`lower.cnma.common`, `lower.cnma.random`
A vector of length *m* of lower confidence interval limits for consistent treatment effects estimated by the additive (common and random effects) model.

`upper.cnma.common`, `upper.cnma.random`
A vector of length *m* of upper confidence interval limits for consistent treatment effects estimated by the additive (common and random effects) model.

`statistic.cnma.common`, `statistic.cnma.random`
A vector of length *m* of z-values for the test of an overall effect estimated by the additive (common and random effects) model.

`pval.cnma.common`, `pval.cnma.random`
A vector of length *m* of p-values for the test of an overall effect estimated by the additive (common and random effects) model.

`TE.common`, `TE.random`
*n*x*n* matrix with overall treatment effects estimated by the additive (common and random effects) model.

`seTE.common`, `seTE.random`
*n*x*n* matrix with standard errors estimated by the additive (common and random effects) model.

`lower.common`, `upper.common`, `lower.random`, `upper.random`
*n*x*n* matrices with lower and upper confidence interval limits estimated by the additive (common and random effects) model.

`statistic.common`, `pval.common`, `statistic.random`, `pval.random`
: *n*x*n* matrices with z-values and p-values for test of overall effect estimated by the additive (common and random effects) model.

`Comp.common`, `Comp.random`
: A vector of component effects (common and random effects model).

`seComp.common`, `seComp.random`
: A vector with corresponding standard errors (common and random effects model).

`lower.Comp.common`, `lower.Comp.random`
: A vector with lower confidence limits for components (common and random effects model).

`upper.Comp.common`, `upper.Comp.random`
: A vector with upper confidence limits for components (common and random effects model).

`statistic.Comp.common`, `statistic.Comp.random`
: A vector with z-values for the overall effect of components (common and random effects model).

`pval.Comp.common`, `pval.Comp.random`
: A vector with p-values for the overall effect of components (common and random effects model).

`Comb.common`, `Comb.random`
: A vector of combination effects (common and random effects model).

`seComb.common`, `seComb.random`
: A vector with corresponding standard errors (common and random effects model).

`lower.Comb.common`, `lower.Comb.random`
: A vector with lower confidence limits for combinations (common and random effects model).

`upper.Comb.common`, `upper.Comb.random`
: A vector with upper confidence limits for combinations (common and random effects model).

`statistic.Comb.common`, `statistic.Comb.random`
: A vector with z-values for the overall effect of combinations (common and random effects model).

`pval.Comb.common`, `pval.Comb.random`
: A vector with p-values for the overall effect of combinations (common and random effects model).

`Q.additive`
: Overall heterogeneity / inconsistency statistic (additive model).

`df.Q.additive`
: Degrees of freedom for test of heterogeneity / inconsistency (additive model).

`pval.Q.additive`
: P-value for test of heterogeneity / inconsistency (additive model).

`tau`
: Square-root of between-study variance (additive model).

`I2`
: I-squared (additive model).

`Q.standard`
: Overall heterogeneity / inconsistency statistic (standard model).

`df.Q.standard`
: Degrees of freedom for test of heterogeneity / inconsistency (standard model).

`pval.Q.standard`
: P-value for test of heterogeneity / inconsistency (standard model).

Q.diff	Test statistic for difference in goodness of fit between standard and additive model.
df.Q.diff	Degrees of freedom for difference in goodness of fit between standard and additive model.
pval.Q.diff	P-value for difference in goodness of fit between standard and additive model.
X.matrix	Design matrix (*m*x*n*).
B.matrix	Edge-vertex incidence matrix (*m*x*n*).
C.matrix	As defined above.
sm	Summary measure.
level.ma	Level for confidence intervals.
common, random, tau.preset	As defined above.
sep.trts	A character used in comparison names as separator between treatment labels.
nchar.comps	A numeric defining the minimum number of characters used to create unique component names.
inactive, sep.comps	As defined above.
backtransf	A logical indicating whether results should be back transformed in printouts and forest plots.
title	Title of meta-analysis / systematic review.
x	As defined above.
call	Function call.
version	Version of R package netmeta used to create object.

Note

This function calculates effects for individual components and complex interventions present in the network.

R function netcomplex can be used to calculate the effect for arbitrary complex interventions in a component network meta-analysis. Furthermore, R function netcomparison can be used to calculate the effect for comparisons of two arbitrary complex intervention in a component network meta-analysis.

Author(s)

Gerta Rücker <ruecker@imbi.uni-freiburg.de>, Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

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See Also

netcomb, forest.netcomb, summary.netcomb, netmeta, netconnection, netcomplex, netcomparison

Examples

```
# Artificial dataset
#
t1 <- c("A + B", "A + C", "A"    , "A"    , "D", "D", "E")
t2 <- c("C"    , "B"    , "B + C", "A + D", "E", "F", "F")
#
mean    <- c(4.1, 2.05, 0, 0, 0.1, 0.1, 0.05)
se.mean <- rep(0.1, 7)
#
study <- paste("study", c(1:4, 5, 5, 5))
#
dat <- data.frame(mean, se.mean, t1, t2, study,
  stringsAsFactors = FALSE)
#
trts <- c("A", "A + B", "A + C", "A + D",
  "B", "B + C", "C", "D", "E", "F")
#
comps <- LETTERS[1:6]

# Use netconnection() to display network information
#
netconnection(t1, t2, study)

dc1 <- discomb(mean, se.mean, t1, t2, study, seq = trts)
dc1

forest(dc1, ref = "F")

# Define C matrix manually (which will produce the same results)
#
C <- rbind(c(1, 0, 0, 0, 0, 0),  # A
  c(1, 1, 0, 0, 0, 0),  # A + B
  c(1, 0, 1, 0, 0, 0),  # A + C
  c(1, 0, 0, 1, 0, 0),  # A + D
  c(0, 1, 0, 0, 0, 0),  # B
  c(0, 1, 1, 0, 0, 0),  # B + C
  c(0, 0, 1, 0, 0, 0),  # C
  c(0, 0, 0, 1, 0, 0),  # D
  c(0, 0, 0, 0, 1, 0),  # E
  c(0, 0, 0, 0, 0, 1))  # F
#
```

```
colnames(C) <- comps
rownames(C) <- trts
#
dc2 <- discomb(mean, se.mean, t1, t2, study, seq = trts,
  C.matrix = C)
#
# Compare C matrices
#
all.equal(dc1$C.matrix, dc2$C.matrix)
```

Dogliotti2014 *Network meta-analysis of antithrombotic treatments in patients with non-valvular atrial fibrillation*

Description

This data set comes from a systematic review aiming to determine the effects of eight antithrombotic treatments in reducing the incidence of major thrombotic events in patients with non-valvular atrial fibrillation (Dogliotti et al., 2014). The review included 20 studies (79,808 participants), four of which were three-arm studies. The primary outcome is stroke reduction.

Format

A data frame with the following columns:

study	study label
id	study ID
treatment	treatment
stroke	number of strokes
total	number of individuals in treatment arm

Source

Dogliotti A, Paolasso E, Giugliano RP (2014): Current and new oral antithrombotics in non-valvular atrial fibrillation: a network meta-analysis of 79 808 patients. *Heart*, **100**, 396–405

See Also

pairwise, metabin, netmetabin

Examples

```
data(Dogliotti2014)
Dogliotti2014

## Not run:
# Transform data from long arm-based format to contrast-based
# format. Argument 'sm' has to be used for odds ratio as summary
```

```
# measure; by default the risk ratio is used in the metabin
# function called internally.
#
p1 <- pairwise(treatment, stroke, total, studlab = study,
  data = Dogliotti2014, sm = "OR")

# Conduct Mantel-Haenszel network meta-analysis
#
netmetabin(p1, ref = "plac")

## End(Not run)
```

Dong2013 *Network meta-analysis for chronic obstructive pulmonary disease*

Description

Network meta-analysis comparing inhaled medications in patients with chronic obstructive pulmonary disease.

Format

A data frame with the following columns:

id	study ID
treatment	treatment
death	mortality
randomized	number of individuals in treatment arm

Source

Dong Y-H, Lin H-H, Shau W-Y, Wu Y-C, Chang C-H, Lai M-S (2013): Comparative safety of inhaled medications in patients with chronic obstructive pulmonary disease: systematic review and mixed treatment comparison meta-analysis of randomised controlled trials. *Thorax*, **68**, 48–56

See Also

pairwise, metabin, netmetabin

Examples

```
data(Dong2013)

# Only consider first ten studies (to reduce runtime of example)
#
first10 <- subset(Dong2013, id <= 10)

# Transform data from long arm-based format to contrast-based
# format. Argument 'sm' has to be used for odds ratio as summary
```

```
# measure; by default the risk ratio is used in the metabin
# function called internally.
#
p1 <- pairwise(treatment, death, randomized, studlab = id,
  data = first10, sm = "OR")

# Conduct Mantel-Haenszel network meta-analysis
#
netmetabin(p1, ref = "plac")

## Not run:
# Conduct Mantel-Haenszel network meta-analysis for the whole
# dataset
#
p2 <- pairwise(treatment, death, randomized, studlab = id,
  data = Dong2013, sm = "OR")
netmetabin(p2, ref = "plac")

## End(Not run)
```

`forest.netbind` *Forest plot showing results of two or more network meta-analyses*

Description

Forest plot to show network estimates of two or more network meta-analyses.

Usage

```
## S3 method for class 'netbind'
forest(
  x,
  pooled = ifelse(x$x$random, "random", "common"),
  equal.size = TRUE,
  leftcols = "studlab",
  leftlabs = "Treatment",
  rightcols = c("effect", "ci"),
  rightlabs = NULL,
  subset.treatments,
  digits = gs("digits.forest"),
  digits.prop = max(gs("digits.pval") - 2, 2),
  backtransf = x$backtransf,
  lab.NA = "",
  smlab,
  ...
)

## S3 method for class 'netbind'
plot(x, ...)
```

Arguments

x	An object of class netbind.
pooled	A character string indicating whether results for the common ("common") or random effects model ("random") should be plotted. Can be abbreviated.
equal.size	A logical indicating whether all squares should be of equal size. Otherwise, the square size is proportional to the precision of estimates.
leftcols	A character vector specifying columns to be plotted on the left side of the forest plot (see Details).
leftlabs	A character vector specifying labels for columns on left side of the forest plot.
rightcols	A character vector specifying columns to be plotted on the right side of the forest plot (see Details).
rightlabs	A character vector specifying labels for columns on right side of the forest plot.
subset.treatments	A character vector specifying treatments to show in forest plot as comparators to the reference.
digits	Minimal number of significant digits for treatment effects and confidence intervals, see print.default.
digits.prop	Minimal number of significant digits for the direct evidence proportion.
backtransf	A logical indicating whether results should be back transformed in forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
lab.NA	A character string to label missing values.
smlab	A label printed at top of figure. By default, text indicating either common or random effects model is printed.
...	Additional arguments for forest.meta function.

Details

A forest plot, also called confidence interval plot, is drawn in the active graphics window.

The arguments leftcols and rightcols can be used to specify columns which are plotted on the left and right side of the forest plot, respectively. If argument rightcols is FALSE, no columns will be plotted on the right side.

For more information see help page of forest.meta function.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netbind, netcomb, forest.meta

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Standard random effects NMA model (with placebo as reference
# treatment)
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, reference.group = "placebo",
  sm = "OR", common = FALSE)

# Additive CNMA model with placebo as inactive component and
# reference
#
nc1 <- netcomb(net1, inactive = "placebo")

# Combine results of standard NMA and CNMA
#
nb1 <- netbind(nc1, net1,
  name = c("Additive CNMA", "Standard NMA"),
  col.study = c("red", "black"),
  col.square = c("red", "black"))
forest(nb1,
  col.by = "black", addrow.subgroups = FALSE,
  fontsize = 10, spacing = 0.7, squaresize = 0.9,
  label.left = "Favours Placebo",
  label.right = "Favours other")
```

`forest.netcomb` *Forest plot for additive network meta-analysis*

Description

Draws a forest plot in the active graphics window (using grid graphics system).

Usage

```
## S3 method for class 'netcomb'
forest(
  x,
  pooled = ifelse(x$random, "random", "common"),
  reference.group = x$reference.group,
  baseline.reference = x$baseline.reference,
  leftcols = "studlab",
```

```
  leftlabs = "Treatment",
  rightcols = c("effect", "ci"),
  rightlabs = NULL,
  digits = gs("digits.forest"),
  smlab = NULL,
  sortvar = x$seq,
  backtransf = x$backtransf,
  lab.NA = ".",
  add.data,
  drop.reference.group = FALSE,
  weight.study = "same",
  ...
)

## S3 method for class 'netcomb'
plot(x, ...)
```

Arguments

x	An object of class netcomb.
pooled	A character string indicating whether results for the common ("common") or random effects model ("random") should be plotted. Can be abbreviated.
reference.group	Reference treatment(s).
baseline.reference	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa.
leftcols	A character vector specifying (additional) columns to be plotted on the left side of the forest plot or a logical value (see forest.meta help page for details).
leftlabs	A character vector specifying labels for (additional) columns on left side of the forest plot (see forest.meta help page for details).
rightcols	A character vector specifying (additional) columns to be plotted on the right side of the forest plot or a logical value (see forest.meta help page for details).
rightlabs	A character vector specifying labels for (additional) columns on right side of the forest plot (see forest.meta help page for details).
digits	Minimal number of significant digits for treatment effects and confidence intervals, see print.default.
smlab	A label printed at top of figure. By default, text indicating either common or random effects model is printed.
sortvar	An optional vector used to sort the individual studies (must be of same length as the total number of treatments).
backtransf	A logical indicating whether results should be back transformed in forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
lab.NA	A character string to label missing values.

add.data	An optional data frame with additional columns to print in forest plot (see Details).
drop.reference.group	A logical indicating whether the reference group should be printed in the forest plot.
weight.study	A character string indicating weighting used to determine size of squares or diamonds.
...	Additional arguments for forest.meta function.

Details

A forest plot, also called confidence interval plot, is drawn in the active graphics window.

Argument sortvar can be either a numeric or character vector with length of number of treatments. If sortvar is numeric the order function is utilised internally to determine the order of values. If sortvar is character it must be a permutation of the treatment names. It is also possible to to sort by treatment comparisons (sortvar = TE, etc.), standard error (sortvar = seTE), and number of studies with direct treatment comparisons (sortvar = k).

Argument add.data can be used to add additional columns to the forest plot. This argument must be a data frame with the same row names as the treatment effects matrices in R object x, i.e., x$TE.common or x$TE.random.

For more information see help page of forest.meta function.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netcomb, discomb, forest.meta

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, ref = "placebo", sm = "OR", common = FALSE)

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")
#
```

```
forest(nc1)

## Not run:
# Specify, order of treatments
#
trts <- c("TCA", "SSRI", "SNRI", "NRI", "Low-dose SARI", "NaSSa",
  "rMAO-A", "Ind drug", "Hypericum", "Face-to-face CBT",
  "Face-to-face PST", "Face-to-face interpsy", "Face-to-face psychodyn",
  "Other face-to-face", "Remote CBT", "Self-help CBT", "No contact CBT",
  "Face-to-face CBT + SSRI", "Face-to-face interpsy + SSRI",
  "Face-to-face PST + SSRI", "UC", "Placebo")
#
# Note, three treatments are actually combinations of 'SSRI' with
# other components:
# "Face-to-face CBT + SSRI",
# "Face-to-face interpsy + SSRI",
# "Face-to-face PST + SSRI"

# Conduct random effects network meta-analysis
#
net2 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = Linde2016, ref = "placebo",
  seq = trts, sm = "OR", common = FALSE)
#
net2

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc2 <- netcomb(net2, inactive = "placebo")
#
forest(nc2)

## End(Not run)
```

`forest.netcomparison` *Forest plot for complex interventions in component network meta-analysis*

Description

Draws a forest plot in the active graphics window (using grid graphics system).

Usage

```
## S3 method for class 'netcomparison'
forest(
  x,
  pooled = ifelse(x$random, "random", "common"),
```

```
  leftcols = c("studlab", "treat1", "treat2"),
  leftlabs = c("Comparison", "Trt 1", "Trt 2"),
  rightcols = c("effect", "ci", "statistic", "pval"),
  rightlabs = c(NA, NA, "z", "p-value"),
  nchar.comps = x$nchar.trts,
  digits = gs("digits.forest"),
  digits.stat = gs("digits.stat"),
  digits.pval = gs("digits.pval"),
  smlab = NULL,
  backtransf = x$backtransf,
  lab.NA = ".",
  weight.study = "same",
  ...
)

## S3 method for class 'netcomparison'
plot(x, ...)
```

Arguments

x	An object of class netcomparison.
pooled	A character string indicating whether results for the common ("common") or random effects model ("random") should be plotted. Can be abbreviated.
leftcols	A character vector specifying (additional) columns to be plotted on the left side of the forest plot or a logical value (see forest.meta help page for details).
leftlabs	A character vector specifying labels for (additional) columns on left side of the forest plot (see forest.meta help page for details).
rightcols	A character vector specifying (additional) columns to be plotted on the right side of the forest plot or a logical value (see forest.meta help page for details).
rightlabs	A character vector specifying labels for (additional) columns on right side of the forest plot (see forest.meta help page for details).
nchar.comps	A numeric defining the minimum number of characters used to create unique names for components.
digits	Minimal number of significant digits for treatment effects and confidence intervals, see print.default.
digits.stat	Minimal number of significant digits for tests of overall effect, see print.default.
digits.pval	Minimal number of significant digits for p-value of overall effects, see print.default.
smlab	A label printed at top of figure. By default, text indicating either common or random effects model is printed.
backtransf	A logical indicating whether results should be back transformed in forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
lab.NA	A character string to label missing values.
weight.study	A character string indicating weighting used to determine size of squares or diamonds.
...	Additional arguments for forest.meta function.

Details

A forest plot, also called confidence interval plot, is drawn in the active graphics window. For more information see help page of forest.meta function.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netcomparison, netcomb, discomb, forest.meta

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, ref = "placebo", sm = "OR", common = FALSE)

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")

# Some comparisons
#
t1 <- c("F + TCA", "F + Plac", "SSRI + Plac + TCA")
t2 <- c("UC", "Plac", "UC")
#
netcomparison(nc1, t1, t2)
#
forest(netcomparison(nc1, t1, t2))
forest(netcomparison(nc1, t1, t2), nchar.comps = 4)
forest(netcomparison(nc1, c("F", "TCA"), "UC"), nchar.comps = 4)
```

---

forest.netcomplex *Forest plot for complex interventions in component network meta-analysis*

Description

Draws a forest plot in the active graphics window (using grid graphics system).

Usage

```
## S3 method for class 'netcomplex'
forest(
  x,
  pooled = ifelse(x$random, "random", "common"),
  leftcols = "studlab",
  leftlabs = NULL,
  rightcols = c("effect", "ci", "statistic", "pval"),
  rightlabs = c(NA, NA, "z", "p-value"),
  nchar.comps = x$nchar.trts,
  digits = gs("digits.forest"),
  digits.stat = gs("digits.stat"),
  digits.pval = gs("digits.pval"),
  smlab = NULL,
  backtransf = x$backtransf,
  lab.NA = ".",
  weight.study = "same",
  ...
)

## S3 method for class 'netcomplex'
plot(x, ...)
```

Arguments

x	An object of class netcomplex.
pooled	A character string indicating whether results for the common ("common") or random effects model ("random") should be plotted. Can be abbreviated.
leftcols	A character vector specifying (additional) columns to be plotted on the left side of the forest plot or a logical value (see forest.meta help page for details).
leftlabs	A character vector specifying labels for (additional) columns on left side of the forest plot (see forest.meta help page for details).
rightcols	A character vector specifying (additional) columns to be plotted on the right side of the forest plot or a logical value (see forest.meta help page for details).
rightlabs	A character vector specifying labels for (additional) columns on right side of the forest plot (see forest.meta help page for details).
nchar.comps	A numeric defining the minimum number of characters used to create unique names for components.
digits	Minimal number of significant digits for treatment effects and confidence intervals, see print.default.
digits.stat	Minimal number of significant digits for tests of overall effect, see print.default.
digits.pval	Minimal number of significant digits for p-value of overall effects, see print.default.
smlab	A label printed at top of figure. By default, text indicating either common or random effects model is printed.

backtransf	A logical indicating whether results should be back transformed in forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
lab.NA	A character string to label missing values.
weight.study	A character string indicating weighting used to determine size of squares or diamonds.
...	Additional arguments for forest.meta function.

Details

A forest plot, also called confidence interval plot, is drawn in the active graphics window. For more information see help page of forest.meta function.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netcomplex, netcomb, discomb, forest.meta

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, ref = "placebo", sm = "OR", common = FALSE)

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")

# Some complex interventions
#
ints <- c("F + TCA", "F + Plac", "SSRI + Plac + TCA")
netcomplex(nc1, ints)
#
forest(netcomplex(nc1, ints))
forest(netcomplex(nc1, ints), nchar.comps = 4)

# Component effects
#
forest(netcomplex(nc1, nc1$comps))
```

`forest.netmeta` *Forest plot for network meta-analysis*

Description

Draws a forest plot in the active graphics window (using grid graphics system).

Usage

```
## S3 method for class 'netmeta'
forest(
  x,
  pooled = ifelse(x$random, "random", "common"),
  reference.group = x$reference.group,
  baseline.reference = x$baseline.reference,
  labels = x$trts,
  equal.size = TRUE,
  leftcols = "studlab",
  leftlabs,
  rightcols = c("effect", "ci"),
  rightlabs,
  digits = gs("digits.forest"),
  small.values = x$small.values,
  nsim = 1000,
  digits.prop = 2,
  smlab = NULL,
  sortvar = x$seq,
  backtransf = x$backtransf,
  lab.NA = ".",
  add.data,
  drop.reference.group = FALSE,
  col.by = "black",
  print.subgroup.name = FALSE,
  ...
)

## S3 method for class 'netmeta'
plot(x, ...)
```

Arguments

`x`	An object of class `netmeta`.
`pooled`	A character string indicating whether results for the common (`"common"`) or random effects model (`"random"`) should be plotted. Can be abbreviated.

`reference.group`
: Reference treatment(s).

`baseline.reference`
: A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa.

`labels`
: An optional vector with treatment labels.

`equal.size`
: A logical indicating whether all squares should be of equal size. Otherwise, the square size is proportional to the precision of estimates.

`leftcols`
: A character vector specifying columns to be plotted on the left side of the forest plot or a logical value (see Details).

`leftlabs`
: A character vector specifying labels for (additional) columns on left side of the forest plot (see Details).

`rightcols`
: A character vector specifying columns to be plotted on the right side of the forest plot or a logical value (see Details).

`rightlabs`
: A character vector specifying labels for (additional) columns on right side of the forest plot (see Details).

`digits`
: Minimal number of significant digits for treatment effects and confidence intervals, see `print.default`.

`small.values`
: A character string specifying whether small treatment effects indicate a beneficial (`"good"`) or harmful (`"bad"`) effect, can be abbreviated; see `netrank`.

`nsim`
: Number of simulations to calculate SUCRAs.

`digits.prop`
: Minimal number of significant digits for P-scores, SUCRAs and direct evidence proportions, see `print.default` and `netrank`.

`smlab`
: A label printed at top of figure. By default, text indicating either common or random effects model is printed.

`sortvar`
: An optional vector used to sort treatments (must be of same length as the total number of treatments).

`backtransf`
: A logical indicating whether results should be back transformed in forest plots. If `backtransf = TRUE`, results for `sm = "OR"` are presented as odds ratios rather than log odds ratios, for example.

`lab.NA`
: A character string to label missing values.

`add.data`
: An optional data frame with additional columns to print in forest plot (see Details).

`drop.reference.group`
: A logical indicating whether the reference group should be printed in the forest plot.

`col.by`
: The colour to print information on subgroups.

`print.subgroup.name`
: A logical indicating whether the name of the grouping variable should be printed in front of the group labels.

`...`
: Additional arguments for `forest.meta` function.

Details

A forest plot, also called confidence interval plot, is drawn in the active graphics window.

Argument `sortvar` can be either a numeric or character vector with length of number of treatments. If `sortvar` is numeric the `order` function is utilised internally to determine the order of values. If `sortvar` is character it must be a permutation of the treatment names. It is also possible to provide either `sortvar = Pscore`, `sortvar = "Pscore"`, `sortvar = -Pscore`, or `sortvar = "-Pscore"` in order to sort treatments according to the ranking generated by `netrank` which is called internally. It is also possible to use "SUCRA" instead of "Pscore". Similar expressions are possible to sort by treatment comparisons (`sortvar = TE`, etc.), standard error (`sortvar = seTE`), number of studies with direct treatment comparisons (`sortvar = k`), and direct evidence proportion (`sortvar = prop.direct`, see also `netmeasures`).

The arguments `leftcols` and `rightcols` can be used to specify columns which are plotted on the left and right side of the forest plot, respectively. The following columns are available:

Name	Definition
`"studlab"`	Treatments
`"TE"`	Network estimates (either from common or random effects model)
`"seTE"`	Corresponding standard errors
`"Pscore"`	P-scores (see `netrank`)
`"SUCRA"`	SUCRAs (see `netrank`)
`"n.trts"`	Number of participants per treatment arm
`"k"`	Number of studies in pairwise comparisons
`"prop.direct"`	Direct evidence proportions (see `netmeasures`)
`"effect"`	(Back-transformed) network estimates
`"ci"`	Confidence intervals
`"effect.ci"`	(Back-transformed) network estimates and confidence intervals

As a sidenote, the rather odd column name `"studlab"` to describe the treatment comparisons comes from internally calling `forest.meta` which uses study labels as the essential information.

Argument `add.data` can be used to add additional columns to the forest plot. This argument must be a data frame with row names equal to the treatment names in R object `x`, i.e., `x$trts`.

See help page of `forest.meta` for more information on the generation of forest plots and additional arguments.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

`forest.meta`

Examples

```
data(Senn2013)

## Not run:
```

```
# Conduct network meta-analysis
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD")

forest(net1, ref = "plac")
forest(net1, xlim = c(-1.5, 1), ref = "plac",
  xlab = "HbA1c difference", rightcols = FALSE)

## End(Not run)

# Random effects effect model
#
net2 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", common = FALSE)

forest(net2, xlim = c(-1.5, 1), ref = "plac",
  xlab = "HbA1c difference")

## Not run:
# Add column with P-Scores on right side of forest plot
#
forest(net2, xlim = c(-1.5, 1), ref = "plac",
  xlab = "HbA1c difference",
  rightcols = c("effect", "ci", "Pscore"),
  just.addcols = "right")

# Add column with P-Scores on left side of forest plot
#
forest(net2, xlim = c(-1.5, 1), ref = "plac",
  xlab = "HbA1c difference",
  leftcols = c("studlab", "Pscore"),
  just.addcols = "right")

# Sort forest plot by descending P-Score
#
forest(net2, xlim = c(-1.5, 1), ref = "plac",
  xlab = "HbA1c difference",
  rightcols = c("effect", "ci", "Pscore"),
  just.addcols = "right",
  sortvar = -Pscore)

# Drop reference group and sort by and print number of studies with
# direct treatment comparisons
#
forest(net2, xlim = c(-1.5, 1), ref = "plac",
  xlab = "HbA1c difference",
  leftcols = c("studlab", "k"),
  leftlabs = c("Contrast\nto Placebo", "Direct\nComparisons"),
  sortvar = -k,
  drop = TRUE,
  smlab = "Random Effects Model")
```

```
## End(Not run)
```

`forest.netsplit` *Forest plot for direct and indirect evidence*

Description

Forest plot to show direct and indirect evidence in network meta-analysis. Furthermore, estimates from network meta-analysis as well as prediction intervals can be printed.

Usage

```
## S3 method for class 'netsplit'
forest(
  x,
  pooled = ifelse(x$x$random, "random", "common"),
  show = "both",
  subgroup = "comparison",
  overall = TRUE,
  direct = TRUE,
  indirect = TRUE,
  prediction = x$prediction,
  only.reference = FALSE,
  sortvar = NULL,
  text.overall = "Network estimate",
  text.direct = "Direct estimate",
  text.indirect = "Indirect estimate",
  text.predict = "Prediction interval",
  type.overall,
  type.direct,
  type.indirect,
  col.square = "gray",
  col.square.lines = col.square,
  col.inside = "white",
  col.diamond = "gray",
  col.diamond.lines = "black",
  col.predict = "red",
  col.predict.lines = "black",
  equal.size = TRUE,
  leftcols,
  leftlabs,
  rightcols = c("effect", "ci"),
  rightlabs = NULL,
  digits = gs("digits.forest"),
  digits.prop = max(gs("digits.pval") - 2, 2),
  backtransf = x$backtransf,
```

```
  lab.NA = "",
  smlab,
  ...
)

## S3 method for class 'netsplit'
plot(x, ...)
```

Arguments

x	An object of class netsplit.
pooled	A character string indicating whether results for the common ("common") or random effects model ("random") should be plotted. Can be abbreviated.
show	A character string indicating which comparisons should be printed (see Details).
subgroup	A character string indicating which layout should be used in forest plot: subgroups by comparisons ("comparison") or subgroups by estimates ("estimate"). Can be abbreviated.
overall	A logical indicating whether network meta-analysis estimates should be printed.
direct	A logical indicating whether direct estimates should be printed.
indirect	A logical indicating whether indirect estimates should be printed.
prediction	A logical indicating whether prediction intervals should be printed.
only.reference	A logical indicating whether only comparisons with the reference group should be printed.
sortvar	An optional vector used to sort comparisons (must be of same length as the total number of comparisons).
text.overall	A character string used in the plot to label the network estimates.
text.direct	A character string used in the plot to label the direct estimates.
text.indirect	A character string used in the plot to label the indirect estimates.
text.predict	A character string used in the plot to label the prediction interval.
type.overall	A character string specifying how to plot treatment effects and confidence intervals for the overall network evidence.
type.direct	A character string specifying how to plot treatment effects and confidence intervals for the direct evidence.
type.indirect	A character string specifying how to plot treatment effects and confidence intervals for the indirect evidence.
col.square	The colour for squares.
col.square.lines	The colour for the outer lines of squares.
col.inside	The colour for results and confidence limits if confidence limits are completely within squares squares.
col.diamond	The colour of diamonds.
col.diamond.lines	The colour of the outer lines of diamonds.

col.predict	Background colour of prediction intervals.
col.predict.lines	Colour of outer lines of prediction intervals.
equal.size	A logical indicating whether all squares should be of equal size. Otherwise, the square size is proportional to the precision of estimates.
leftcols	A character vector specifying columns to be plotted on the left side of the forest plot (see Details).
leftlabs	A character vector specifying labels for columns on left side of the forest plot.
rightcols	A character vector specifying columns to be plotted on the right side of the forest plot (see Details).
rightlabs	A character vector specifying labels for columns on right side of the forest plot.
digits	Minimal number of significant digits for treatment effects and confidence intervals, see print.default.
digits.prop	Minimal number of significant digits for the direct evidence proportion.
backtransf	A logical indicating whether results should be back transformed in forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
lab.NA	A character string to label missing values.
smlab	A label printed at top of figure. By default, text indicating either common or random effects model is printed.
...	Additional arguments for forest.meta function.

Details

A forest plot, also called confidence interval plot, is drawn in the active graphics window.

The arguments leftcols and rightcols can be used to specify columns which are plotted on the left and right side of the forest plot, respectively. If argument rightcols is FALSE, no columns will be plotted on the right side.

If direct estimates are included in the forest plot (direct = TRUE, default), the following columns will be printed on the left side of the forest plot: the comparisons (column "studlab" in forest.meta), number of pairwise comparisons ("k"), direct evidence proportion ("k"), and I^2 from pairwise comparison ("I2").

If direct estimates are not included in the forest plot (direct = FALSE), only the comparisons ("studlab") are printed on the left side of the forest plot.

For more information see help page of forest.meta function.

Argument show determines which comparisons are printed:

"all"	All comparisons
"both"	Only comparisons contributing both direct and indirect evidence
"with.direct"	Comparisons providing direct evidence
"direct.only"	Comparisons providing only direct evidence
"indirect.only"	Comparisons providing only indirect evidence

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

forest.meta

Examples

```
data(Senn2013)

# Only consider first five studies (to reduce runtime of example)
#
studies <- unique(Senn2013$studlab)
Senn2013.5 <- subset(Senn2013, studlab %in% studies[1:5])

net1 <- netmeta(TE, seTE, treat1.long, treat2.long,
  studlab, data = Senn2013.5, common = FALSE)
#
ns1 <- netsplit(net1)

# Forest plot showing comparisons contributing both direct and
# indirect evidence
#
forest(ns1, fontsize = 6, spacing = 0.5, addrow.subgroups = FALSE)

## Not run:
# Forest plot showing comparisons contributing direct evidence
#
forest(ns1, fontsize = 6, spacing = 0.5, addrow.subgroups = FALSE,
  show = "with.direct")


# Forest plot only showing network estimates compared to reference
# group and prediction intervals
#
forest(ns1, fontsize = 8, spacing = 0.75,
  show = "ref", prediction = TRUE, direct = FALSE, indirect = FALSE)

## End(Not run)
```

Franchini2012 *Network meta-analysis of treatments for Parkinson's disease*

Description

Network meta-analysis comparing the effects of a number of treatments for Parkinson's disease.

The data are the mean lost work-time reduction in patients given dopamine agonists as adjunct therapy in Parkinson's disease (Franchini et al. 2012). The data are given as sample size, mean and standard deviation in each trial arm. Treatments are placebo and four active drugs. These data are used as an example in the supplemental material of Dias et al. (2013) where placebo is coded as 1 and the four active drugs as 2 to 5.

Format

A data frame with the following columns:

Study	study label
Treatment1	treatment 1
y1	treatment effect arm 1
sd1	Standard deviation arm 1
n1	Sample size arm 1
Treatment2	treatment 2
y2	treatment effect arm 2
sd2	Standard deviation arm 2
n2	Sample size arm 2
Treatment3	treatment 3
y3	treatment effect arm 3
sd3	Standard deviation arm 3
n3	Sample size arm 3

Source

Dias S, Sutton AJ, Ades AE and Welton NJ (2013): Evidence synthesis for decision making 2: A generalized linear modeling framework for pairwise and network meta-analysis of randomized controlled trials. *Medical Decision Making*, **33**, 607–17

Franchini AJ, Dias S, Ades AE, Jansen JP, Welton NJ (2012): Accounting for correlation in network meta-analysis with multi-arm trials. *Research Synthesis Methods*, **3**, 142–60

See Also

pairwise, metacont, netmeta, netgraph.netmeta

Examples

```
data(Franchini2012)

# Transform data from arm-based format to contrast-based format
#
p1 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = Franchini2012, studlab = Study)
p1

# Conduct network meta-analysis
net1 <- netmeta(p1)
```

```
net1

# Draw network graphs
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.5,
  thickness = "se.common")
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.5,
  thickness = "se.common",
  iterate = TRUE, plastic = TRUE)
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.5,
  thickness = "se.common",
  iterate = TRUE, plastic = TRUE, start = "eigen")
```

`funnel.netmeta` *'Comparison-adjusted' funnel plot*

Description

Draw a 'comparison-adjusted' funnel plot to assess funnel plot asymmetry in network meta-analysis.

Usage

```
## S3 method for class 'netmeta'
funnel(
  x,
  order,
  pooled = ifelse(x$random, "random", "common"),
  xlab = NULL,
  level = x$level,
  pch,
  col = "black",
  legend = TRUE,
  pos.legend = "topright",
  pos.tests = "topleft",
  lump.comparator = FALSE,
  text.comparator = "comparator",
  method.bias,
  text.linreg = "(Egger)",
  text.rank = "(Begg-Mazumdar)",
  text.mm = "(Thompson-Sharp)",
  sep.trts = x$sep.trts,
  nchar.trts = x$nchar.trts,
  backtransf = x$backtransf,
  digits.pval = gs("digits.pval"),
  warn.deprecated = gs("warn.deprecated"),
  linreg = FALSE,
  rank = FALSE,
  mm = FALSE,
  ...
)
```

Arguments

x	An object of class netmeta.
order	A mandatory character or numerical vector specifying the order of treatments or list of comparators (see Details).
pooled	A character string indicating whether results for the common ("common") or random effects model ("random") should be plotted. Can be abbreviated.
xlab	A label for the x-axis.
level	The confidence level utilised in the plot. For the funnel plot, confidence limits are not drawn if yaxis = "size".
pch	The plotting symbol(s) used for individual studies within direct comparisons.
col	The colour(s) used for individual studies within direct comparisons.
legend	A logical indicating whether a legend with information on direct comparisons should be added to the plot.
pos.legend	The position of the legend describing plotting symbols and colours for direct comparisons.
pos.tests	The position of results for test(s) of funnel plot asymmetry.
lump.comparator	A logical indicating whether comparators should be lumped, e.g., to specify inactive treatments. information on direct comparisons should be added to the plot.
text.comparator	A character string used in the plot to label the comparator if lump.comparator is TRUE.
method.bias	A character vector indicating which test(s) for funnel plot asymmatrx to use. Admissible values are "Begg", "Egger", and "Thompson", can be abbreviated. See function metabias.
text.linreg	A character string used in the plot to label the Egger test for funnel plot asymmetry.
text.rank	A character string used in the plot to label the Begg test for funnel plot asymmetry.
text.mm	A character string used in the plot to label the Thompson-Sharp test for funnel plot asymmetry.
sep.trts	A character used in comparison names as separator between treatment labels.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names (see netmeta).
backtransf	A logical indicating whether results for relative summary measures (argument sm equal to "OR", "RR", "HR", or "IRR") should be back transformed in funnel plots. If backtransf = TRUE, results for sm = "OR" are printed as odds ratios rather than log odds ratios, for example.
digits.pval	Minimal number of significant digits for p-value of test(s) for funnel plot asymmetry.

warn.deprecated
: A logical indicating whether warnings should be printed if deprecated arguments are used.

linreg
: Deprecated argument (replaced by method.bias).

rank
: Deprecated argument (replaced by method.bias).

mm
: Deprecated argument (replaced by method.bias).

...
: Additional graphical arguments passed as arguments to funnel.meta.

Details

A 'comparison-adjusted' funnel plot (Chaimani & Salanti, 2012) is drawn in the active graphics window.

Argument order is mandatory to determine the order of treatments (Chaimani et al., 2013):

"Before using this plot, investigators should order the treatments in a meaningful way and make assumptions about how small studies differ from large ones. For example, if they anticipate that newer treatments are favored in small trials, then they could name the treatments from oldest to newest so that all comparisons refer to 'old versus new intervention'. Other possibilities include defining the comparisons so that all refer to an active treatment versus placebo or sponsored versus non-sponsored intervention."

Alternatively, it is possible to only provide a single or few treatment name(s) in argument order to define the comparator(s). In this case only comparisons with this / these treatment(s) will be considered. If argument lump.comparator is TRUE, all comparators will be lumped into a single group. The text for this group can be specified with argument text.comparator.

In the funnel plot, if yaxis is "se", the standard error of the treatment estimates is plotted on the y-axis which is likely to be the best choice (Sterne & Egger, 2001). Other possible choices for yaxis are "invvar" (inverse of the variance), "invse" (inverse of the standard error), and "size" (study size).

Value

A data frame with the following columns:

studlab
: Study label.

treat1
: Label/Number for first treatment.

treat2
: Label/Number for second treatment.

comparison
: Treatment comparison.

TE
: Estimate of treatment effect, e.g., log odds ratio.

TE.direct
: Pooled estimate from direct evidence.

TE.adj
: 'Comparison-adjusted' treatment effect (TE - TE.direct).

seTE
: Standard error of treatment estimate.

pch
: Plotting symbol(s).

col
: Colour of plotting symbol(s).

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Chaimani A & Salanti G (2012): Using network meta-analysis to evaluate the existence of small-study effects in a network of interventions. *Research Synthesis Methods*, **3**, 161–76

Chaimani A, Higgins JP, Mavridis D, Spyridonos P, Salanti G (2013): Graphical tools for network meta-analysis in STATA. PLOS ONE, **8**, e76654

Sterne JAC & Egger M (2001): Funnel plots for detecting bias in meta-analysis: Guidelines on choice of axis. *Journal of Clinical Epidemiology*, **54**, 1046–55

See Also

netmeta, funnel.meta, metabias

Examples

```
data(Senn2013)

net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD")

# 'Comparison-adjusted' funnel plot not created as argument 'order'
# is missing
#
try(funnel(net1))

# Only show comparisons with placebo
#
funnel(net1, order = "pl")

# Add result for Egger test of funnel plot asymmetry
#
funnel(net1, order = "pl", method.bias = "Egger",
  digits.pval = 2)

# (Non-sensical) alphabetic order of treatments with placebo as
# last treatment
#
ord <- c("a", "b", "me", "mi", "pi", "r", "si", "su", "v", "pl")
funnel(net1, order = ord)

## Not run:
# Add results for tests of funnel plot asymmetry and use different
# plotting symbols and colours
#
funnel(net1, order = ord,
  pch = rep(c(15:18, 1), 3), col = 1:3,
  method.bias = c("Egger", "Begg", "Thompson"), digits.pval = 2)
```

```
# Same results for tests of funnel plot asymmetry using reversed
# order of treatments
#
funnel(net1, order = rev(ord),
  pch = rep(c(15:18, 1), 3), col = 1:3,
  method.bias = c("Egger", "Begg", "Thompson"), digits.pval = 2)

# Calculate tests for funnel plot asymmetry
#
f1 <- funnel(net1, order = ord)
#
metabias(metagen(TE.adj, seTE, data = f1))
metabias(metagen(TE.adj, seTE, data = f1), method = "Begg")
metabias(metagen(TE.adj, seTE, data = f1), method = "Thompson")

## End(Not run)
```

Gurusamy2011 *Network meta-analysis on blood loss during liver transplantation*

Description

Network meta-analysis comparing the effects of a number of interventions for decreasing blood loss and blood transfusion requirements during liver transplantation.

Format

A data frame with the following columns:

study	study information (first author, year)
treatment	treatment
death	mortality at 60 days post-transplantation
n	number of individuals in treatment arm

Source

Gurusamy KS, Pissanou T, Pikhart H, Vaughan J, Burroughs AK, Davidson BR (2011): Methods to decrease blood loss and transfusion requirements for liver transplantation. *Cochrane Database of Systematic Reviews*, CD009052

See Also

pairwise, metabin, netmetabin

Examples

```
data(Gurusamy2011)
```

```
# Only consider three studies (to reduce runtime of example)
#
studies <- c("Findlay 2001", "Garcia-Huete 1997", "Dalmau 2000")
three <- subset(Gurusamy2011, study %in% studies)

# Transform data from long arm-based format to contrast-based
# format. Argument 'sm' has to be used for odds ratio as summary
# measure; by default the risk ratio is used in the metabin
# function called internally.
#
p1 <- pairwise(treatment, death, n, studlab = study,
  data = three, sm = "OR")

# Conduct Mantel-Haenszel network meta-analysis
#
netmetabin(p1, ref = "cont")

## Not run:
p2 <- pairwise(treatment, death, n, studlab = study,
  data = Gurusamy2011, sm = "OR")

# Conduct Mantel-Haenszel network meta-analysis
netmetabin(p2, ref = "cont")

## End(Not run)
```

`hasse` *Hasse diagram*

Description

This function generates a Hasse diagram for a partial order of treatment ranks in a network meta-analysis.

Usage

```
hasse(x, pooled = ifelse(x$random, "random", "common"), newpage = TRUE)
```

Arguments

`x`	An object of class `netposet` (mandatory).
`pooled`	A character string indicating whether Hasse diagram show be drawn for common (`"common"`) or random effects model (`"random"`). Can be abbreviated.
`newpage`	A logical value indicating whether a new figure should be printed in an existing graphics window. Otherwise, the Hasse diagram is added to the existing figure.

Details

Generate a Hasse diagram (Carlsen & Bruggemann, 2014) for a partial order of treatment ranks in a network meta-analysis (Rücker & Schwarzer, 2017).

This R function is a wrapper function for R function hasse in R package **hasseDiagram** (Krzysztof Ciomek, https://github.com/kciomek/hasseDiagram), i.e., function hasse can only be used if R package **hasseDiagram** is installed.

Author(s)

Gerta Rücker <ruecker@imbi.uni-freiburg.de>, Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Carlsen L, Bruggemann R (2014): Partial order methodology: a valuable tool in chemometrics. *Journal of Chemometrics*, **28**, 226–34

Rücker G, Schwarzer G (2017): Resolve conflicting rankings of outcomes in network meta-analysis: Partial ordering of treatments. *Research Synthesis Methods*, **8**, 526–36

See Also

netmeta, netposet, netrank, plot.netrank

Examples

```
## Not run:
# Use depression dataset
#
data(Linde2015)

# Define order of treatments
#
trts <- c("TCA", "SSRI", "SNRI", "NRI",
  "Low-dose SARI", "NaSSa", "rMAO-A", "Hypericum", "Placebo")

# Outcome labels
#
outcomes <- c("Early response", "Early remission")

# (1) Early response
#
p1 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(resp1, resp2, resp3),
  n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net1 <- netmeta(p1, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "bad")

# (2) Early remission
#
```

```
p2 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(remi1, remi2, remi3),
  n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net2 <- netmeta(p2, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "bad")

# Partial order of treatment rankings
#
po <- netposet(netrank(net1), netrank(net2), outcomes = outcomes)

# Hasse diagram
#
hasse(po)

## End(Not run)
```

---

hatmatrix *Derive hat matrix from network meta-analysis*

Description

Auxiliary function to derive hat matrix from network meta-analysis

Usage

```
hatmatrix(
  x,
  method = "Ruecker",
  type,
  common = x$common,
  random = x$random,
  nchar.trts = x$nchar.trts,
  nchar.studlab = x$nchar.studlab
)

## S3 method for class 'hatmatrix'
print(
  x,
  common = x$x$common,
  random = x$x$random,
  nchar.trts = x$x$nchar.trts,
  nchar.studlab = x$x$nchar.studlab,
  digits = gs("digits"),
  legend = TRUE,
  legend.studlab = TRUE,
  ...
)
```

Arguments

x	A netmeta object.
method	A character string indicating which method is used to derive the hat matrix. Either "Ruecker", "Krahn" or "Davies" (can be abbreviated, see Details).
type	A character string indicating which specific hat matrix should be derived (can be abbreviated, see Details).
common	A logical indicating whether a hat matrix should be printed for the common effects network meta-analysis.
random	A logical indicating whether a hat matrix should be printed for the random effects network meta-analysis.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
nchar.studlab	A numeric defining the minimum number of characters used to create unique study labels.
digits	Minimal number of significant digits, see print.default.
legend	A logical indicating whether a legend should be printed.
legend.studlab	A logical indicating whether a legend should be printed for abbreviated study labels.
...	Additional arguments (ignored).

Details

This auxiliary function can be used to derive various hat matrices from a network meta-analysis object.

Hat matrix by Rücker (2012):

This hat matrix is estimated if method = "Ruecker".

Let n be the number of different treatments (nodes, vertices) in a network and let m be the number of existing comparisons (edges) between the treatments. If there are only two-arm studies, m is equal to the number of studies, k. Let seTE.adj.common and seTE.adj.random be the vectors of adjusted standard errors under the common and random effects model (see netmeta). Let $\mathbf{W}$ be the m x m diagonal matrix that contains the inverse variance 1 / seTE.adj.common^2 or 1 / seTE.adj.random^2.

The given comparisons define the network structure. Therefrom an m x n design matrix X (edge-vertex incidence matrix) is formed; for more precise information, see Rücker (2012). Moreover, the n x n Laplacian matrix $\mathbf{L}$ and its Moore-Penrose pseudoinverse $\mathbf{L}^+$ are calculated (both matrices play an important role in graph theory and electrical network theory). Using these matrices, the variances based on both direct and indirect comparisons can be estimated. The hat matrix $\mathbf{H}$ is estimated by $\mathbf{H = XL^+X^TW = X(X^TW\,X)^+X^TW}$.

Hat matrices by Krahn et al. (2013):

One of the following hat matrices is estimated if method = "Krahn".

Use of type = "design" (default) results in a hat matrix of dimension $n(n-1)/2$ x d, where d is the sum of the number of independent comparisons from each design.

Use of type = "studies" results in a hat matrix of dimension $n(n-1)/2$ x l, where l is the number of independent pairwise comparisons, i.e., a three-arm study contributes two pairwise comparisons.

Hat matrices by Davies et al. (2021):

One of three hat matrices is estimated if `method = "Davies"`.

Here, we focus on the hat matrix of the aggregate (two-step) version of the graph theoretical NMA model. In the first step, a pairwise meta-analysis is performed across each edge using the adjusted weights (these account for correlations due to multi-arm trials). From this we obtain direct treatment effect estimates (and corresponding aggregate weights) associated with each edge. In step two, we combine these direct estimates in a network meta analysis to obtain the network estimates. This is done using weighted least squares regression. The hat matrix associated with this second step is called the *aggregate hat matrix*.

All three versions of the aggregate hat matrix contain the same information: the second two can be derived directly from the first. They differ in their dimensionality.

Each row of the hat matrix that represents a treatment comparison (*ij*) describes the flow of evidence through each edge for that comparison. This defines a directed acyclic 'flow graph' from node *i* to node *j*.

(1) Use of `type = "short"` (default) results in a hat matrix of dimension *e* x *e*, where *e* is the number of (unique) edges (direct comparisons) in the network. This is the aggregate hat matrix described in Davies et al. (2021). Each row and column represents a pair of treatments for which there is at least one direct comparison.

(2) Use of `type = "long"` results in a hat matrix of dimension *n(n-1)/2* x *e*. There is a row for every possible pair of treatments in the network - regardless of whether there is direct evidence for this comparison. Each column represents a pair of treatments for which there is at least one direct comparison. The extra rows can be calculated from the short hat matrix using consistency equations.

(3) Use of `type = "full"` results in a hat matrix of dimension *n(n-1)/2* x *n(n-1)/2*. In comparison to the long hat matrix, columns of zeroes are added for comparisons that do not have any direct evidence. Therefore, there is a row and column for every pair of treatments in the network. This hat matrix is used to calculate the transition matrices for the random walk in `netcontrib`.

Value

A list with two hat matrices: `common` (common effects model) and `random` (random effects model).

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Davies AL, Papakonstantinou T, Nikolakopoulou A, Rücker G, Galla T (2021): Network meta-analysis and random walks. Available from: http://arxiv.org/abs/2107.02886

Krahn U, Binder H, König J (2013): A graphical tool for locating inconsistency in network meta-analyses. *BMC Medical Research Methodology*, **13**, 35

Rücker G (2012): Network meta-analysis, electrical networks and graph theory. *Research Synthesis Methods*, **3**, 312–24

See Also

`netmeta`, `netcontrib`, `netheat`

Examples

```
data(Dong2013)
# Only consider first ten studies for concise output
first10 <- subset(Dong2013, id <= 10)
p1 <- pairwise(treatment, death, randomized, studlab = id,
  data = first10, sm = "OR")
net1 <- netmeta(p1, common = FALSE)

hatmatrix(net1)
hatmatrix(net1, method = "k")
hatmatrix(net1, method = "k", type = "studies")
hatmatrix(net1, method = "d")
hatmatrix(net1, method = "d", type = "long")
hatmatrix(net1, method = "d", type = "full")
```

---

invmat *Moore-Penrose Pseudoinverse of a Matrix*

Description

Calculates the Moore-Penrose pseudoinverse of a square matrix **X**.

Usage

```
invmat(X)
```

Arguments

X	A square matrix.

Details

This function is used by default in R package **netmeta** to calculate the Moore-Penrose pseudoinverse $\mathbf{L}^+$ of the Laplacian matrix **L** (Rücker, 2012):

$\mathbf{L}^+ = (\mathbf{X} - \mathbf{J} / n)^{-1} + \mathbf{J} / n$ with identity matrix **J** of dimension nxn.

Value

The Moore-Penrose pseudoinverse for matrix **X**.

Author(s)

Gerta Rücker <ruecker@imbi.uni-freiburg.de>, Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Rücker G (2012): Network meta-analysis, electrical networks and graph theory. *Research Synthesis Methods*, **3**, 312–24

See Also

netmeta, solve

Examples

```
data(Senn2013)

net1 <- netmeta(TE, seTE, treat1.long, treat2.long, studlab,
  data = Senn2013)
L1 <- net1$L.matrix.common
L2 <- invmat(net1$Lplus.matrix.common)
all.equal(round(L1, 10), round(L2, 10))
```

Linde2015 *Network meta-analysis of treatments for depression*

Description

Network meta-analysis of nine classes of antidepressants including placebo for the primary care setting; partly shown in Linde et al. (2015), supplementary Table 2.

Format

A data frame with the following columns:

id	Study ID
author	First author
year	Publication year
treatment1	First treatment
treatment2	Second treatment
treatment3	Third treatment
n1	Number of patients receiving first treatment
resp1	Number of early responder (treatment 1)
remi1	Number of early remissions (treatment 1)
loss1	Number of patients loss to follow-up (treatment 1)
loss.ae1	Number of patients loss to follow-up due to adverse events (treatment 1)
ae1	Number of patients with adverse events (treatment 1)
n2	Number of patients receiving second treatment
resp2	Number of early responder (treatment 2)
remi2	Number of early remissions (treatment 2)
loss2	Number of patients loss to follow-up (treatment 2)
loss.ae2	Number of patients loss to follow-up due to adverse events (treatment 2)
ae2	Number of patients with adverse events (treatment 2)
n3	Number of patients receiving third treatment
resp3	Number of early responder (treatment 3)
remi3	Number of early remissions (treatment 3)

loss3	Number of patients loss to follow-up (treatment 3)
loss.ae3	Number of patients loss to follow-up due to adverse events (treatment 3)
ae3	Number of patients with adverse events (treatment 3)

Source

Linde K, Kriston L, Rücker G, et al. (2015): Efficacy and acceptability of pharmacological treatments for depressive disorders in primary care: Systematic review and network meta-analysis. *Annals of Family Medicine*, **13**, 69–79

See Also

pairwise, metabin, netmeta, netposet

Examples

```
data(Linde2015)

# Transform data from arm-based format to contrast-based format
# Outcome: early response
p1 <- pairwise(list(treatment1, treatment2, treatment3),
  event = list(resp1, resp2, resp3),
  n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
p1

## Not run:
# Define order of treatments
trts <- c("TCA", "SSRI", "SNRI", "NRI", "Low-dose SARI",
  "NaSSa", "rMAO-A", "Hypericum", "Placebo")

# Conduct random effects network meta-analysis
net1 <- netmeta(p1, common = FALSE, reference = "Placebo", seq = trts)
print(net1, digits = 2)

## End(Not run)
```

Linde2016 *Network meta-analysis of primary care depression treatments*

Description

Network meta-analysis of 22 treatments (including placebo and usual care) for the primary care of depression.

Format

A data frame with the following columns:

id	Study ID
author	First author
year	Year of publication
treat1	First treatment (abbreviated)
treat2	Second treatment (abbreviated)
treat1.long	First treatment
treat2.long	Second treatment
lnOR	Response after treatment (log odds ratio)
selnOR	Standard error of log odds ratio
resp1	Responder (first treatment)
n1	Sample size (first treatment)
resp2	Responder (second treatment)
n2	Sample size (second treatment)

Source

Linde K, Rücker G, Schneider A et al. (2016): Questionable assumptions hampered interpretation of a network meta-analysis of primary care depression treatments. *Journal of Clinical Epidemiology*, **71**, 86–96

See Also

netmeta, netcomb

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, reference.group = "placebo",
  sm = "OR", common = FALSE, nchar.trts = 6)
#
net1

## Not run:
# Conduct random effects network meta-analysis
#
net2 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = Linde2016, reference.group = "placebo",
```

```
  sm = "OR", common = FALSE, nchar.trts = 6)
#
net2

## End(Not run)
```

`metabias.netmeta` *Test of funnel plot asymmetry in network meta-analysis*

Description

Test of funnel plot asymmetry in network meta-analysis

Usage

```
## S3 method for class 'netmeta'
metabias(
  x,
  order,
  pooled = ifelse(x$random, "random", "common"),
  method.bias = "Egger",
  lump.comparator = FALSE,
  ...
)
```

Arguments

`x`	An object of class `netmeta`.
`order`	A mandatory character or numerical vector specifying the order of treatments or list of comparators (see Details).
`pooled`	A character string indicating whether results for the common (`"common"`) or random effects model (`"random"`) should be used in test of funnel plot asymmetry. Can be abbreviated.
`method.bias`	A character vector indicating which test(s) for funnel plot asymmatrx to use. Admissible values are `"Begg"`, `"Egger"`, and `"Thompson"`, can be abbreviated. See function `metabias.meta`.
`lump.comparator`	A logical indicating whether comparators should be lumped, e.g., to specify inactive treatments. information on direct comparisons should be added to the plot.
`...`	Additional arguments (passed on to `metabias.meta`).

Details

Test of funnel plot asymmetry in network meta-analysis

Argument `order` is mandatory to determine the order of treatments (Chaimani et al., 2013):

"[...] investigators should order the treatments in a meaningful way and make assumptions about how small studies differ from large ones. For example, if they anticipate that newer treatments are favored in small trials, then they could name the treatments from oldest to newest so that all comparisons refer to 'old versus new intervention'. Other possibilities include defining the comparisons so that all refer to an active treatment versus placebo or sponsored versus non-sponsored intervention."

Alternatively, it is possible to only provide a single or few treatment name(s) in argument `order` to define the comparator(s). In this case only comparisons with this / these treatment(s) will be considered. If argument `lump.comparator` is `TRUE`, all comparators will be lumped into a single group.

Value

A list with class `metabias` containing the following components if a test for funnel plot asymmetry is conducted:

`statistic`	Test statistic.
`df`	The degrees of freedom of the test statistic in the case that it follows a t distribution.
`pval`	The p-value for the test.
`estimate`	Estimates used to calculate test statisic.
`method`	A character string indicating what type of test was used.
`title`	Title of Cochrane review.
`complab`	Comparison label.
`outclab`	Outcome label.
`var.model`	A character string indicating whether none, multiplicative, or additive residual heterogeneity variance was assumed.
`method.bias`	As defined above.
`x`	Network meta-analysis object.
`version`	Version of R package **meta** used to create object.
`version.netmeta`	Version of R package **netmeta** used to create object.

Or a list with the following elements if test is not conducted due to the number of studies:

`k`	Number of comparisons.
`k.min`	Minimum number of comparisons to perform test for funnel plot asymmetry.
`version`	Version of R package **meta** used to create object.
`version.netmeta`	Version of R package **netmeta** used to create object.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Chaimani A & Salanti G (2012): Using network meta-analysis to evaluate the existence of small-study effects in a network of interventions. *Research Synthesis Methods*, **3**, 161–76

Chaimani A, Higgins JP, Mavridis D, Spyridonos P, Salanti G (2013): Graphical tools for network meta-analysis in STATA. PLOS ONE, **8**, e76654

See Also

netmeta, funnel.netmeta, metabias

Examples

```
data(Senn2013)

net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD")

# Test for asymmetry in 'comparison-adjusted' funnel plot not
# conducted as argument 'order' is missing
#
try(metabias(net1))

# Test for funnel plot asymmetry comparing active treatments with
# placebo
metabias(net1, order = "pl")

# Rank test
#
metabias(net1, order = "pl", method.bias = "Begg")


# Test for funnel plot asymmetry based on (non-sensical) alphabetic
# order of treatments with placebo as last treatment
#
ord <- c("a", "b", "me", "mi", "pi", "r", "si", "su", "v", "pl")
metabias(net1, order = ord)
```

---

netbind *Combine network meta-analysis objects*

Description

This function can be used to combine network meta-analysis objects which is especially useful to generate a forest plot with results of several network meta-analyses.

Usage

```
netbind(
  ...,
  name,
  common,
  random,
  col.study = "black",
  col.inside = "white",
  col.square = "gray",
  col.square.lines = col.square,
  backtransf,
  reference.group,
  baseline.reference,
  warn.deprecated = gs("warn.deprecated"),
  fixed,
  comb.fixed,
  comb.random
)
```

Arguments

`...`	Any number of network meta-analysis objects or a single list with network meta-analyses.
`name`	An optional character vector providing descriptive names for network meta-analysis objects.
`common`	A logical indicating whether results for the common effects model should be reported.
`random`	A logical indicating whether results for the random effects model should be reported.
`col.study`	The colour for network estimates and confidence limits.
`col.inside`	The colour for network estimates and confidence limits if confidence limits are completely within squares.
`col.square`	The colour for squares.
`col.square.lines`	The colour for the outer lines of squares.
`backtransf`	A logical indicating whether results should be back transformed. If `backtransf = TRUE` (default), results for `sm = "OR"` are printed as odds ratios rather than log odds ratios, for example.
`reference.group`	Reference treatment.
`baseline.reference`	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa. This argument is only considered if `reference.group` has been specified.

warn.deprecated
: A logical indicating whether warnings should be printed if deprecated arguments are used.

fixed
: Deprecated argument; replaced by common.

comb.fixed
: Deprecated argument; replaced by common.

comb.random
: Deprecated argument; replaced by random.

Value

An object of class "netbind" with corresponding forest function. The object is a list containing the following components:

common
: A data frame with results for the common effects model.

random
: A data frame with results for the random effects model.

sm
: Summary measure used in network meta-analyses.

level.ma
: Level for confidence intervals.

reference.group, baseline.reference
: As defined above.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netmeta, netcomb, discomb, forest.netbind

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Standard random effects NMA model (with placebo as reference
# treatment)
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, reference.group = "placebo",
  sm = "OR", common = FALSE)

# Additive CNMA model with placebo as inactive component and
# reference
#
nc1 <- netcomb(net1, inactive = "placebo")

# Combine results of standard NMA and CNMA
#
```

```
nb1 <- netbind(nc1, net1,
  name = c("Additive CNMA", "Standard NMA"),
  col.study = c("red", "black"), col.square = c("red", "black"))
forest(nb1,
  col.by = "black", addrow.subgroups = FALSE,
  fontsize = 10, spacing = 0.7, squaresize = 0.9,
  label.left = "Favours Placebo",
  label.right = "Favours other")
```

---

netcomb *Additive network meta-analysis for combinations of treatments*

Description

Some treatments in a network meta-analysis may be combinations of other treatments or have common components. The influence of individual components can be evaluated in an additive network meta-analysis model assuming that the effect of treatment combinations is the sum of the effects of its components. This function implements this additive model in a frequentist way.

Usage

```
netcomb(
  x,
  inactive = NULL,
  sep.comps = "+",
  C.matrix,
  common = x$common,
  random = x$random | !is.null(tau.preset),
  tau.preset = NULL,
  details.chkident = FALSE,
  nchar.comps = x$nchar.trts,
  func.inverse = invmat,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netmeta.
inactive	A character string defining the inactive treatment component (see Details).
sep.comps	A single character to define separator between treatment components.
C.matrix	C matrix (see Details).
common	A logical indicating whether a common effects network meta-analysis should be conducted.
random	A logical indicating whether a random effects network meta-analysis should be conducted.

`tau.preset` An optional value for the square-root of the between-study variance τ^2.

`details.chkident`
A logical indicating whether details on unidentifiable components should be printed.

`nchar.comps` A numeric defining the minimum number of characters used to create unique names for components (see Details).

`func.inverse` R function used to calculate the pseudoinverse of the Laplacian matrix L (see `netmeta`).

`warn.deprecated`
A logical indicating whether warnings should be printed if deprecated arguments are used.

`...` Additional arguments (to catch deprecated arguments).

Details

Treatments in network meta-analysis (NMA) can be complex interventions. Some treatments may be combinations of others or have common components. The standard analysis provided by `netmeta` is a NMA where all existing (single or combined) treatments are considered as different nodes in the network. Exploiting the fact that some treatments are combinations of common components, an additive component network meta-analysis (CNMA) model can be used to evaluate the influence of individual components. This model assumes that the effect of a treatment combination is the sum of the effects of its components which implies that common components cancel out in comparisons.

The additive CNMA model has been implemented using Bayesian methods (Mills et al., 2012; Welton et al., 2013). This function implements the additive model in a frequentist way (Rücker et al., 2020).

The underlying multivariate model is given by

$$\boldsymbol{\delta} = \boldsymbol{B}\boldsymbol{\theta}, \boldsymbol{\theta} = \boldsymbol{C}\boldsymbol{\beta}$$

with

$\boldsymbol{\delta}$ vector of true treatment effects (differences) from individual studies,

$\boldsymbol{B}$ design matrix describing the structure of the network,

$\boldsymbol{\theta}$ parameter vector that represents the existing combined treatments,

$\boldsymbol{C}$ matrix describing how the treatments are composed,

$\boldsymbol{\beta}$ parameter vector representing the treatment components.

All parameters are estimated using weighted least squares regression.

Argument `inactive` can be used to specify a single component that does not have any therapeutic value. Accordingly, it is assumed that the treatment effect of the combination of this component with an additional treatment component is equal to the treatment effect of the additional component alone.

Argument `sep.comps` can be used to specify the separator between individual components. By default, the matrix **C** is calculated internally from treatment names. However, it is possible to specify a different matrix using argument `C.matrix`.

By default, component names are not abbreviated in printouts. However, in order to get more concise printouts, argument nchar.comps can be used to define the minimum number of characters for abbreviated component names (see abbreviate, argument minlength). R function treats is utilised internally to create abbreviated component names.

Value

An object of class netcomb with corresponding print, summary, and forest functions. The object is a list containing the following components:

studlab	Study labels.
treat1	Label/Number for first treatment.
treat2	Label/Number for second treatment.
TE	Estimate of treatment effect, i.e. difference between first and second treatment.
seTE	Standard error of treatment estimate.
seTE.adj.common, seTE.adj.random	Standard error of treatment estimate, adjusted for multi-arm studies.
design	Design of study providing pairwise comparison.
event1	Number of events in first treatment group.
event2	Number of events in second treatment group.
n1	Number of observations in first treatment group.
n2	Number of observations in second treatment group.
k	Total number of studies.
m	Total number of pairwise comparisons.
n	Total number of treatments.
d	Total number of designs (corresponding to the unique set of treatments compared within studies).
c	Total number of components.
trts	Treatments included in network meta-analysis.
k.trts	Number of studies evaluating a treatment.
n.trts	Number of observations receiving a treatment (if arguments n1 and n2 are provided).
events.trts	Number of events observed for a treatment (if arguments event1 and event2 are provided).
studies	Study labels coerced into a factor with its levels sorted alphabetically.
narms	Number of arms for each study.
designs	Unique list of designs present in the network. A design corresponds to the set of treatments compared within a study.
comps	Unique list of components present in the network.
TE.nma.common, TE.nma.random	A vector of length *m* of consistent treatment effects estimated by network meta-analysis (nma) (common and random effects model).

`seTE.nma.common`, `seTE.nma.random`
: A vector of length *m* of effective standard errors estimated by network meta-analysis (common and random effects model).

`lower.nma.common`, `lower.nma.random`
: A vector of length *m* of lower confidence interval limits for consistent treatment effects estimated by network meta-analysis (common and random effects model).

`upper.nma.common`, `upper.nma.random`
: A vector of length *m* of upper confidence interval limits for the consistent treatment effects estimated by network meta-analysis (common and random effects model).

`statistic.nma.common`, `statistic.nma.random`
: A vector of length *m* of z-values for test of treatment effect for individual comparisons (common and random effects model).

`pval.nma.common`, `pval.nma.random`
: A vector of length *m* of p-values for test of treatment effect for individual comparisons (common and random effects model).

`TE.cnma.common`, `TE.cnma.random`
: A vector of length *m* of consistent treatment effects estimated by the additive (common and random effects) model.

`seTE.cnma.common`, `seTE.cnma.random`
: A vector of length *m* with standard errors estimated by the additive (common and random effects) model.

`lower.cnma.common`, `lower.cnma.random`
: A vector of length *m* of lower confidence interval limits for consistent treatment effects estimated by the additive (common and random effects) model.

`upper.cnma.common`, `upper.cnma.random`
: A vector of length *m* of upper confidence interval limits for consistent treatment effects estimated by the additive (common and random effects) model.

`statistic.cnma.common`, `statistic.cnma.random`
: A vector of length *m* of z-values for the test of an overall effect estimated by the additive (common and random effects) model.

`pval.cnma.common`, `pval.cnma.random`
: A vector of length *m* of p-values for the test of an overall effect estimated by the additive (common and random effects) model.

`TE.common`, `TE.random`
: *n*x*n* matrix with overall treatment effects estimated by the additive (common and random effects) model.

`seTE.common`, `seTE.random`
: *n*x*n* matrix with standard errors estimated by the additive (common and random effects) model.

`lower.common`, `upper.common`, `lower.random`, `upper.random`
: *n*x*n* matrices with lower and upper confidence interval limits estimated by the additive (common and random effects) model.

`statistic.common`, `pval.common`, `statistic.random`, `pval.random`
: *n*x*n* matrices with z-values and p-values for test of overall effect estimated by the additive (common and random effects) model.

`Comp.common, Comp.random`
: A vector of component effects (common and random effects model).

`seComp.common, seComp.random`
: A vector with corresponding standard errors (common and random effects model).

`lower.Comp.common, lower.Comp.random`
: A vector with lower confidence limits for components (common and random effects model).

`upper.Comp.common, upper.Comp.random`
: A vector with upper confidence limits for components (common and random effects model).

`statistic.Comp.common, statistic.Comp.random`
: A vector with z-values for the overall effect of components (common and random effects model).

`pval.Comp.common, pval.Comp.random`
: A vector with p-values for the overall effect of components (common and random effects model).

`Comb.common, Comb.random`
: A vector of combination effects (common and random effects model).

`seComb.common, seComb.random`
: A vector with corresponding standard errors (common and random effects model).

`lower.Comb.common, lower.Comb.random`
: A vector with lower confidence limits for combinations (common and random effects model).

`upper.Comb.common, upper.Comb.random`
: A vector with upper confidence limits for combinations (common and random effects model).

`statistic.Comb.common, statistic.Comb.random`
: A vector with z-values for the overall effect of combinations (common and random effects model).

`pval.Comb.common, pval.Comb.random`
: A vector with p-values for the overall effect of combinations (common and random effects model).

`Q.additive`
: Overall heterogeneity / inconsistency statistic (additive model).

`df.Q.additive`
: Degrees of freedom for test of heterogeneity / inconsistency (additive model).

`pval.Q.additive`
: P-value for test of heterogeneity / inconsistency (additive model).

`tau`
: Square-root of between-study variance (additive model).

`I2, lower.I2, upper.I2`
: I-squared, lower and upper confidence limits.

`Q.standard`
: Overall heterogeneity / inconsistency statistic (standard model).

`df.Q.standard`
: Degrees of freedom for test of heterogeneity / inconsistency (standard model).

`pval.Q.standard`
: P-value for test of heterogeneity / inconsistency (standard model).

`Q.diff`
: Test statistic for difference in goodness of fit between standard and additive model.

df.Q.diff	Degrees of freedom for difference in goodness of fit between standard and additive model.
pval.Q.diff	P-value for difference in goodness of fit between standard and additive model.
A.matrix	Adjacency matrix (*n*x*n*).
B.matrix	Edge-vertex incidence matrix (*m*x*n*).
C.matrix	As defined above.
sm	Summary measure.
level.ma	Level for confidence intervals.
common, random, tau.preset	As defined above.
sep.trts	A character used in comparison names as separator between treatment labels.
nchar.comps	A numeric defining the minimum number of characters used to create unique component names.
inactive, sep.comps	As defined above.
backtransf	A logical indicating whether results should be back transformed in printouts and forest plots.
title	Title of meta-analysis / systematic review.
x	As defined above.
call	Function call.
version	Version of R package netmeta used to create object.

Note

This function calculates effects for individual components and complex interventions present in the network.

R function netcomplex can be used to calculate the effect for arbitrary complex interventions in a component network meta-analysis. Furthermore, R function netcomparison can be used to calculate the effect for comparisons of two arbitrary complex intervention in a component network meta-analysis.

Author(s)

Gerta Rücker <ruecker@imbi.uni-freiburg.de>, Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

König J, Krahn U, Binder H (2013): Visualizing the flow of evidence in network meta-analysis and characterizing mixed treatment comparisons. *Statistics in Medicine*, **32**, 5414–29

Mills EJ, Thorlund K, Ioannidis JP (2012): Calculating additive treatment effects from multiple randomized trials provides useful estimates of combination therapies. *Journal of Clinical Epidemiology*, **65**, 1282–8

Rücker G, Petropoulou M, Schwarzer G (2020): Network meta-analysis of multicomponent interventions. *Biometrical Journal*, **62**, 808–21

Welton NJ, Caldwell DM, Adamopoulos E, Vedhara K (2009): Mixed treatment comparison meta-analysis of complex interventions: psychological interventions in coronary heart disease. *American Journal of Epidemiology*, **169**: 1158–65

See Also

discomb, netmeta, forest.netcomb, print.netcomb, netcomplex, netcomparison

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, ref = "placebo", sm = "OR", common = FALSE)
net1
forest(net1, xlim = c(0.2, 50))

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")
nc1
forest(nc1, xlim = c(0.2, 50))

## Not run:
# Specify, order of treatments
#
trts <- c("TCA", "SSRI", "SNRI", "NRI", "Low-dose SARI", "NaSSa",
  "rMAO-A", "Ind drug", "Hypericum", "Face-to-face CBT",
  "Face-to-face PST", "Face-to-face interpsy", "Face-to-face psychodyn",
  "Other face-to-face", "Remote CBT", "Self-help CBT", "No contact CBT",
  "Face-to-face CBT + SSRI", "Face-to-face interpsy + SSRI",
  "Face-to-face PST + SSRI", "UC", "Placebo")
#
# Note, three treatments are actually combinations of 'SSRI' with
# other components:
# "Face-to-face CBT + SSRI",
# "Face-to-face interpsy + SSRI",
# "Face-to-face PST + SSRI"

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = Linde2016, ref = "placebo",
  seq = trts, sm = "OR", common = FALSE)
```

```
net1
forest(net1, xlim = c(0.2, 50))

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")
nc1
forest(nc1, xlim = c(0.2, 50))

## End(Not run)
```

`netcomparison` *Calculate comparison effects of two arbitrary complex interventions in component network meta-analysis*

Description

Calculate comparison effects of two arbitrary complex interventions (i.e., combinations of several components) in component network meta-analysis.

Usage

```
netcomparison(
  x,
  treat1,
  treat2,
  common = x$common,
  random = x$random,
  level = x$level.ma,
  nchar.comps = x$nchar.comps,
  warn.deprecated = gs("warn.deprecated"),
  ...
)

## S3 method for class 'netcomparison'
print(
  x,
  common = x$common,
  random = x$random,
  backtransf = x$backtransf,
  nchar.comps = x$nchar.comps,
  digits = gs("digits"),
  digits.stat = gs("digits.stat"),
  digits.pval = gs("digits.pval"),
  scientific.pval = gs("scientific.pval"),
  zero.pval = gs("zero.pval"),
```

```
  JAMA.pval = gs("JAMA.pval"),
  big.mark = gs("big.mark"),
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netcomb or netcomparison (print function).
treat1	A character vector defining the first complex intervention(s).
treat2	A character vector defining the second complex intervention(s).
common	A logical indicating whether results for common effects model should be conducted.
random	A logical indicating whether results for random effects model should be conducted.
level	The level used to calculate confidence intervals for combinations of components.
nchar.comps	A numeric defining the minimum number of characters used to create unique names for components (see Details).
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).
backtransf	A logical indicating whether printed results should be back transformed. If backtransf=TRUE, results for sm="OR" are printed as odds ratios rather than log odds ratios.
digits	Minimal number of significant digits, see print.default.
digits.stat	Minimal number of significant digits for z-value of test for overall effect, see print.default.
digits.pval	Minimal number of significant digits for p-values, see print.default.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
zero.pval	A logical specifying whether p-values should be printed with a leading zero.
JAMA.pval	A logical specifying whether p-values for test of combination effect should be printed according to JAMA reporting standards.
big.mark	A character used as thousands separator.
legend	A logical indicating whether a legend should be printed.

Details

R functions netcomb and discomb calculate effects for individual components and complex interventions present in the component network meta-analysis (CNMA). This function can be used to

calculate the effect for comparisons of two arbitrary complex interventions defined by arguments `treat1` and `treat2`.

All complex interventions occuring in the network are considered for the first complex intervention if argument `treat1` is missing. The reference group defined in the (C)NMA is used as second complex intervention if argument `treat2` is missing. The first complex intervention in the (C)NMA is used if the reference group is not defined.

The following matrices are needed to calculate comparison effects of arbitrary complex interventions, (Rücker et al., 2020, Section 3.2):

- B matrix describing how comparisons are composed by complex intervetions,
- C matrix describing how the complex interventions are composed by the components.

Internally, both matrices are constructed based on arguments `x`, `treat1` and `treat2`.

By default, component names are not abbreviated in printouts. However, in order to get more concise printouts, argument `nchar.comps` can be used to define the minimum number of characters for abbreviated component names (see `abbreviate`, argument `minlength`). R function `treats` is utilised internally to create abbreviated component names.

Value

A list is returned by the function `netcomparison` with the following elements:

`comparison` Comparison.

`TE.common`, `TE.random`
: A vector of comparison effects (common and random effects model).

`seTE.common`, `seTE.random`
: A vector with corresponding standard errors (common and random effects model).

`lower.common`, `lower.random`
: A vector with lower confidence limits for comparisons (common and random effects model).

`upper.common`, `upper.random`
: A vector with upper confidence limits for comparisons (common and random effects model).

`statistic.common`, `statistic.random`
: A vector with z-values for the overall effect of comparisons (common and random effects model).

`pval.common`, `pval.random`
: A vector with p-values for the overall effect of comparisons (common and random effects model).

`trts` Treatments included in comparisons.

`comps` Components included in comparisons.

`treat1`, `treat2` A defined above.

`common`, `random` A defined above.

`level`, `nchar.comps`, `backtransf`, `x`
: A defined above.

`B.matrix` B matrix.

`C.matrix` C matrix.

Note

R function netcomplex can be used to calculate the effect for arbitrary complex interventions in a component network meta-analysis.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Rücker G, Petropoulou M, Schwarzer G (2020): Network meta-analysis of multicomponent interventions. *Biometrical Journal*, **62**, 808–21

See Also

netcomb, discomb, netcomplex

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, ref = "placebo", sm = "OR", common = FALSE)

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")

# Result for comparison Face-to-face PST vs TCA
netcomparison(nc1, "Face-to-face PST", "TCA", nchar.comps = 4)
netcomparison(nc1, "F", "T", nchar.comps = 4)

# Result for comparison Face-to-face PST vs TCA + Placebo
netcomparison(nc1, "Face-to-face PST", "TCA + Plac", nchar.comps = 4)

## Not run:
# Artificial example
t1 <- rep("A", 3)
t2 <- c("B+C", "A+C", "C+D")
TE <- c(0, 1, 0)
seTE <- rep(1, 3)
# Conduct (C)NMA
net2 <- netmeta(TE, seTE, t1, t2, random = FALSE)
nc2 <- netcomb(net2)
```

```
# Result for comparison A vs B + D
netcomparison(nc2, "A", "B + D")
# Same results
netcomparison(nc2, "A", "B+D")
netcomparison(nc2, "A", "D+B")
netcomparison(nc2, "a", "d+b")

# Generated B matrix
netcomparison(nc2, "A", "B + D")$C.matrix
# Generated B matrix
netcomparison(nc2, "A", "B + D")$B.matrix

## End(Not run)
```

`netcomplex` *Calculate effect of arbitrary complex interventions in component network meta-analysis*

Description

Calculate effect of arbitrary complex interventions (i.e., combinations of several components) in component network meta-analysis.

Usage

```
netcomplex(
  x,
  complex,
  common = x$common,
  random = x$random,
  level = x$level.ma,
  nchar.comps = x$nchar.trts,
  warn.deprecated = gs("warn.deprecated"),
  ...
)

## S3 method for class 'netcomplex'
print(
  x,
  common = x$common,
  random = x$random,
  backtransf = x$backtransf,
  nchar.comps = x$nchar.comps,
  digits = gs("digits"),
  digits.stat = gs("digits.stat"),
  digits.pval = gs("digits.pval"),
```

```
  scientific.pval = gs("scientific.pval"),
  zero.pval = gs("zero.pval"),
  JAMA.pval = gs("JAMA.pval"),
  big.mark = gs("big.mark"),
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netcomb or netcomplex (print function).
complex	A matrix, vector or single numeric defining the complex intervention(s) (see Details).
common	A logical indicating whether results for common effects model should be conducted.
random	A logical indicating whether results for random effects model should be conducted.
level	The level used to calculate confidence intervals for combinations of components.
nchar.comps	A numeric defining the minimum number of characters used to create unique names for components (see Details).
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).
backtransf	A logical indicating whether printed results should be back transformed. If backtransf=TRUE, results for sm="OR" are printed as odds ratios rather than log odds ratios.
digits	Minimal number of significant digits, see print.default.
digits.stat	Minimal number of significant digits for z-value of test for overall effect, see print.default.
digits.pval	Minimal number of significant digits for p-values, see print.default.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
zero.pval	A logical specifying whether p-values should be printed with a leading zero.
JAMA.pval	A logical specifying whether p-values for test of combination effect should be printed according to JAMA reporting standards.
big.mark	A character used as thousands separator.
legend	A logical indicating whether a legend should be printed.

Details

R functions netcomb and discomb only report results for complex interventions present in the network. This function can be used to calculate the effect for arbitrary complex interventions.

Complex interventions can be specified using argument complex:

- a character vector with definition of complex interventions,
- a single numeric defining the number of components to combine in a complex intervention,
- a dedicated C matrix.

In order to calculate effects of arbitrary complex interventions, a C matrix is needed which describes how the complex interventions are composed by the components (Rücker et al., 2020, Section 3.2). The C matrix is constructed internally if not provided by argument complex. All complex interventions occuring in the network are considered if argument complex is missing.

By default, component names are not abbreviated in printouts. However, in order to get more concise printouts, argument nchar.comps can be used to define the minimum number of characters for abbreviated component names (see abbreviate, argument minlength). R function treats is utilised internally to create abbreviated component names.

Value

A list is returned by the function netcomplex with the following elements:

complex — Complex intervention(s).

Comb.common, Comb.random — A vector of combination effects (common and random effects model).

seComb.common, seComb.random — A vector with corresponding standard errors (common and random effects model).

lower.Comb.common, lower.Comb.random — A vector with lower confidence limits for combinations (common and random effects model).

upper.Comb.common, upper.Comb.random — A vector with upper confidence limits for combinations (common and random effects model).

statistic.Comb.common, statistic.Comb.random — A vector with z-values for the overall effect of combinations (common and random effects model).

pval.Comb.common, pval.Comb.random — A vector with p-values for the overall effect of combinations (common and random effects model).

common, random — A defined above.

level, nchar.comps, backtransf, x — A defined above.

C.matrix — C matrix.

Note

R function netcomparison can be used to calculate the effect for comparisons of two arbitrary complex intervention in a component network meta-analysis.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Rücker G, Petropoulou M, Schwarzer G (2020): Network meta-analysis of multicomponent interventions. *Biometrical Journal*, **62**, 808–21

See Also

netcomb, discomb, netcomparison

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, ref = "placebo", sm = "OR", common = FALSE)

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")

# Result for combination Face-to-face PST + SSRI
netcomplex(nc1, "Face-to-face PST + SSRI", nchar.comps = 4)
netcomplex(nc1, "F + S", nchar.comps = 4)

# Result for combination Face-to-face PST + SSRI + Placebo
netcomplex(nc1, "Face-to-face PST + SSRI + Plac", nchar.comps = 4)

## Not run:
# Artificial example
t1 <- rep("A", 3)
t2 <- c("B+C", "A+C", "C+D")
TE <- c(0, 1, 0)
seTE <- rep(1, 3)
# Conduct (C)NMA
net2 <- netmeta(TE, seTE, t1, t2, random = FALSE)
nc2 <- netcomb(net2)

# Result for combination A + B + C
netcomplex(nc2, "A + B + C")
# Same results
netcomplex(nc2, "A+B+C")
```

```
netcomplex(nc2, "B+C+A")
netcomplex(nc2, "C+B+A")
netcomplex(nc2, "c+b+a")

# Generated C matrix
netcomplex(nc2, c(LETTERS[1:4], "A+B+C"))$C.matrix

# Results for all possible combinations of two components
netcomplex(nc2, 2)
# Results for all possible combinations of three components
netcomplex(nc2, 3)

## End(Not run)
```

`netconnection` *Get information on network connectivity (number of subnetworks, distance matrix)*

Description

To determine the network structure and to test whether a given network is fully connected. Network information is provided as a triple of vectors `treat1`, `treat2`, and `studlab` where each row corresponds to an existing pairwise treatment comparison (`treat1`, `treat2`) in a study (`studlab`). The function calculates the number of subnetworks (connectivity components; value of 1 corresponds to a fully connected network) and the distance matrix (in block-diagonal form in the case of subnetworks). If some treatments are combinations of other treatments or have common components, an analysis based on the additive network meta-analysis model might be possible, see discomb function.

Usage

```
netconnection(
  treat1,
  treat2,
  studlab,
  data = NULL,
  subset = NULL,
  sep.trts = ":",
  nchar.trts = 666,
  title = "",
  details.disconnected = FALSE,
  warn = FALSE
)

## S3 method for class 'netconnection'
print(
  x,
```

```
  digits = max(4, .Options$digits - 3),
  nchar.trts = x$nchar.trts,
  details = FALSE,
  details.disconnected = x$details.disconnected,
  ...
)
```

Arguments

treat1	Label / number for first treatment or a data frame created with pairwise.
treat2	Label / number for second treatment.
studlab	An optional - but important! - vector with study labels (see Details).
data	An optional data frame containing the study information.
subset	An optional vector specifying a subset of studies to be used.
sep.trts	A character used in comparison names as separator between treatment labels.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
title	Title of meta-analysis / systematic review.
details.disconnected	A logical indicating whether to print more details for disconnected networks.
warn	A logical indicating whether warnings should be printed.
x	An object of class netconnection.
digits	Minimal number of significant digits, see print.default.
details	A logical indicating whether to print the distance matrix.
...	Additional arguments (ignored at the moment)

Value

An object of class netconnection with corresponding print function. The object is a list containing the following components:

treat1, treat2, studlab, title, warn, nchar.trts	As defined above.
k	Total number of studies.
m	Total number of pairwise comparisons.
n	Total number of treatments.
n.subnets	Number of subnetworks; equal to 1 for a fully connected network.
D.matrix	Distance matrix.
A.matrix	Adjacency matrix.
L.matrix	Laplace matrix.
call	Function call.
version	Version of R package netmeta used to create object.

Author(s)

Gerta Rücker <ruecker@imbi.uni-freiburg.de>, Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netmeta, netdistance, discomb

Examples

```
data(Senn2013)

nc1 <- netconnection(treat1, treat2, studlab, data = Senn2013)
nc1

# Extract number of (sub)networks
#
nc1$n.subnets

# Extract distance matrix
#
nc1$D.matrix

## Not run:
# Conduct network meta-analysis (results not shown)
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab, data = Senn2013)

# Artificial example with two subnetworks
#
t1 <- c("G", "B", "B", "D", "A", "F")
t2 <- c("B", "C", "E", "E", "H", "A")
#
nc2 <- netconnection(t1, t2)
print(nc2, details = TRUE)

# Number of subnetworks
#
nc2$n.subnets

# Extract distance matrix
#
nc2$D.matrix

# Conduct network meta-analysis (results in an error message due to
# unconnected network)
try(net2 <- netmeta(1:6, 1:6, t1, t2, 1:6))

# Conduct network meta-analysis on first subnetwork
#
net2.1 <- netmeta(1:6, 1:6, t1, t2, 1:6, subset = nc2$subnet == 1)

# Conduct network meta-analysis on second subnetwork
```

```
#
net2.2 <- netmeta(1:6, 1:6, t1, t2, 1:6, subset = nc2$subnet == 2)

net2.1
net2.2

## End(Not run)
```

---

netcontrib *Contribution matrix in network meta-analysis*

Description

This function generates the contribution of direct comparisons to every network treatment comparison. The output is a matrix where rows represent network treatment effects and columns represent the contribution of direct treatment effects.

Usage

```
netcontrib(
  x,
  method = "shortestpath",
  hatmatrix.F1000 = FALSE,
  common = x$common,
  random = x$random,
  nchar.trts = x$nchar.trts,
  warn.deprecated = gs("warn.deprecated"),
  verbose = FALSE,
  ...
)

## S3 method for class 'netcontrib'
print(
  x,
  common = x$x$common,
  random = x$x$random,
  digits = 4,
  nchar.trts = x$nchar.trts,
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netmeta or netcontrib.

method	A character string indicating which method is to calculate the contribution matrix. Either "randomwalk" or "shortestpath", can be abbreviated.
hatmatrix.F1000	A logical indicating whether hat matrix given in F1000 article should be used for method = "shortestpath".
common	A logical indicating whether a contribution matrix should be printed for the common effects network meta-analysis.
random	A logical indicating whether a contribution matrix should be printed for the random effects network meta-analysis.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names (see Details).
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
verbose	A logical indicating whether progress information should be printed.
...	Additional arguments.
digits	Minimal number of significant digits, see print.default.
legend	A logical indicating whether a legend should be printed.

Details

In network meta-analysis (NMA), it is important to assess the influence of limitations or other characteristics of individual studies on the estimates obtained from the network. To this end, the contribution matrix shows how much each direct treatment effect contributes to each treatment effect estimate from network meta-analysis.

We use ideas from graph theory to derive the proportion that is contributed by each direct treatment effect. We start with the 'projection' matrix in a two-step network meta-analysis model, called the H matrix, which is analogous to the hat matrix in a linear regression model. H entries are translated to proportion contributions based on the observation that the rows of H can be interpreted as flow networks. A stream is defined as the composition of a path and its associated flow (Papakonstantinou et al., 2018).

To account for multi-arm trials, we use the H matrix from a two-step (aggregate) version of the graph theoretical NMA model (Davies et al., 2021). This H matrix can be obtained from hatmatrix with argument method = "davies".

Two methods are implemented to estimate the streams and as a result, the proportion contributions:

(1) If argument method = "randomwalk", an analytical random-walk (RW) approach is used (Davies et al., 2021). Here, the "full" version of the aggregate H matrix (hatmatrix with arguments method = "davies" and type = "full") is used to define RW transition matrices. For each pair of treatments (ij) in the network, the elements in the corresponding row of H-full define a transition matrix from node i to node j. We use the **igraph** package to find every (directed) path from node i to node j. The flow through each path is then equal to the probability that a walker takes that path. This is simply the product of the transition probabilities associated with each edge along the path.

(2) If argument method = "shortestpath", an iterative algorithm is used (Papakonstantinou et al., 2018). Broadly speaking, each iteration of the algorithm consists of the following steps: (i) A path in the evidence flow network is selected. (ii) The minimum flow through the edges making up the

path is identified. This is assigned as the flow associated with the path. (iii) The flow of the path is subtracted from the values of flow in the edges that make up that path. This means that the edge corresponding to the minimum flow in that path is removed from the graph. (iv) A new path is then selected from the remaining graph. The process repeats until all the evidence flow in the edges has been assigned to a path.

In the original F1000 paper (Papakonstantinou et al., 2018), the hat matrix used did not account for correlations due to multi-arm trials. For reproducibility the result of this version can be obtained by specifying `hatmatrix.F1000 = TRUE` for `method = "shortestpath"`. For other purposes, this method is not recommended.

Once the streams have been identified (either by method (1) or (2)), the proportion contribution of each direct comparison is equal to the sum over the flow of evidence in each path containing that edge divided by the number of edges that make up that path.

By default, treatment names are not abbreviated in printouts. However, in order to get more concise printouts, argument `nchar.trts` can be used to define the minimum number of characters for abbreviated treatment names (see `abbreviate`, argument `minlength`). R function `treats` is utilised internally to create abbreviated treatment names.

Calculation of network contributions can be compute-intensive for the random-walk approach in large networks. Crude information on the computation progress is printed if argument `verbose` is `TRUE`. In addition, computation times are printed if R package **tictoc** is installed.

Value

An object of class `netcontrib` with corresponding `print` function. The object is a list containing the following components:

`common`	Numeric matrix of percentage contributions of direct comparisons for each network comparison for the common effects model.
`random`	Numeric matrix of percentage contributions of direct comparisons for each network comparison for the random effects model.
`x`	As defined above.
`tictoc.common`	Computation times under common effects model (if R package **tictoc** is installed).
`tictoc.random`	Computation times under random effects model (if R package **tictoc** is installed).

with the contribution matrices for common and random NMA. Each matrix has the percentage contributions of each direct comparison as columns for each network comparison, direct or indirect as rows.

Author(s)

Theodoros Papakonstantinou <dev@tpapak.com>, Annabel Davies <annabel.davies@manchester.ac.uk>

References

Davies AL, Papakonstantinou T, Nikolakopoulou A, Rücker G, Galla T (2021): Network meta-analysis and random walks. Available from: http://arxiv.org/abs/2107.02886

Papakonstantinou, T., Nikolakopoulou, A., Rücker, G., Chaimani, A., Schwarzer, G., Egger, M., Salanti, G. (2018): Estimating the contribution of studies in network meta-analysis: paths, flows and streams. *F1000Research*

See Also

netmeta

Examples

```
# Use the Woods dataset
#
data("Woods2010")
p1 <- pairwise(treatment, event = r, n = N,
  studlab = author, data = Woods2010, sm = "OR")

net1 <- netmeta(p1)
cm <- netcontrib(net1)
cm

netcontrib(net1, method = "r")
```

---

netdistance *Calculate distance matrix for an adjacency matrix*

Description

Calculate distance matrix for an adjacency matrix based on distance algorithm by Müller et al. (1987).

Usage

```
netdistance(x)
```

Arguments

x	Either a netmeta object or an adjacency matrix.

Author(s)

Gerta Rücker <ruecker@imbi.uni-freiburg.de>

References

Müller WR, Szymanski K, Knop JV, and Trinajstic N (1987): An algorithm for construction of the molecular distance matrix. *Journal of Computational Chemistry*, **8**, 170–73

See Also

netmeta, netconnection

Examples

```
data(Senn2013)

net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD")

netdistance(net1)
netdistance(net1$A.matrix)
```

---

netgraph *Generic function for network graphs*

Description

Generic function for network graphs

Usage

```
netgraph(x, ...)
```

Arguments

x	An R object.
...	Additional arguments.

Details

For more details, look at the following functions to generate network graphs:

- netgraph.netmeta
- netgraph.netimpact
- netgraph.netconnection
- netgraph.netcomb
- netgraph.discomb

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de >

Examples

```
data(Senn2013)

# Generation of an object of class 'netmeta' with reference
# treatment 'plac'
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", reference = "plac")

# Network graph with default settings
#
netgraph(net1)

## Not run:
data(Woods2010)
p1 <- pairwise(treatment, event = r, n = N,
  studlab = author, data = Woods2010, sm = "OR")
net2 <- netmeta(p1)

# Network graph with default settings
#
netgraph(net2)

# Network graph with
# - number of studies for each pairwise comparison and
# - number of participants for each treatment arm
#
netgraph(net2, number.of.studies = TRUE,
  labels = paste0(trts, " (n=", n.trts, ")"))

## End(Not run)
```

---

netgraph.discomb *Network graph for objects of class discomb*

Description

This function generates a graph of the evidence network.

Usage

```
## S3 method for class 'discomb'
netgraph(x, plastic = FALSE, ...)
```

Arguments

x	An object of class discomb.

plastic	A logical indicating whether the appearance of the comparisons should be in '3D look'.
...	Additional arguments passed on to netgraph.netmeta (see Details).

Details

The arguments seq and iterate are used internally and cannot be specified by the user.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Gerta Rücker <ruecker@imbi.uni-freiburg.de>

See Also

discomb, netgraph.netmeta

Examples

```
# Artificial dataset
#
t1 <- c("A + B", "A + C", "A"    , "A"    , "D", "D", "E")
t2 <- c("C"    , "B"    , "B + C", "A + D", "E", "F", "F")
#
mean <- c(4.1, 2.05, 0, 0, 0.1, 0.1, 0.05)
se.mean <- rep(0.1, 7)
#
study <- paste("study", c(1:4, 5, 5, 5))
#
dat <- data.frame(mean, se.mean, t1, t2, study,
                  stringsAsFactors = FALSE)
#
trts <- c("A", "A + B", "A + C", "A + D",
  "B", "B + C", "C", "D", "E", "F")
#
comps <- LETTERS[1:6]

# Use netconnection() to display network information
#
netconnection(t1, t2, study)

dc1 <- discomb(mean, se.mean, t1, t2, study, seq = trts)

netgraph(dc1)
```

`netgraph.netcomb` *Network graph for objects of class netcomb*

Description

This function generates a graph of the evidence network.

Usage

```
## S3 method for class 'netcomb'
netgraph(x, ...)
```

Arguments

`x`	An object of class `netcomb`.
`...`	Additional arguments passed on to `netgraph.netmeta`.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Gerta Rücker <ruecker@imbi.uni-freiburg.de>

See Also

`netcomb`, `netgraph.netmeta`

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, ref = "placebo", sm = "OR", common = FALSE)

# Additive model for treatment components (with placebo as inactive
# treatment)
#
nc1 <- netcomb(net1, inactive = "placebo")

netgraph(nc1)
```

netgraph.netconnection

Network graph for objects of class netconnection

Description

This function generates a graph of the evidence network.

Usage

```
## S3 method for class 'netconnection'
netgraph(
  x,
  seq,
  col = seq_len(x$n.subnets),
  reference.group = NULL,
  plastic = FALSE,
  ...
)
```

Arguments

x	An object of class netconnection.
seq	A character or numerical vector specifying the sequence of treatments arrangement (anticlockwise if start.layout = "circle").
col	A single color (or vector of colors) for lines connecting treatments (edges) if argument plastic = FALSE (see Details).
reference.group	Reference treatment (only relevant for disconnected networks).
plastic	A logical indicating whether the appearance of the comparisons should be in '3D look'.
...	Additional arguments passed on to netgraph.netmeta (see Details).

Details

Argument col can be a single color for all edges, a vector of length equal to the number of edges, or a vector of length equal to the number of subnetworks. Argument reference.group is only considered in disconnected networks, i.e., if more than one (sub)network exists, and if argument col provides colors for subnetworks. In this case, the first color provided in argument col defines the color for the subnetwork with the reference treatment.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Gerta Rücker <ruecker@imbi.uni-freiburg.de>

See Also

netconnection, netgraph.netmeta

Examples

```
# Artificial example with two subnetworks
#
t1 <- c("G", "B", "B", "D", "A", "F")
t2 <- c("B", "C", "E", "E", "H", "A")
#
nc1 <- netconnection(t1, t2)
print(nc1, details = TRUE)

netgraph(nc1, points = TRUE, adj = 0.5, bg.points = "lightgray")
netgraph(nc1, points = TRUE, adj = 0.5, bg.points = "lightgray",
  plastic = TRUE)
```

---

netgraph.netimpact *Network graph for objects of class netimpact*

Description

This function generates a graph of the evidence network.

Usage

```
## S3 method for class 'netimpact'
netgraph(
  x,
  col.ignore = "red",
  number.of.studies = TRUE,
  main,
  sub,
  multiarm = FALSE,
  col.multiarm = NULL,
  alpha.transparency = 0.5,
  col.ignore.multiarm = "transparent",
  col = "black",
  plastic = FALSE,
  ...
)
```

Arguments

x	An object of class netimpact.
col.ignore	A character string indicating color for comparisons removed from network, either "transparent" or any color defined in colours.

number.of.studies
: A logical indicating whether number of studies should be added to network graph.

main
: Main title.

sub
: Subtitle.

multiarm
: A logical indicating whether multi-arm studies should be marked in plot.

col.multiarm
: Either a function from R package colorspace or grDevice to define colors for multi-arm studies or a character vector with colors to highlight multi-arm studies.

alpha.transparency
: The alpha transparency of colors used to highlight multi-arm studies (0 means transparent and 1 means opaque).

col.ignore.multiarm
: A character string indicating color to mark multi-arm studies removed from network, either "transparent" or any color defined in colours.

col
: A single color (or vector of colors) for lines connecting treatments (edges) if argument plastic = FALSE. Length of the vector must be equal to the number of edges.

plastic
: A logical indicating whether the appearance of the comparisons should be in '3D look'.

...
: Additional arguments passed on to netgraph.netmeta.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Gerta Rücker <ruecker@imbi.uni-freiburg.de>

See Also

netimpact, netgraph.netmeta

Examples

```
data(Franchini2012)

# Only consider first two studies (to reduce runtime of example)
#
studies <- unique(Franchini2012$Study)
p1 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = subset(Franchini2012, Study %in% studies[1:2]),
  studlab = Study)

net1 <- netmeta(p1)
ni1 <- netimpact(net1, verbose = TRUE)
netgraph(ni1)
netgraph(ni1, plastic = TRUE)
```

```
## Not run:
p2 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = Franchini2012,
  studlab = Study)

net2 <- netmeta(p2)
ni2 <- netimpact(net2, verbose = TRUE)
netgraph(ni2)
netgraph(ni2, plastic = TRUE)

## End(Not run)
```

`netgraph.netmeta` *Network graph*

Description

This function generates a graph of the evidence network.

Usage

```
## S3 method for class 'netmeta'
netgraph(
  x,
  seq = x$seq,
  labels = x$trts,
  cex = 1,
  adj = NULL,
  srt.labels = 0,
  offset = if (!is.null(adj) && all(unique(adj) == 0.5)) 0 else 0.02,
  scale = 1.1,
  col = if (iterate) "slateblue" else "black",
  plastic = !(iterate & allfigures),
  thickness = "number.of.studies",
  lwd = 5,
  lwd.min = lwd/2.5,
  lwd.max = lwd * 4,
  rescale.thickness = if (thickness == "number.of.studies") sqrt else I,
  dim = "2d",
  rotate = 0,
  highlight = NULL,
  col.highlight = "red2",
  scale.highlight = 1,
  multiarm = FALSE,
```

```
  col.multiarm = NULL,
  alpha.transparency = 0.5,
  points = !missing(cex.points),
  cex.points = 4,
  pch.points = 21,
  col.points = "black",
  bg.points = "red",
  points.max = if (length(unique(cex.points)) != 1) 8 else cex.points,
  points.min = if (length(unique(cex.points)) != 1) 1 else cex.points,
  rescale.pointsize = sqrt,
  number.of.studies = FALSE,
  cex.number.of.studies = cex,
  col.number.of.studies = "white",
  bg.number.of.studies = "black",
  pos.number.of.studies = 0.5,
  start.layout = ifelse(dim == "2d", "circle", "eigen"),
  eig1 = 2,
  eig2 = 3,
  eig3 = 4,
  iterate = FALSE,
  tol = 1e-04,
  maxit = 500,
  allfigures = FALSE,
  A.matrix = x$A.matrix,
  N.matrix = sign(A.matrix),
  D.matrix = netdistance(N.matrix),
  xpos = NULL,
  ypos = NULL,
  zpos = NULL,
  figure = TRUE,
  ...
)
```

Arguments

x	An object of class netmeta (mandatory).
seq	A character or numerical vector specifying the sequence of treatments arrangement (anticlockwise if start.layout = "circle").
labels	An optional vector with treatment labels.
cex	The magnification to be used for treatment labels.
adj	One, two, or three values in [0, 1] (or a vector / matrix with length / number of rows equal to the number of treatments) specifying the x (and optionally y and z) adjustment for treatment labels.
srt.labels	The character string "orthogonal" (can be abbreviated), a single numeric or numerical vector with value(s) between -180 and 180 specifying the angle to rotate treatment labels (see Details).

`offset` Distance between edges (i.e. treatments) in graph and treatment labels for 2-D plots (value of 0.0175 corresponds to a difference of 1.75% of the range on x- and y-axis).

`scale` Additional space added outside of edges (i.e. treatments). Increase this value for larger treatment labels (value of 1.10 corresponds to an additional space of 10% around the network graph).

`col` A single color (or vector of colors) for lines connecting treatments (edges) if argument `plastic = FALSE`. Length of the vector must be equal to the number of edges (see list element 'comparisons' in `netmeta`).

`plastic` A logical indicating whether the appearance of the comparisons should be in '3D look' (not to be confused with argument `dim`).

`thickness` Either a character variable to determine the method to plot line widths (see Details) or a matrix of the same dimension and row and column names as argument `A.matrix` with information on line width.

`lwd` A numeric for scaling the line width of comparisons.

`lwd.min` Minimum line width in network graph. All connections with line widths below this values will be set to `lwd.min`.

`lwd.max` Maximum line width in network graph. The connection with the largest value according to argument `thickness` will be set to this value.

`rescale.thickness`
R function to scale the thickness of lines.

`dim` A character string indicating whether a 2- or 3-dimensional plot should be produced, either `"2d"` or `"3d"`.

`rotate` A single numeric with value between -180 and 180 specifying the angle to rotate nodes in a circular network.

`highlight` A character vector identifying comparisons that should be marked in the network graph, e.g. `highlight = "treat1:treat2"`.

`col.highlight` Color(s) to highlight the comparisons given by `highlight`.

`scale.highlight`
Scaling factor(s) for the line width(s) to highlight the comparisons given by `highlight`.

`multiarm` A logical indicating whether multi-arm studies should be marked in plot.

`col.multiarm` Either a function from R package colorspace or grDevice to define colors for multi-arm studies or a character vector with colors to highlight multi-arm studies.

`alpha.transparency`
The alpha transparency of colors used to highlight multi-arm studies (0 means transparent and 1 means opaque).

`points` A logical indicating whether points should be printed at nodes (i.e. treatments) of the network graph.

`cex.points`, `pch.points`, `col.points`, `bg.points`
Corresponding color, background color, size, and type for points. Can be a vector with length equal to the number of treatments.

`points.max` Maximum point size in network graph. The node with the largest value according to argument `cex.points` will be set to this value.

`points.min` Minimum point size. All points with size below this values will be set to `points.min`.

`rescale.pointsize` R function to scale the point size.

`number.of.studies` A logical indicating whether number of studies should be added to network graph.

`cex.number.of.studies` The magnification to be used for number of studies.

`col.number.of.studies` Color for number of studies.

`bg.number.of.studies` Color for shadow around number of studies.

`pos.number.of.studies` A single value (or vector of values) in [0, 1] specifying the position of the number of studies on the lines connecting treatments (edges). Length of the vector must be equal to the number of edges.

`start.layout` A character string indicating which starting layout is used if `iterate` = `TRUE`. If "circle" (default), the iteration starts with a circular ordering of the vertices; if "eigen", eigenvectors of the Laplacian matrix are used, calculated via generic function `eigen` (spectral decomposition); if "prcomp", eigenvectors of the Laplacian matrix are calculated via generic function `prcomp` (principal component analysis); if "random", a random layout is used, drawn from a bivariate normal.

`eig1` A numeric indicating which eigenvector is used as x coordinate if `start` = `"eigen"` or `"prcomp"` and `iterate` = `TRUE`. Default is 2, the eigenvector to the second-smallest eigenvalue of the Laplacian matrix.

`eig2` A numeric indicating which eigenvector is used as y-coordinate if `start` = `"eigen"` or `"prcomp"` and `iterate` = `TRUE`. Default is 3, the eigenvector to the third-smallest eigenvalue of the Laplacian matrix.

`eig3` A numeric indicating which eigenvector is used as z-coordinate if `start` = `"eigen"` or `"prcomp"` and `iterate` = `TRUE`. Default is 4, the eigenvector to the fourth-smallest eigenvalue of the Laplacian matrix.

`iterate` A logical indicating whether the stress majorization algorithm is carried out for optimization of the layout.

`tol` A numeric for the tolerance for convergence if `iterate` = `TRUE`.

`maxit` An integer defining the maximum number of iteration steps if `iterate` = `TRUE`.

`allfigures` A logical indicating whether all iteration steps are shown if `iterate` = `TRUE`. May slow down computations if set to `TRUE` (especially if `plastic` = `TRUE`).

`A.matrix` Adjacency matrix (*n*x*n*) characterizing the structure of the network graph. Row and column names must be the same set of values as provided by argument `seq`.

`N.matrix` Neighborhood matrix (*n*x*n*) replacing A.matrix if neighborhood is to be specified differently from node adjacency in the network graph, for example content-based. Row and column names must be the same set of values as provided by argument `seq`.

D.matrix	Distance matrix (nxn) replacing A.matrix and N.matrix if distances should be provided directly. Row and column names must be the same set of values as provided by argument seq.
xpos	Vector (n) of x coordinates.
ypos	Vector (n) of y coordinates.
zpos	Vector (n) of z coordinates.
figure	A logical indicating whether network graph should be shown.
...	Additional graphical arguments.

Details

The network is laid out in the plane, where the nodes in the graph layout correspond to the treatments and edges display the observed treatment comparisons. For the default setting, nodes are placed on a circle. Other starting layouts are "eigen", "prcomp", and "random" (Rücker & Schwarzer 2015). If iterate = TRUE, the layout is further optimized using the stress majorization algorithm. This algorithm specifies an 'ideal' distance (e.g., the graph distance) between two nodes in the plane. In the optimal layout, these distances are best approximated in the sense of least squares. Starting from an initial layout, the optimum is approximated in an iterative process called stress majorization (Kamada and Kawai 1989, Michailidis and de Leeuw 2001, Hu 2012). The starting layout can be chosen as a circle or coming from eigenvectors of the Laplacian matrix (corresponding to Hall's algorithm, Hall 1970), calculated in different ways, or random. Moreover, it can be chosen whether the iteration steps are shown (argument allfigures = TRUE).

An optimized circular presentation which typically has a reduced (sometimes minimal) number of crossings can be achieved by using argument seq = "optimal" in combination with argument start.layout. Note, is is not possible of prespecify the best value for argument start.layout for any situation as the result depends on the network structure.

Argument thickness providing the line width of the nodes (comparisons) can be a matrix of the same dimension as argument A.matrix or any of the following character variables:

- Proportional to number of studies comparing two treatments (thickness = "number.of.studies", default)
- Proportional to inverse standard error of common effects model comparing two treatments (thickness = "se.common")
- Proportional to inverse standard error of random effects model comparing two treatments (thickness = "se.random")
- Weight from common effects model comparing two treatments (thickness = "w.common")
- Weight from random effects model comparing two treatments (thickness = "w.random")
- Same line width (argument lwd) for all comparisons (thickness = "equal")

Only evidence from direct treatment comparisons is considered to determine the line width if argument thickness is equal to any but the last method.

Argument srt.labels can be used to specific the rotation (in degrees) of the treatment labels. If srt.labels is equal to "orthogonal", treatment labels are orthogonal to the circle. If srt.labels is a single numeric, all labels are rotated by this degree. If srt.labels is a numeric vector, it must be of the same length as the number of treatments and labels are rotated counter-clockwise starting

on the right side. Finally, if `srt.labels` is a named numeric vector, it must be of the same length as the number of treatments and the names must be equal to the treatment names (and treatment labels are rotated according to the specified values).

Further, a couple of graphical parameters can be specified, such as color and appearance of the edges (treatments) and the nodes (comparisons), whether special comparisons should be highlighted and whether multi-arm studies should be indicated as colored polygons. By default, if R package colorspace is available the `sequential_hcl` function is used to highlight multi-arm studies; otherwise the `rainbow` is used.

In order to generate 3-D plots (argument `dim = "3d"`), R package **rgl** is necessary. Note, under macOS the X.Org X Window System must be available (see `https://www.xquartz.org`).

Value

A list containing two data frames with information on nodes and edges.

List element 'nodes'

`trts`	Treatment names.
`labels`	Treatment labels.
`seq`	Sequence of treatment labels.
`srt`	String rotation.
`xpos`	Position of treatment / edge on x-axis.
`ypos`	Position of treatment / edge on y-axis.
`zpos`	Position of treatment / edge on z-axis (for 3-D plots).
`xpos.labels`	Position of treatment labels on x-axis (for 2-D plots).
`ypos.labels`	Position of treatment labels on y-axis (for 2-D plots).
`offset.x`	Offset of treatment labels on x-axis (for 2-D plots).
`offset.y`	Offset of treatment labels on y-axis (for 2-D plots).
`cex`	Point size of treatments / edges.
`col`	Color for points.
`pch`	Point type.
`bg`	Background color for points.
`adj.x`	Adjustment for treatment label on x-axis.
`adj.y`	Adjustment for treatment label on y-axis.
`adj.z`	Adjustment for treatment label on z-axis (for 3-D plots).

List element 'edges'

`treat1`	Name of first treatment.
`treat2`	Name of second treatment.
`n.stud`	Number of studies directly comparing treatments.
`xpos`	Position of number of studies on x-axis.
`ypos`	Position of number of studies on y-axis.

`adj` Adjustment of number of studies.

`pos.number.of.studies` Position of number of studies on edge.

`col` Color for edges.

Author(s)

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See Also

`netmeta`

Examples

```
data(Senn2013)

# Generation of an object of class 'netmeta' with reference
# treatment 'plac'
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", reference = "plac")

# Network graph with default settings
#
netgraph(net1)

## Not run:
# Network graph with specified order of the treatments and one
# highlighted comparison
#
trts <- c("plac", "benf", "migl", "acar", "sulf",
  "metf", "rosi", "piog", "sita", "vild")
```

```
netgraph(net1, highlight = "rosi:plac", seq = trts)

# Same network graph using argument 'seq' in netmeta function
#
net2 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", reference = "plac", seq = trts)
netgraph(net2, highlight = "rosi:plac")

# Network graph optimized, starting from a circle, with multi-arm
# study colored
#
netgraph(net1, start = "circle", iterate = TRUE,
  multiarm = TRUE, col.multiarm = "purple")

# Network graph optimized, starting from a circle, with multi-arm
# study colored and all intermediate iteration steps visible
#
netgraph(net1, start = "circle", iterate = TRUE,
  multiarm = TRUE, col.multiarm = "purple",
  allfigures = TRUE)

# Network graph optimized, starting from Laplacian eigenvectors,
# with multi-arm study colored
#
netgraph(net1, start = "eigen",
  multiarm = TRUE, col.multiarm = "purple")

# Network graph optimized, starting from different Laplacian
# eigenvectors, with multi-arm study colored
#
netgraph(net1, start = "prcomp",
  multiarm = TRUE, col.multiarm = "purple")

# Network graph optimized, starting from random initial layout,
# with multi-arm study colored
#
netgraph(net1, start = "random",
  multiarm = TRUE, col.multiarm = "purple")

# Network graph without plastic look and one highlighted comparison
#
netgraph(net1, plastic = FALSE, highlight = "rosi:plac")

# Network graph with same thickness for all comparisons
#
netgraph(net1, thickness = "equal")

# Network graph with changed labels and specified order of the
# treatments
#
netgraph(net1, seq = c(1, 3, 5, 2, 9, 4, 7, 6, 8, 10),
  labels = LETTERS[1:10])
```

```
# Rotate treatment labels (orthogonal to circle)
#
netgraph(net1, srt.labels = "o")

# Network graph in 3-D (opens a new device, where you may rotate and
# zoom the plot using the mouse / the mouse wheel).
# The rgl package must be installed for 3-D plots.
#
netgraph(net1, dim = "3d")

## End(Not run)
```

`netheat` *Net heat plot*

Description

This function creates a net heat plot, a graphical tool for locating inconsistency in network meta-analyses.

Usage

```
netheat(
  x,
  random = FALSE,
  tau.preset = NULL,
  showall = TRUE,
  nchar.trts = x$nchar.trts,
  ...
)
```

Arguments

`x`	An object of class `netmeta`.
`random`	A logical indicating whether the net heat plot should be based on a random effects model.
`tau.preset`	An optional value for the square-root of the between-study variance τ^2 for a random effects model on which the net heat plot will be based.
`showall`	A logical indicating whether results should be shown for all designs or only a sensible subset (see Details).
`nchar.trts`	A numeric defining the minimum number of characters used to create unique treatment names.
`...`	Additional arguments.

Details

The net heat plot is a matrix visualization proposed by Krahn et al. (2013) that highlights hot spots of inconsistency between specific direct evidence in the whole network and renders transparent possible drivers.

In this plot, the area of a gray square displays the contribution of the direct estimate of one design in the column to a network estimate in a row. In combination, the colors show the detailed change in inconsistency when relaxing the assumption of consistency for the effects of single designs. The colors on the diagonal represent the inconsistency contribution of the corresponding design. The colors on the off-diagonal are associated with the change in inconsistency between direct and indirect evidence in a network estimate in the row after relaxing the consistency assumption for the effect of one design in the column. Cool colors indicate an increase and warm colors a decrease: the stronger the intensity of the color, the greater the difference between the inconsistency before and after the detachment. So, a blue colored element indicates that the evidence of the design in the column supports the evidence in the row. A clustering procedure is applied to the heat matrix in order to find warm colored hot spots of inconsistency. In the case that the colors of a column corresponding to design d are identical to the colors on the diagonal, the detaching of the effect of design d dissolves the total inconsistency in the network.

The pairwise contrasts corresponding to designs of three- or multi-arm studies are marked by '_' following the treatments of the design.

Designs where only one treatment is involved in other designs of the network or where the removal of corresponding studies would lead to a splitting of the network do not contribute to the inconsistency assessment. By default (`showall = TRUE`), these designs are not incorporated into the net heat plot. If `showall = FALSE`, additional designs with minimal contribution to the inconsistency Q statistic are not incorporated (i.e., designs with `abs(Q.inc.design) <= .Machine$double.eps^0.5`).

In the case of `random = TRUE`, the net heat plot is based on a random effects model generalised for multivariate meta-analysis in which the between-study variance τ^2 is estimated by the method of moments (see Jackson et al., 2012) and embedded in a full design-by-treatment interaction model (see Higgins et al., 2012).

Author(s)

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References

Krahn U, Binder H, König J (2013): A graphical tool for locating inconsistency in network meta-analyses. *BMC Medical Research Methodology*, **13**, 35

Jackson D, White IR and Riley RD (2012): Quantifying the impact of between-study heterogeneity in multivariate meta-analyses. *Statistics in Medicine*, **31**, 3805–20

Higgins JPT, Jackson D, Barrett JK, Lu G, Ades AE, White IR (2012): Consistency and inconsistency in network meta-analysis: concepts and models for multi-arm studies. *Research Synthesis Methods*, **3**, 98–110

See Also

netmeta

Examples

```
data(Senn2013)

# Only consider first five studies (to reduce runtime of example)
#
studies <- unique(Senn2013$studlab)
Senn2013.5 <- subset(Senn2013, studlab %in% studies[1:5])

# Conduct network meta-analysis with placebo as reference treatment
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013.5, sm = "MD", reference = "plac")

# Generate a net heat plot based on a common effects model
#
netheat(net1)

## Not run:
# Generate a net heat plot based on a random effects model
#
netheat(net1, random = TRUE)

## End(Not run)
```

---

netimpact *Determine the importance of individual studies in network meta-analysis*

Description

This function measures the importance of individual studies in network meta-analysis by the reduction of the precision if the study is removed / ignored from the network.

Usage

```
netimpact(
  x,
  seTE.ignore = 100 * max(x$seTE, na.rm = TRUE),
  event.ignore = 0.01,
  verbose = FALSE
)
```

Arguments

x	An object of class netmeta.
seTE.ignore	Assumed (large) standard error in order to mimicking the removal of individual studies from the network meta-analysis (ignored for netmetabin objects).

event.ignore — Assumed event number mimicking the removal of individual studies from the network meta-analysis (considered for netmetabin objects).

verbose — A logical indicating whether information on the estimation progress should be printed.

Value

An object of class "netimpact" with corresponding netgraph and print function. The object is a list containing the following components:

impact.common — A matrix with contributions of individual studies (columns) to comparisons (rows) under the common effects model.

impact.random — A matrix with contributions of individual studies (columns) to comparisons (rows) under the random effects model.

ignored.comparisons — List with comparisons of ignored study.

seTE.ignore, event.ignore, x — As defined above.

nets — List of all network meta-analyses (removing a single study).

version — Version of R package netmeta used to create object.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Gerta Rücker <ruecker@imbi.uni-freiburg.de>

See Also

netmeta, netmetabin, netgraph.netimpact, print.netimpact

Examples

```
data(Franchini2012)

# Only consider first two studies (to reduce runtime of example)
#
studies <- unique(Franchini2012$Study)
p1 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = subset(Franchini2012, Study %in% studies[1:2]),
  studlab = Study)

net1 <- netmeta(p1)
ni1 <- netimpact(net1, verbose = TRUE)
ni1

netgraph(ni1)
```

netleague *Create league table with network meta-analysis results*

Description

A league table is a square matrix showing all pairwise comparisons in a network meta-analysis. Typically, both treatment estimates and confidence intervals are shown.

Usage

```
netleague(
  x,
  y,
  common = x$common,
  random = x$random,
  seq = x$seq,
  ci = TRUE,
  backtransf = TRUE,
  direct = FALSE,
  digits = gs("digits"),
  big.mark = gs("big.mark"),
  text.NA = ".",
  bracket = gs("CIbracket"),
  separator = gs("CIseparator"),
  lower.blank = gs("CIlower.blank"),
  upper.blank = gs("CIupper.blank"),
  writexl = !missing(path),
  path = "leaguetable.xlsx",
  overwrite = FALSE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)

## S3 method for class 'netleague'
print(
  x,
  common = x$x$common,
  random = x$x$random,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netmeta or netleague (mandatory).
y	An object of class netmeta (optional).

common	A logical indicating whether a league table should be printed for the common effects network meta-analysis.
random	A logical indicating whether a league table should be printed for the random effects network meta-analysis.
seq	A character or numerical vector specifying the sequence of treatments in rows and columns of a league table.
ci	A logical indicating whether confidence intervals should be shown.
backtransf	A logical indicating whether printed results should be back transformed. If backtransf = TRUE, results for sm = "OR" are printed as odds ratios rather than log odds ratios, for example.
direct	A logical indicating whether league table with network estimates (default) or estimates from direct comparisons should be generated if argument y is not missing.
digits	Minimal number of significant digits, see print.default.
big.mark	A character used as thousands separator.
text.NA	A character string to label missing values.
bracket	A character with bracket symbol to print lower confidence interval: "[", "(", "{", "".
separator	A character string with information on separator between lower and upper confidence interval.
lower.blank	A logical indicating whether blanks between left bracket and lower confidence limit should be printed.
upper.blank	A logical indicating whether blanks between separator and upper confidence limit should be printed.
writexl	A logical indicating whether an Excel file should be created (R package **writexl** must be available).
path	A character string specifying the filename of the Excel file.
overwrite	A logical indicating whether an existing Excel file should be overwritten.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (passed on to write_xlsx to create Excel file).

Details

A league table is a square matrix showing all pairwise comparisons in a network meta-analysis (Hutton et al., 2015). Typically, both treatment estimates and confidence intervals are shown.

If argument y is not provided, the league table contains the network estimates from network meta-analysis object x in the lower triangle and the direct treatment estimates from pairwise comparisons in the upper triangle. Note, for the random-effects model, the direct treatment estimates are based on the common between-study variance τ^2 from the network meta-analysis, i.e. the square of list element x$tau.

If argument y is provided, the league table contains information on treatment comparisons from network meta-analysis object x in the lower triangle and from network meta-analysis object y in the upper triangle. This is, for example, useful to print information on efficacy and safety in the same league table.

By default, an R object with the league tables is generated. Alternatively, an Excel file is created if argument writexl = TRUE.

This implementation reports pairwise comparisons of the treatment in the column versus the treatment in the row in the lower triangle and row versus column in the upper triangle. This is a common presentation for network meta-analyses which allows to easily compare direction and magnitude of treatment effects. For example, given treatments A, B, and C, the results reported in the first row and second column as well as second row and first column are from the pairwise comparison A versus B. Note, this presentation is different from the printout of a network meta-analysis object which reports opposite pairwise comparisons in the lower and upper triangle, e.g., A versus B in the first row and second column and B versus A in the second row and first column.

If the same network meta-analysis object is used for arguments x and y, reciprocal treatment estimates will be shown in the upper triangle (see examples), e.g., the comparison B versus A.

R function netrank can be used to change the order of rows and columns in the league table (see examples).

Value

An object of class netleague with corresponding print function if writexl = FALSE. The object is a list containing the league tables in list elements 'common' and 'random'. An Excel file is created if writexl = TRUE. In this case, NULL is returned in R.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Gerta Rücker <ruecker@imbi.uni-freiburg.de>

References

Hutton B, Salanti G, Caldwell DM, et al. (2015): The PRISMA Extension Statement for Reporting of Systematic Reviews Incorporating Network Meta-analyses of Health Care Interventions: Checklist and Explanations. *Annals of Internal Medicine*, **162**, 777

See Also

netmeta, netposet, netrank

Examples

```
# Network meta-analysis of count mortality statistics
#
data(Woods2010)

p0 <- pairwise(treatment, event = r, n = N,
  studlab = author, data = Woods2010, sm = "OR")
net0 <- netmeta(p0)
```

```
oldopts <- options(width = 100)

# League table for common and random effects model with
# - network estimates in lower triangle
# - direct estimates in upper triangle
#
netleague(net0, digits = 2, bracket = "(", separator = " - ")

# League table for common effects model
#
netleague(net0, random = FALSE, digits = 2)

# Change order of treatments according to treatment ranking (random
# effects model)
#
netleague(net0, common = FALSE, digits = 2, seq = netrank(net0))
#
print(netrank(net0), common = FALSE)

## Not run:
# Create a CSV file with league table for random effects model
#
league0 <- netleague(net0, digits = 2, bracket = "(", separator = " to ")
#
write.table(league0$random, file = "league0-random.csv",
  row.names = FALSE, col.names = FALSE, sep = ",")
#
# Create Excel files with league tables
# (if R package writexl is available)
#
netleague(net0, digits = 2, bracket = "(", separator = " to ",
          path = tempfile(fileext = ".xlsx"))

## End(Not run)


# Use depression dataset
#
data(Linde2015)

# Define order of treatments
#
trts <- c("TCA", "SSRI", "SNRI", "NRI",
  "Low-dose SARI", "NaSSa", "rMAO-A", "Hypericum", "Placebo")

# Outcome labels
#
outcomes <- c("Early response", "Early remission")

# (1) Early response
#
p1 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(resp1, resp2, resp3), n = list(n1, n2, n3),
```

```
  studlab = id, data = Linde2015, sm = "OR")
#
net1 <- netmeta(p1, common = FALSE,
                seq = trts, ref = "Placebo", small = "bad")

# (2) Early remission
#
p2 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(remi1, remi2, remi3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net2 <- netmeta(p2, common = FALSE,
                seq = trts, ref = "Placebo", small = "bad")

options(width = 200)
netleague(net1, digits = 2)

netleague(net1, digits = 2, ci = FALSE)
netleague(net2, digits = 2, ci = FALSE)

# League table for two outcomes with
# - network estimates of first outcome in lower triangle
# - network estimates of second outcome in upper triangle
#
netleague(net1, net2, digits = 2, ci = FALSE)

netleague(net1, net2, seq = netrank(net1), ci = FALSE)
netleague(net1, net2, seq = netrank(net2), ci = FALSE)

print(netrank(net1))
print(netrank(net2))


# Report results for network meta-analysis twice
#
netleague(net1, net1, seq = netrank(net1), ci = FALSE,
  backtransf = FALSE)
netleague(net1, net1, seq = netrank(net1), ci = FALSE,
  backtransf = FALSE, direct = TRUE)


options(oldopts)

## Not run:
# Generate a partial order of treatment rankings
#
np <- netposet(net1, net2, outcomes = outcomes)

# Requires R package 'hasse'
#
hasse(np)
plot(np)
```

```
## End(Not run)
```

`netmatrix` *Create a matrix with additional information for pairwise comparisons*

Description

Auxiliary function to create a matrix with additional information for pairwise comparisons

Usage

```
netmatrix(
  x,
  var,
  levels,
  labels = levels,
  func = "mode",
  ties.method = "random"
)
```

Arguments

`x`	A `netmeta` object.
`var`	Variable with additional information.
`levels`	An optional vector of the values that `var` might have taken (see `factor`).
`labels`	An optional vector with labels for `var` (see `factor`).
`func`	A character string with the function name to summarize values within pairwise comparisons; see Details.
`ties.method`	A character string describing how ties are handled if `func = "mode"`; see Details.

Details

For each pairwise comparison, unique values will be calculated for the variable `var` based on the argument `func`: "mode" (most common value), "min" (minimum value), "max", "mean", "median", and "sum". In order to determine the most common value, the argument `ties.method` can be used in the case of ties with "first" meaning that the first / smallest value will be selected; similar for "last" (last / largest value) and "random" (random selection).

Value

A matrix with the same row and column names as the adjacency matrix `x$A.matrix`.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netmeta, netgraph.netmeta

Examples

```
data(smokingcessation)
# Add variable with (fictious) risk of bias values
# with 1 = "low risk" and 2 = "high risk"
#
smokingcessation$rob <- rep(1:2, 12)

p1 <- pairwise(list(treat1, treat2, treat3),
  event = list(event1, event2, event3), n = list(n1, n2, n3),
  data = smokingcessation, sm = "OR")
net1 <- netmeta(p1, common = FALSE, ref = "A")

# Generate network graph with information on risk of bias
#
col.rob <- netmatrix(net1, rob, ties.method = "last",
  levels = 1:2, labels = c("green", "yellow"))
#
netgraph(net1, plastic = FALSE, col = col.rob,
  cex.points = 5, bg.points = "gray", adj = 0.5)

netgraph(net1, plastic = FALSE, col = col.rob,
  cex.points = n.trts, bg.points = "blue",
  labels = paste0(trts, " (n=", n.trts, ")"),
  offset = c(0.05, 0.035, 0.05, 0.025))
```

---

netmeasures *Measures for characterizing a network meta-analysis*

Description

This function provides measures for quantifying the direct evidence proportion, the mean path length and the minimal parallelism (the latter on aggregated and study level) of mixed treatment comparisons (network estimates) as well as the evidence flow per design, see König et al. (2013). These measures support the critical evaluation of the network meta-analysis results by rendering transparent the process of data pooling.

Usage

```
netmeasures(
  x,
  random = x$random | !missing(tau.preset),
  tau.preset = x$tau.preset,
  warn = TRUE,
  warn.deprecated = gs("warn.deprecated"),
```

```
  ...
)
```

Arguments

x	An object of class netmeta.
random	A logical indicating whether random effects model should be used to calculate network measures.
tau.preset	An optional value for the square-root of the between-study variance τ^2.
warn	A logical indicating whether warnings should be printed.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).

Details

The direct evidence proportion gives the absolute contribution of direct effect estimates combined for two-arm and multi-arm studies to one network estimate.

Concerning indirectness, comparisons with a mean path length beyond two should be interpreted with particular caution, as more than two direct comparisons have to be combined serially on average.

Large indices of parallelism, either on study-level or on aggregated level, can be considered as supporting the validity of a network meta-analysis if there is only a small amount of heterogeneity.

The network estimates for two treatments are linear combinations of direct effect estimates comparing these or other treatments. The linear coefficients can be seen as the generalization of weights known from classical meta-analysis. These coefficients are given in the projection matrix H of the underlying model. For multi-arm studies, the coefficients depend on the choice of the study-specific baseline treatment, but the absolute flow of evidence can be made explicit for each design as shown in König et al. (2013) and is given in H.tilde.

All measures are calculated based on the common effects meta-analysis by default. In the case that in function netmeta the argument random = TRUE, all measures are calculated for a random effects model. The value of the square-root of the between-study variance τ^2 can also be prespecified by argument tau.preset in function netmeta.

Value

A list containing the following components:

random, tau.preset	As defined above.
proportion	A named vector of the direct evidence proportion of each network estimate.
meanpath	A named vector of the mean path length of each network estimate.
minpar	A named vector of the minimal parallelism on aggregated level of each network estimate.

minpar.study	A named vector of the minimal parallelism on study level of each network estimate.
H.tilde	Design-based hat matrix with information on absolute evidence flow per design. The number of rows is equal to the number of possible pairwise treatment comparisons and the number of columns is equal to the number of designs.

Author(s)

Ulrike Krahn <ulrike.krahn@bayer.com>, Jochem König <koenigjo@uni-mainz.de>

References

König J, Krahn U, Binder H (2013): Visualizing the flow of evidence in network meta-analysis and characterizing mixed treatment comparisons. *Statistics in Medicine*, **32**, 5414–29

See Also

netmeta

Examples

```
data(Senn2013)

# Conduct common effects network meta-analysis with reference
# treatment 'plac', i.e. placebo
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", reference = "plac", random = FALSE)

# Calculate measures based on a common effects model
#
nm1 <- netmeasures(net1)

# Plot of minimal parallelism versus mean path length
#
plot(nm1$meanpath, nm1$minpar, pch = "",
  xlab = "Mean path length", ylab = "Minimal parallelism")
text(nm1$meanpath, nm1$minpar, names(nm1$meanpath), cex = 0.8)

## Not run:
# Conduct random effects network meta-analysis with reference
# treatment 'plac', i.e. placebo
#
net2 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", reference = "plac", common = FALSE)

# Calculate measures based on a random effects model
#
nm2 <- netmeasures(net2)

## End(Not run)
```

netmeta *Network meta-analysis using graph-theoretical method*

Description

Network meta-analysis is a generalisation of pairwise meta-analysis that compares all pairs of treatments within a number of treatments for the same condition. The graph-theoretical approach for network meta-analysis uses methods that were originally developed in electrical network theory. It has been found to be equivalent to the frequentist approach to network meta-analysis which is based on weighted least squares regression (Rücker, 2012).

Print method for objects of class `netmeta`.

Usage

```
netmeta(
  TE,
  seTE,
  treat1,
  treat2,
  studlab,
  data = NULL,
  subset = NULL,
  sm,
  level = gs("level"),
  level.ma = gs("level.ma"),
  common = gs("common"),
  random = gs("random") | !is.null(tau.preset),
  prediction = FALSE,
  level.predict = gs("level.predict"),
  reference.group,
  baseline.reference = TRUE,
  small.values = "good",
  all.treatments = NULL,
  seq = NULL,
  method.tau = "DL",
  tau.preset = NULL,
  tol.multiarm = 0.001,
  tol.multiarm.se = NULL,
  details.chkmultiarm = FALSE,
  sep.trts = ":",
  nchar.trts = 666,
  nchar.studlab = 666,
  func.inverse = invmat,
  n1 = NULL,
  n2 = NULL,
  event1 = NULL,
```

```
  event2 = NULL,
  incr = NULL,
  sd1 = NULL,
  sd2 = NULL,
  time1 = NULL,
  time2 = NULL,
  backtransf = gs("backtransf"),
  title = "",
  keepdata = gs("keepdata"),
  control = NULL,
  warn = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  nchar = nchar.trts,
  ...
)

## S3 method for class 'netmeta'
print(
  x,
  common = x$common,
  random = x$random,
  prediction = x$prediction,
  reference.group = x$reference.group,
  baseline.reference = x$baseline.reference,
  all.treatments = x$all.treatments,
  backtransf = x$backtransf,
  nchar.trts = x$nchar.trts,
  header = TRUE,
  digits = gs("digits"),
  digits.stat = gs("digits.stat"),
  digits.pval = max(gs("digits.pval"), 2),
  digits.pval.Q = max(gs("digits.pval.Q"), 2),
  digits.Q = gs("digits.Q"),
  digits.tau2 = gs("digits.tau2"),
  digits.tau = gs("digits.tau"),
  digits.I2 = gs("digits.I2"),
  scientific.pval = gs("scientific.pval"),
  big.mark = gs("big.mark"),
  text.tau2 = gs("text.tau2"),
  text.tau = gs("text.tau"),
  text.I2 = gs("text.I2"),
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

TE	Estimate of treatment effect, i.e. difference between first and second treatment (e.g. log odds ratio, mean difference, or log hazard ratio). Or an R object created with pairwise.
seTE	Standard error of treatment estimate.
treat1	Label/Number for first treatment.
treat2	Label/Number for second treatment.
studlab	An optional - but important! - vector with study labels (see Details).
data	An optional data frame containing the study information.
subset	An optional vector specifying a subset of studies to be used.
sm	A character string indicating underlying summary measure, e.g., "RD", "RR", "OR", "ASD", "HR", "MD", "SMD", or "ROM".
level	The level used to calculate confidence intervals for individual comparisons.
level.ma	The level used to calculate confidence intervals for network estimates.
common	A logical indicating whether results for the common effects model should be printed.
random	A logical indicating whether results for the random effects model should be printed.
prediction	A logical indicating whether prediction intervals should be printed.
level.predict	The level used to calculate prediction intervals for a new study.
reference.group	Reference treatment.
baseline.reference	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa. This argument is only considered if reference.group has been specified.
small.values	A character string specifying whether small treatment effects indicate a beneficial ("good") or harmful ("bad") effect (passed on to netrank, can be abbreviated.
all.treatments	A logical or "NULL". If TRUE, matrices with all treatment effects, and confidence limits will be printed.
seq	A character or numerical vector specifying the sequence of treatments in printouts.
method.tau	A character string indicating which method is used to estimate the between-study variance τ^2 and its square root τ. Either "DL", "REML", or "ML", can be abbreviated.
tau.preset	An optional value for manually setting the square-root of the between-study variance τ^2.
tol.multiarm	A numeric for the tolerance for consistency of treatment estimates in multi-arm studies which are consistent by design.

`tol.multiarm.se`
: A numeric for the tolerance for consistency of standard errors in multi-arm studies which are consistent by design. This check is not conducted if the argument is `NULL`.

`details.chkmultiarm`
: A logical indicating whether treatment estimates and / or variances of multi-arm studies with inconsistent results or negative multi-arm variances should be printed.

`sep.trts`
: A character used in comparison names as separator between treatment labels.

`nchar.trts`
: A numeric defining the minimum number of characters used to create unique treatment names.

`nchar.studlab`
: A numeric defining the minimum number of characters used to create unique study labels.

`func.inverse`
: R function used to calculate the pseudoinverse of the Laplacian matrix L (see Details).

`n1`
: Number of observations in first treatment group.

`n2`
: Number of observations in second treatment group.

`event1`
: Number of events in first treatment group.

`event2`
: Number of events in second treatment group.

`incr`
: Numerical value added to cell frequencies (for details, see `pairwise`).

`sd1`
: Standard deviation in first treatment group.

`sd2`
: Standard deviation in second treatment group.

`time1`
: Person time at risk in first treatment group.

`time2`
: Person time at risk in second treatment group.

`backtransf`
: A logical indicating whether results should be back transformed in printouts and forest plots. If `backtransf = TRUE`, results for `sm = "OR"` are presented as odds ratios rather than log odds ratios, for example.

`title`
: Title of meta-analysis / systematic review.

`keepdata`
: A logical indicating whether original data (set) should be kept in netmeta object.

`control`
: An optional list to control the iterative process to estimate the between-study variance τ^2. This argument is passed on to `rma.mv`.

`warn`
: A logical indicating whether warnings should be printed (e.g., if studies are excluded from meta-analysis due to zero standard errors).

`warn.deprecated`
: A logical indicating whether warnings should be printed if deprecated arguments are used.

`nchar`
: Deprecated argument (replaced by `nchar.trts`).

`...`
: Additional arguments.

`x`
: An object of class `netmeta`.

`header`
: A logical indicating whether information on title of meta-analysis, comparison and outcome should be printed at the beginning of the printout.

digits	Minimal number of significant digits, see print.default.
digits.stat	Minimal number of significant digits for tests of overall effect, see print.default.
digits.pval	Minimal number of significant digits for p-value of overall effects, see print.default.
digits.pval.Q	Minimal number of significant digits for p-value of heterogeneity tests, see print.default.
digits.Q	Minimal number of significant digits for heterogeneity statistics, see print.default.
digits.tau2	Minimal number of significant digits for between-study variance, see print.default.
digits.tau	Minimal number of significant digits for square root of between-study variance, see print.default.
digits.I2	Minimal number of significant digits for I-squared statistic, see print.default.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
big.mark	A character used as thousands separator.
text.tau2	Text printed to identify between-study variance τ^2.
text.tau	Text printed to identify τ, the square root of the between-study variance τ^2.
text.I2	Text printed to identify heterogeneity statistic I^2.
legend	A logical indicating whether a legend should be printed.

Details

Network meta-analysis using R package **netmeta** is described in detail in Schwarzer et al. (2015), Chapter 8.

Let *n* be the number of different treatments (nodes, vertices) in a network and let *m* be the number of existing comparisons (edges) between the treatments. If there are only two-arm studies, *m* is the number of studies. Let TE and seTE be the vectors of observed effects and their standard errors. Let W be the *m*x*m* diagonal matrix that contains the inverse variance 1 / seTE^2.

The given comparisons define the network structure. Therefrom an *m*x*n* design matrix X (edge-vertex incidence matrix) is formed; for more precise information, see Rücker (2012). Moreover, the *n*x*n* Laplacian matrix L and its Moore-Penrose pseudoinverse L+ are calculated (both matrices play an important role in graph theory and electrical network theory). Using these matrices, the variances based on both direct and indirect comparisons can be estimated. Moreover, the hat matrix H can be estimated by **H = XL+X^tW = X(X^t W X)^+X^tW** and finally consistent treatment effects can be estimated by applying the hat matrix to the observed (potentially inconsistent) effects. H is a projection matrix which maps the observed effects onto the consistent (n-1)-dimensional subspace. This is the Aitken estimator (Senn et al., 2013). As in pairwise meta-analysis, the Q statistic measures the deviation from consistency. Q can be separated into parts for each pairwise meta-analysis and a part for remaining inconsistency between comparisons.

Often multi-arm studies are included in a network meta-analysis. In multi-arm studies, the treatment effects on different comparisons are not independent, but correlated. This is accounted for by reweighting all comparisons of each multi-arm study. The method is described in Rücker (2012) and Rücker and Schwarzer (2014).

Comparisons belonging to multi-arm studies are identified by identical study labels (argument studlab). It is therefore important to use identical study labels for all comparisons belonging to the

same multi-arm study, e.g., study label "Willms1999" for the three-arm study in the data example (Senn et al., 2013). The function netmeta then automatically accounts for within-study correlation by reweighting all comparisons of each multi-arm study.

Data entry for this function is in *contrast-based* format, that is, data are given as contrasts (differences) between two treatments (argument `TE`) with standard error (argument `seTE`). In principle, meta-analysis functions from R package **meta**, e.g. `metabin` for binary outcomes or `metacont` for continuous outcomes, can be used to calculate treatment effects separately for each treatment comparison which is a rather tedious enterprise. If data are provided in *arm-based* format, that is, data are given for each treatment arm separately (e.g. number of events and participants for binary outcomes), a much more convenient way to transform data into contrast-based form is available. Function `pairwise` can automatically transform data with binary outcomes (using the `metabin` function from R package **meta**), continuous outcomes (`metacont` function), incidence rates (`metainc` function), and generic outcomes (`metagen` function). Additional arguments of these functions can be provided (see help page of function `pairwise`).

Note, all pairwise comparisons must be provided for a multi-arm study. Consider a multi-arm study of p treatments with known variances. For this study, treatment effects and standard errors must be provided for each of $p(p - 1) / 2$ possible comparisons. For instance, a three-arm study contributes three pairwise comparisons, a four-arm study even six pairwise comparisons. Function `pairwise` automatically calculates all pairwise comparisons for multi-arm studies.

A simple random effects model assuming that a constant heterogeneity variance is added to each comparison of the network can be defined via a generalised methods of moments estimate of the between-studies variance τ^2 (Jackson et al., 2012). This is added to the observed sampling variance `seTE^2` of each comparison in the network (before appropriate adjustment for multi-arm studies). Then, as in standard pairwise meta-analysis, the procedure is repeated with the resulting enlarged standard errors.

For the random-effects model, the direct treatment estimates are based on the common between-study variance τ^2 from the network meta-analysis.

Internally, both common and random effects models are calculated regardless of values choosen for arguments `common` and `random`. Accordingly, the network estimates for the random effects model can be extracted from component `TE.random` of an object of class `"netmeta"` even if argument `random = FALSE`. However, all functions in R package **netmeta** will adequately consider the values for `common` and `random`. E.g. function `print.summary.netmeta` will not print results for the random effects model if `random = FALSE`.

By default, treatment names are not abbreviated in printouts. However, in order to get more concise printouts, argument `nchar.trts` can be used to define the minimum number of characters for abbreviated treatment names (see `abbreviate`, argument `minlength`). R function `treats` is utilised internally to create abbreviated treatment names.

Names of treatment comparisons are created by concatenating treatment labels of pairwise comparisons using `sep.trts` as separator (see `paste`). These comparison names are used in the covariance matrices `Cov.common` and `Cov.random` and in some R functions, e.g, `decomp.design`. By default, a colon is used as the separator. If any treatment label contains a colon the following characters are used as separator (in consecutive order): `"-"`, `"_"`, `"/"`, `"+"`, `"."`, `"|"`, and `"*"`. If all of these characters are used in treatment labels, a corresponding error message is printed asking the user to specify a different separator.

Value

An object of class netmeta with corresponding print, summary, forest, and netrank functions. The object is a list containing the following components:

studlab, treat1, treat2, TE, seTE
: As defined above.

seTE.adj.common, seTE.adj.random
: Standard error of treatment estimate, adjusted for multi-arm studies.

design
: Design of study providing pairwise comparison.

n1, n2, event1, event2, incr
: As defined above.

mean1, mean2, sd1, sd2, time1, time2
: As defined above.

sd1, sd2, time1, time2
: As defined above.

k
: Total number of studies.

m
: Total number of pairwise comparisons.

n
: Total number of treatments.

d
: Total number of designs (corresponding to the unique set of treatments compared within studies).

trts
: Treatments included in network meta-analysis.

k.trts
: Number of studies evaluating a treatment.

n.trts
: Number of observations receiving a treatment (if arguments n1 and n2 are provided).

events.trts
: Number of events observed for a treatment (if arguments event1 and event2 are provided).

multiarm
: Logical vector to identify pairwise comparisons from multi-arm studies.

n.arms
: Number of treatment arms in study providing pairwise comparison.

studies
: Vector with unique study labels.

narms
: Number of arms for each study.

designs
: Vector with unique designs present in the network. A design corresponds to the set of treatments compared within a study.

designs
: Vector with unique direct comparisons present in the network.

TE.nma.common, TE.nma.random
: A vector of length *m* of consistent treatment effects estimated by network meta-analysis (nma) (common / random effects model).

seTE.nma.common, seTE.nma.random
: A vector of length *m* of effective standard errors estimated by network meta-analysis (common / random effects model).

lower.nma.common, lower.nma.random
: A vector of length *m* of lower confidence interval limits for consistent treatment effects estimated by network meta-analysis (common effects / random effects model).

`upper.nma.common`, `upper.nma.random`
: A vector of length *m* of upper confidence interval limits for the consistent treatment effects estimated by network meta-analysis (common effects / random effects model).

`statistic.nma.common`, `statistic.nma.random`
: A vector of length *m* of z-values for test of treatment effect for individual comparisons (common / random effects model).

`pval.nma.common`, `pval.nma.random`
: A vector of length *m* of p-values for test of treatment effect for individual comparisons (common / random effects model).

`leverage.common`
: A vector of length *m* of leverages, interpretable as factors by which variances are reduced using information from the whole network.

`w.common`, `w.random`
: A vector of length *m* of weights of individual studies (common / random effects model).

`Q.common`
: A vector of length *m* of contributions to total heterogeneity / inconsistency statistic.

`TE.common`, `TE.random`
: *n*x*n* matrix with estimated overall treatment effects (common / random effects model).

`seTE.common`, `seTE.random`
: *n*x*n* matrix with standard errors (common / random effects model).

`lower.common`, `upper.common`, `lower.random`, `upper.random`
: *n*x*n* matrices with lower and upper confidence interval limits (common / random effects model).

`statistic.common`, `pval.common`, `statistic.random`, `pval.random`
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect (common / random effects model).

`seTE.predict`
: *n*x*n* matrix with standard errors for prediction intervals.

`lower.predict`, `upper.predict`
: *n*x*n* matrices with lower and upper prediction interval limits.

`prop.direct.common`, `prop.direct.random`
: A named vector of the direct evidence proportion of each network estimate. (common effects / random effects model).

`TE.direct.common`, `TE.direct.random`
: *n*x*n* matrix with estimated treatment effects from direct evidence (common effects / random effects model).

`seTE.direct.common`, `seTE.direct.random`
: *n*x*n* matrix with estimated standard errors from direct evidence (common effects / random effects model).

`lower.direct.common`, `upper.direct.common`, `lower.direct.random`,
: *n*x*n* matrices with lower and upper confidence interval limits from direct evidence (common / random effects model).

`upper.direct.random`
: *n*x*n* matrices with lower and upper confidence interval limits from direct evidence (common effects / random effects model).

`statistic.direct.common`, `pval.direct.common`, `statistic.direct.random`,
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect from direct evidence (common / random effects model).

`pval.direct.random`
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect from direct evidence (common / random effects model).

`TE.indirect.common`, `TE.indirect.random`
: *n*x*n* matrix with estimated treatment effects from indirect evidence (common / random effects model).

`seTE.indirect.common`, `seTE.indirect.random`
: *n*x*n* matrix with estimated standard errors from indirect evidence (common / random effects model).

`lower.indirect.common`, `upper.indirect.common`, `lower.indirect.random`,
: *n*x*n* matrices with lower and upper confidence interval limits from indirect evidence (common / random effects model).

`upper.indirect.random`
: *n*x*n* matrices with lower and upper confidence interval limits from indirect evidence (common / random effects model).

`statistic.indirect.common`, `pval.indirect.common`, `statistic.indirect.random`,
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect from indirect evidence (common / random effects model).

`pval.indirect.random`
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect from indirect evidence (common / random effects model).

`Q`
: Overall heterogeneity / inconsistency statistic.

`df.Q`
: Degrees of freedom for test of heterogeneity / inconsistency.

`pval.Q`
: P-value for test of heterogeneity / inconsistency.

`I2`, `lower.I2`, `upper.I2`
: I-squared, lower and upper confidence limits.

`tau`
: Square-root of between-study variance.

`Q.heterogeneity`
: Overall heterogeneity statistic.

`df.Q.heterogeneity`
: Degrees of freedom for test of overall heterogeneity.

`pval.Q.heterogeneity`
: P-value for test of overall heterogeneity.

`Q.inconsistency`
: Overall inconsistency statistic.

`df.Q.inconsistency`
: Degrees of freedom for test of overall inconsistency.

`pval.Q.inconsistency`
: P-value for test of overall inconsistency.

`Q.decomp`
: Data frame with columns 'treat1', 'treat2', 'Q', 'df' and 'pval.Q', providing heterogeneity statistics for each pairwise meta-analysis of direct comparisons.

`A.matrix` Adjacency matrix (*n*x*n*).

`X.matrix` Design matrix (*m*x*n*).

`B.matrix` Edge-vertex incidence matrix (*m*x*n*).

`L.matrix.common`, `L.matrix.random`
Laplacian matrix (*n*x*n*).

`Lplus.matrix.common`, `Lplus.matrix.random`
Moore-Penrose pseudoinverse of the Laplacian matrix (*n*x*n*).

`Q.matrix` Matrix of heterogeneity statistics for pairwise meta-analyses, where direct comparisons exist (*n*x*n*).

`G.matrix` Matrix with variances and covariances of comparisons (*m*x*m*). G is defined as **BL+B^t**.

`H.matrix.common`, `H.matrix.random`
Hat matrix (*m*x*m*), defined as **H = GW = BL+B^tW**.

`n.matrix` *n*x*n* matrix with number of observations in direct comparisons (if arguments `n1` and `n2` are provided).

`events.matrix` *n*x*n* matrix with number of events in direct comparisons (if arguments `event1` and `event2` are provided).

`P.common`, `P.random`
*n*x*n* matrix with direct evidence proportions (common / random effects model).

`Cov.common` Variance-covariance matrix (common effects model)

`Cov.random` Variance-covariance matrix (random effects model)

`sm`, `level`, `level.ma`
As defined above.

`common`, `random` As defined above.

`prediction`, `level.predict`
As defined above.

`reference.group`, `baseline.reference`, `small.values`, `all.treatments`
As defined above.

`seq`, `tau.preset`, `tol.multiarm`, `tol.multiarm.se`
As defined above.

`details.chkmultiarm`, `sep.trts`, `nchar.trts`
As defined above.

`backtransf`, `title`, `warn`, `warn.deprecated`
As defined above.

`call` Function call.

`version` Version of R package netmeta used to create object.

Note

R function `rma.mv` from R package **metafor** (Viechtbauer 2010) is called internally to estimate the between-study variance τ^2 for the (restricted) maximum likelihood method. For binary outcomes, incidence rates, and the mean difference, the variance-covariance matrix is calculated if arguments `event1`, `event2`, `n1`, and `n2` (binary outcomes); `event1`, `event2`, `time1`, and `time2` (incidence rates); `n1`, `n2`, `sd1`, and `sd2` (mean difference) are provided. For data sets preprocessed with `pairwise` the respective variables are selected automatically.

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Senn S, Gavini F, Magrez D, Scheen A (2013): Issues in performing a network meta-analysis. *Statistical Methods in Medical Research*, **22**, 169–89

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See Also

pairwise, forest.netmeta, netrank, metagen

Examples

```
data(Senn2013)

# Conduct common effects network meta-analysis
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", random = FALSE)
net1
net1$Q.decomp

# Comparison with reference group
#
print(net1, reference = "plac")

## Not run:
# Conduct random effects network meta-analysis
#
net2 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", common = FALSE)
net2

# Change printing order of treatments with placebo last and use
# long treatment names
#
trts <- c("acar", "benf", "metf", "migl", "piog",
  "rosi", "sita", "sulf", "vild", "plac")
```

```
net3 <- netmeta(TE, seTE, treat1.long, treat2.long, studlab,
  data = Senn2013, sm = "MD", common = FALSE,
  seq = trts, reference = "Placebo")
print(net3, digits = 2)

## End(Not run)
```

`netmetabin` *Network meta-analysis of binary outcome data*

Description

Provides three models for the network meta-analysis of binary data (Mantel-Haenszel method, based on the non-central hypergeometric distribution, and the inverse variance method).

Usage

```
netmetabin(
  event1,
  n1,
  event2,
  n2,
  treat1,
  treat2,
  studlab,
  data = NULL,
  subset = NULL,
  sm,
  method = "MH",
  cc.pooled = FALSE,
  incr,
  allincr,
  addincr,
  allstudies,
  level = gs("level"),
  level.ma = gs("level.ma"),
  common = gs("common"),
  random = method == "Inverse" & (gs("random") | !is.null(tau.preset)),
  prediction = FALSE,
  level.predict = gs("level.predict"),
  reference.group = "",
  baseline.reference = TRUE,
  all.treatments = NULL,
  seq = NULL,
  tau.preset = NULL,
  tol.multiarm = 0.001,
```

```
  tol.multiarm.se = NULL,
  details.chkmultiarm = FALSE,
  details.chkdata = TRUE,
  sep.trts = ":",
  nchar.trts = 666,
  func.inverse = invmat,
  backtransf = gs("backtransf"),
  title = "",
  keepdata = gs("keepdata"),
  warn = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

event1	Number of events (first treatment).
n1	Number of observations (first treatment).
event2	Number of events (second treatment).
n2	Number of observations (second treatment)
treat1	Label/Number for first treatment.
treat2	Label/Number for second treatment.
studlab	An optional - but important! - vector with study labels (see Details).
data	An optional data frame containing the study information.
subset	An optional vector specifying a subset of studies to be used.
sm	A character string indicating underlying summary measure, i.e., "RD", "RR", "OR", "ASD".
method	A character string indicating which method is to be used for pooling of studies. One of "Inverse", "MH", or "NCH", can be abbreviated.
cc.pooled	A logical indicating whether incr should be used as a continuity correction, when calculating the network meta-analysis estimates.
incr	A numerical value which is added to each cell count, i.e., to the numbers of events and non-events, of all treatment arms in studies with zero events or non-events in any of the treatment arms ("continuity correction").
allincr	A logical indicating whether incr should be added to each cell count of all studies if a continuity correction was used for at least one study (only considered if method = "Inverse"). If FALSE (default), incr is used as continuity correction only for studies with zero events or zero non-events in any of the treatment arms.
addincr	A logical indicating whether incr should be added to each cell count of all studies, irrespective of zero cell counts (only considered if method = "Inverse").
allstudies	A logical indicating whether studies with zero events or non-events in all treatment arms should be included in an inverse variance meta-analysis (applies only if method = "Inverse" and sm is equal to either "RR" or "OR").

`level` The level used to calculate confidence intervals for individual studies.

`level.ma` The level used to calculate confidence intervals for network estimates.

`common` A logical indicating whether a common effects network meta-analysis should be conducted.

`random` A logical indicating whether a random effects network meta-analysis should be conducted.

`prediction` A logical indicating whether a prediction interval should be printed (only considered if `method = "Inverse"`).

`level.predict` The level used to calculate prediction interval for a new study (only considered if `method = "Inverse"`).

`reference.group`
Reference treatment.

`baseline.reference`
A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa. This argument is only considered if `reference.group` has been specified.

`all.treatments` A logical or `"NULL"`. If `TRUE`, matrices with all treatment effects, and confidence limits will be printed.

`seq` A character or numerical vector specifying the sequence of treatments in printouts.

`tau.preset` An optional value for manually setting the square-root of the between-study variance τ^2 (only considered if `method = "Inverse"`).

`tol.multiarm` A numeric for the tolerance for consistency of treatment estimates in multi-arm studies which are consistent by design (only considered if `method = "Inverse"`).

`tol.multiarm.se`
A numeric for the tolerance for consistency of standard errors in multi-arm studies which are consistent by design (only considered if the argument is not `NULL` and `method = "Inverse"`).

`details.chkmultiarm`
A logical indicating whether treatment estimates and / or variances of multi-arm studies with inconsistent results or negative multi-arm variances should be printed (only considered if `method = "Inverse"`).

`details.chkdata`
A logical indicating whether number of events and participants of studies with inconsistent data should be printed.

`sep.trts` A character used in comparison names as separator between treatment labels.

`nchar.trts` A numeric defining the minimum number of characters used to create unique treatment names (see Details).

`func.inverse` R function used to calculate the pseudoinverse of the Laplacian matrix L (see `netmeta`).

`backtransf` A logical indicating whether results should be back transformed in printouts and forest plots. If `backtransf = TRUE`, results for `sm = "OR"` are presented as odds ratios rather than log odds ratios, for example.

`title` Title of meta-analysis / systematic review.

keepdata	A logical indicating whether original data (set) should be kept in netmeta object.
warn	A logical indicating whether warnings should be printed (e.g., if studies are excluded from meta-analysis due to zero standard errors).
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).

Details

This function implements three models for the network meta-analysis of binary data:

- The Mantel-Haenszel network meta-analysis model, as described in Efthimiou et al. (2019) (method = "MH");
- a network meta-analysis model using the non-central hypergeometric distribution with the Breslow approximation, as described in Stijnen et al. (2010) (method = "NCH");
- the inverse variance method for network meta-analysis (method = "Inverse"), also provided by netmeta.

Comparisons belonging to multi-arm studies are identified by identical study labels (argument studlab). It is therefore important to use identical study labels for all comparisons belonging to the same multi-arm study.

Data entry for this function is in *contrast-based* format, that is, each line of the data corresponds to a single pairwise comparison between two treatments (arguments treat1, treat2, event1, n1, event2, and n2). If data are provided in *arm-based* format, that is, number of events and participants are given for each treatment arm separately, function pairwise can be used to transform the data to *contrast-based* format (see help page of function pairwise).

Note, all pairwise comparisons must be provided for a multi-arm study. Consider a multi-arm study of p treatments with known variances. For this study, the number of events and observations must be provided for each treatment, for each of $p(p - 1) / 2$ possible comparisons in separate lines in the data. For instance, a three-arm study contributes three pairwise comparisons, a four-arm study even six pairwise comparisons. Function pairwise automatically calculates all pairwise comparisons for multi-arm studies.

For method = "Inverse", both common and random effects models are calculated regardless of values choosen for arguments common and random. Accordingly, the network estimates for the random effects model can be extracted from component TE.random of an object of class "netmeta" even if argument random = FALSE. However, all functions in R package **netmeta** will adequately consider the values for common and random. E.g. function print.summary.netmeta will not print results for the random effects model if random = FALSE.

For the random-effects model, the direct treatment estimates are based on the common between-study variance τ^2 from the network meta-analysis.

For method = "MH" and method = "NCH", only a common effects model is available.

By default, treatment names are not abbreviated in printouts. However, in order to get more concise printouts, argument nchar.trts can be used to define the minimum number of characters for abbreviated treatment names (see abbreviate, argument minlength). R function treats is utilised internally to create abbreviated treatment names.

Names of treatment comparisons are created by concatenating treatment labels of pairwise comparisons using `sep.trts` as separator (see `paste`). These comparison names are used in the covariance matrices `Cov.common` and `Cov.random` and in some R functions, e.g, `decomp.design`. By default, a colon is used as the separator. If any treatment label contains a colon the following characters are used as separator (in consecutive order): `"-"`, `"_"`, `"/"`, `"+"`, `"."`, `"|"`, and `"*"`. If all of these characters are used in treatment labels, a corresponding error message is printed asking the user to specify a different separator.

Value

An object of class `netmetabin` and `netmeta` with corresponding `print`, `summary`, `forest`, and `netrank` functions. The object is a list containing the following components:

`studlab, treat1, treat2`
: As defined above.

`n1, n2, event1, event2`
: As defined above.

`TE`
: Estimate of treatment effect, i.e. difference between first and second treatment (e.g. log odds ratio).

`seTE`
: Standard error of treatment estimate.

`k`
: Total number of studies.

`m`
: Total number of pairwise comparisons.

`n`
: Total number of treatments.

`d`
: Total number of designs (corresponding to the unique set of treatments compared within studies).

`trts`
: Treatments included in network meta-analysis.

`k.trts`
: Number of studies evaluating a treatment.

`n.trts`
: Number of observations receiving a treatment.

`events.trts`
: Number of events observed for a treatment.

`studies`
: Study labels coerced into a factor with its levels sorted alphabetically.

`narms`
: Number of arms for each study.

`designs`
: Unique list of designs present in the network. A design corresponds to the set of treatments compared within a study.

`TE.common, seTE.common`
: *n*x*n* matrix with estimated overall treatment effects and standard errors for common effects model.

`lower.common, upper.common`
: *n*x*n* matrices with lower and upper confidence interval limits for common effects model.

`statistic.common, pval.common`
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect under common effects model.

`TE.random, seTE.random`
: *n*x*n* matrix with estimated overall treatment effects and standard errors for random effects model (only available if `method = "Inverse"`).

`lower.random`, `upper.random`
: *n*x*n* matrices with lower and upper confidence interval limits for random effects model (only available if `method = "Inverse"`).

`statistic.random`, `pval.random`
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect under random effects model (only available if `method = "Inverse"`).

`TE.direct.common`, `seTE.direct.common`
: *n*x*n* matrix with estimated treatment effects and standard errors from direct evidence under common effects model.

`lower.direct.common`, `upper.direct.common`
: *n*x*n* matrices with lower and upper confidence interval limits from direct evidence under common effects model.

`statistic.direct.common`, `pval.direct.common`
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect from direct evidence under common effects model.

`TE.direct.random`, `seTE.direct.random`
: *n*x*n* matrix with estimated treatment effects and standard errors from direct evidence under random effects model (only available if `method = "Inverse"`).

`lower.direct.random`, `upper.direct.random`
: *n*x*n* matrices with lower and upper confidence interval limits from direct evidence under random effects model (only available if `method = "Inverse"`).

`statistic.direct.random`, `pval.direct.random`
: *n*x*n* matrices with z-value and p-value for test of overall treatment effect from direct evidence under random effects model (only available if `method = "Inverse"`).

`Q`
: Overall heterogeneity / inconsistency statistic. (only available if `method = "Inverse"`)

`df.Q`
: Degrees of freedom for test of heterogeneity / inconsistency.

`pval.Q`
: P-value for test of heterogeneity / inconsistency.

`I2`, `lower.I2`, `upper.I2`
: I-squared, lower and upper confidence limits (only available if `method = "Inverse"`).

`tau`
: Square-root of between-study variance (only available if `method = "Inverse"`).

`Q.heterogeneity`
: Overall heterogeneity statistic. (only available if `method = "Inverse"`)

`df.Q.heterogeneity`
: Degrees of freedom for test of overall heterogeneity.

`pval.Q.heterogeneity`
: P-value for test of overall heterogeneity.

`Q.inconsistency`
: Overall inconsistency statistic.

`df.Q.inconsistency`
: Degrees of freedom for test of overall inconsistency.

`pval.Q.inconsistency`
: P-value for test of overall inconsistency.

`A.matrix`
: Adjacency matrix (*n*x*n*).

`H.matrix`
: Hat matrix (*m*x*m*)

`n.matrix` *nxn* matrix with number of observations in direct comparisons.

`events.matrix` *nxn* matrix with number of events in direct comparisons.

`sm, method, level, level.ma`
As defined above.

`incr, allincr, addincr, allstudies, cc.pooled`
As defined above.

`common, random` As defined above.

`prediction, level.predict`
As defined above.

`reference.group, baseline.reference, all.treatments`
As defined above.

`seq, tau.preset, tol.multiarm, tol.multiarm.se`
As defined above.

`details.chkmultiarm, details.chkdata`
As defined above.

`sep.trts, nchar.trts`
As defined above.

`backtransf, title, warn, warn.deprecated`
As defined above.

`data` Data set (in contrast-based format).

`data.design` List with data in arm-based format (each list element corresponds to a single design).

`call` Function call.

`version` Version of R package netmeta used to create object.

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References

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Senn S, Gavini F, Magrez D, Scheen A (2013): Issues in performing a network meta-analysis. *Statistical Methods in Medical Research*, **22**, 169–89

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See Also

pairwise, netmeta

Examples

```
data(Dong2013)

# Only consider first ten studies (to reduce runtime of example)
#
first10 <- subset(Dong2013, id <= 10)

# Transform data from long arm-based format to contrast-based
# format. Argument 'sm' has to be used for odds ratio as summary
# measure; by default the risk ratio is used in the metabin
# function called internally.
#
p1 <- pairwise(treatment, death, randomized, studlab = id,
  data = first10, sm = "OR")

# Conduct Mantel-Haenszel network meta-analysis (without continuity
# correction)
#
nb1 <- netmetabin(p1, ref = "plac")
nb1

# Obtain the league table
#
netleague(nb1)

## Not run:
# Conduct Mantel-Haenszel network meta-analysis for the whole
# dataset
#
p2 <- pairwise(treatment, death, randomized, studlab = id,
  data = Dong2013, sm = "OR")
netmetabin(p2, ref = "plac")

# Conduct network meta-analysis using the non-central
# hypergeometric model (without continuity correction)
#
netmetabin(p2, ref = "plac", method = "NCH")

# Conduct Mantel-Haenszel network meta-analysis (with continuity
# correction of 0.5; include all studies)
#
netmetabin(p2, ref = "plac", cc.pooled = TRUE)

data(Gurusamy2011)

p3 <- pairwise(treatment, death, n, studlab = study,
  data = Gurusamy2011, sm = "OR")

# Conduct Mantel-Haenszel network meta-analysis (without continuity
# correction)
#
netmetabin(p3, ref = "cont")
```

```
## End(Not run)
```

`netpairwise` *Conduct pairwise meta-analyses for all comparisons with direct evidence in a network meta-analysis*

Description

Conduct pairwise meta-analyses for all comparisons with direct evidence in a network meta-analysis.

Usage

```
netpairwise(
  x,
  separate = FALSE,
  common = x$common,
  random = x$random,
  level = x$level,
  level.ma = x$level.ma,
  prediction = x$prediction,
  level.predict = x$level.predict,
  reference.group = x$reference.group,
  baseline.reference = x$baseline.reference,
  method.tau = x$method.tau,
  sep.trts = x$sep.trts,
  nchar.trts = x$nchar.trts,
  backtransf = x$backtransf,
  warn.deprecated = gs("warn.deprecated"),
  ...
)

## S3 method for class 'netpairwise'
print(x, ...)

## S3 method for class 'netpairwise'
summary(object, ...)

## S3 method for class 'summary.netpairwise'
print(x, ...)

## S3 method for class 'netpairwise'
forest(x, ...)

## S3 method for class 'netpairwise'
plot(x, ...)
```

Arguments

x	An object of class netmeta or netpairwise.
separate	A logical indicating whether results for pairwise comparisons should be printed as separate meta-analyses or as subgroups which is more concise.
common	A logical indicating whether a common effects network meta-analysis should be conducted.
random	A logical indicating whether a random effects network meta-analysis should be conducted.
level	The level used to calculate confidence intervals for individual comparisons.
level.ma	The level used to calculate confidence intervals for pooled estimates.
prediction	A logical indicating whether prediction intervals should be printed.
level.predict	The level used to calculate prediction intervals for a new study.
reference.group	Reference treatment.
baseline.reference	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa. This argument is only considered if reference.group has been specified.
method.tau	A character string indicating which method is used to estimate the between-study variance τ^2 and its square root τ. Either "DL", "REML", or "ML", can be abbreviated.
sep.trts	A character used in comparison names as separator between treatment labels.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names (see Details).
backtransf	A logical indicating whether results should be back transformed in printouts and forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (passed on to metagen or print functions and to catch deprecated arguments).
object	An object of class netpairwise.

Details

Conduct pairwise meta-analyses for all comparisons with direct evidence in a network meta-analysis. In contrast to netmeta and netsplit, unadjusted standard errors are used in the calculations and the between-study heterogeneity variance is allowed to differ between comparisons.

The R function metagen is called internally.

Value

Either a single metagen object with pairwise comparisons as subgroups or a list with metagen objects for each direct pairwise comparison.

Note

This function must not be confused with pairwise which can be used as a pre-processing step to convert data from arm-based to contrast-based format by calculating all pairwise comparisons within a study.

Author(s)

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See Also

netmeta, netsplit, pairwise

Examples

```
data(Senn2013)
oldsets <- settings.meta(digits = 2, digits.tau2 = 2, digits.tau = 2)

# Random effects model
#
net1 <- netmeta(TE, seTE, treat1.long, treat2.long, studlab,
  data = Senn2013, sm = "MD", common = FALSE)

# Calculate and print consise results for all pairwise
# meta-analyses
#
np1 <- netpairwise(net1)
np1
print(np1, details.method = FALSE)

## Not run:
forest(np1)

# Print detailed information for each pairwise comparison
#
np2 <- netpairwise(net1, separate = TRUE)
forest(np2)

## End(Not run)

settings.meta(oldsets)
```

---

netposet *Partial order of treatments in network meta-analysis*

Description

Partial order of treatments in network meta-analysis. The set of treatments in a network is called a partially ordered set (in short, a *poset*), if different outcomes provide different treatment ranking lists.

Usage

```
netposet(
  ...,
  outcomes,
  treatments,
  small.values,
  common,
  random,
  fixed,
  comb.fixed,
  comb.random
)

## S3 method for class 'netposet'
print(x, pooled = ifelse(x$random, "random", "common"), ...)
```

Arguments

`...`	See details.
`outcomes`	A character vector with outcome names.
`treatments`	A character vector with treatment names.
`small.values`	See details.
`common`	A logical indicating whether to show results for the common effects model.
`random`	A logical indicating whether to show results for the random effects model.
`fixed`	Ignored deprecated argument (replaced by `common`).
`comb.fixed`	Ignored deprecated argument (replaced by `common`).
`comb.random`	Ignored deprecated argument (replaced by `random`).
`x`	An object of class `netposet`.
`pooled`	A character string indicating whether Hasse diagram should be drawn for common (`"common"`) or random effects model (`"random"`). Can be abbreviated.

Details

In network meta-analysis, frequently different outcomes are considered which may each provide a different ordering of treatments. The concept of a partially ordered set (in short, a *poset*, Carlsen & Bruggemann, 2014) of treatments can be used to gain further insights in situations with apparently conflicting orderings. This implementation for rankings in network meta-analyis is described in Rücker & Schwarzer (2017).

In function `netposet`, argument `...{}` can be any of the following:

- arbitrary number of netrank objects providing P-scores;
- arbitrary number of netmeta objects;
- single ranking matrix with each column providing P-scores (Rücker & Schwarzer 2015) or SUCRA values (Salanti et al. 2011) for an outcome and rows corresponding to treatments.

Note, albeit in general a ranking matrix is not constrained to have values between 0 and 1, netposet stops with an error in this case as this function expects a matrix with P-scores or SUCRA values.

Argument outcomes can be used to label outcomes. If argument outcomes is missing,

- column names of the ranking matrix are used as outcome labels (if first argument is a ranking matrix and column names are available);
- capital letters 'A', 'B', ... are used as outcome labels and a corresponding warning is printed.

Argument treatments can be used to provide treatment labels if the first argument is a ranking matrix. If argument treatment is missing,

- row names of the ranking matrix are used as treatment labels (if available);
- letters 'a', 'b', ... are used as treatment labels and a corresponding warning is printed.

If argument ...{} consists of netmeta objects, netrank is called internally to calculate P-scores. In this case, argument small.values can be used to specify for each outcome whether small values are good or bad; see netrank. This argument is ignored for a ranking matrix and netrank objects.

Arguments common and random can be used to define whether results should be printed and plotted for common and random effects model. If netmeta and netrank objects are provided in argument ...{}, values for common and random within these objects are considered; if these values are not unique, argument common or random are set to TRUE.

In function print.netposet, argument ...{} is passed on to the printing function.

Value

An object of class netposet with corresponding print, plot, and hasse functions. The object is a list containing the following components:

P.common	Ranking matrix with rows corresponding to treatments and columns corresponding to outcomes (common effects model).
M0.common	Hasse matrix skipping unnecessary paths (common effects model).
M.common	"Full" Hasse matrix (common effects model).
O.common	Matrix with information about partial ordering (common effects model).
P.random	Ranking matrix with rows corresponding to treatments and columns corresponding to outcomes (random effects model).
M0.random	Hasse matrix skipping unnecessary paths (random effects model).
M.random	"Full" Hasse matrix (random effects model).
O.random	Matrix with information about partial ordering (random effects model).
small.values, common, random	As.defined above.
call	Function call.
version	Version of R package netmeta used to create object.

Author(s)

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Rücker G, Schwarzer G (2015): Ranking treatments in frequentist network meta-analysis works without resampling methods. *BMC Medical Research Methodology*, **15**, 58

Rücker G, Schwarzer G (2017): Resolve conflicting rankings of outcomes in network meta-analysis: Partial ordering of treatments. *Research Synthesis Methods*, **8**, 526–36

Salanti G, Ades AE, Ioannidis JP (2011): Graphical methods and numerical summaries for presenting results from multiple-treatment meta-analysis: an overview and tutorial. *Journal of Clinical Epidemiology*, **64**, 163–71

See Also

netmeta, netrank, plot.netrank, hasse, plot.netposet

Examples

```
## Not run:
# Use depression dataset
#
data(Linde2015)

# Define order of treatments
#
trts <- c("TCA", "SSRI", "SNRI", "NRI",
  "Low-dose SARI", "NaSSa", "rMAO-A", "Hypericum", "Placebo")

# Outcome labels
#
outcomes <- c("Early response", "Early remission")

# (1) Early response
#
p1 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(resp1, resp2, resp3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net1 <- netmeta(p1, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "bad")

# (2) Early remission
#
p2 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(remi1, remi2, remi3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
```

```
net2 <- netmeta(p2, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "bad")

# Partial order of treatment rankings (two outcomes)
#
po <- netposet(netrank(net1), netrank(net2), outcomes = outcomes)

# Hasse diagram
#
hasse(po)


#
# Outcome labels
#
outcomes <- c("Early response", "Early remission",
  "Lost to follow-up", "Lost to follow-up due to AEs",
  "Adverse events (AEs)")

# (3) Loss to follow-up
#
p3 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(loss1, loss2, loss3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net3 <- netmeta(p3, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "good")

# (4) Loss to follow-up due to adverse events
#
p4 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(loss.ae1, loss.ae2, loss.ae3), n = list(n1, n2, n3),
  studlab = id, data = subset(Linde2015, id != 55), sm = "OR")
#
net4 <- netmeta(p4, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "good")

# (5) Adverse events
#
p5 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(ae1, ae2, ae3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net5 <- netmeta(p5, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "good")

# Partial order of treatment rankings (all five outcomes)
#
po.ranks <- netposet(netrank(net1), netrank(net2),
  netrank(net3), netrank(net4), netrank(net5), outcomes = outcomes)

# Same result
#
```

```
po.nets <- netposet(net1, net2, net3, net4, net5,
  outcomes = outcomes)
#
all.equal(po.ranks, po.nets)

# Print matrix with P-scores (random effects model)
#
po.nets$P.random

# Hasse diagram for all outcomes (random effects model)
#
hasse(po.ranks)

# Hasse diagram for outcomes early response and early remission
#
po12 <- netposet(netrank(net1), netrank(net2),
  outcomes = outcomes[1:2])
hasse(po12)

# Scatter plot
#
oldpar <- par(pty = "s")
plot(po12)
par(oldpar)

## End(Not run)

# Example using ranking matrix with P-scores
#
# Ribassin-Majed L, Marguet S, Lee A.W., et al. (2017):
# What is the best treatment of locally advanced nasopharyngeal
# carcinoma? An individual patient data network meta-analysis.
# Journal of Clinical Oncology, 35, 498-505
#
outcomes <- c("OS", "PFS", "LC", "DC")
treatments <- c("RT", "IC-RT", "IC-CRT", "CRT",
  "CRT-AC", "RT-AC", "IC-RT-AC")
#
# P-scores (from Table 1)
#
pscore.os  <- c(15, 33, 63, 70, 96, 28, 45) / 100
pscore.pfs <- c( 4, 46, 79, 52, 94, 36, 39) / 100
pscore.lc  <- c( 9, 27, 47, 37, 82, 58, 90) / 100
pscore.dc  <- c(16, 76, 95, 48, 72, 32, 10) / 100
#
pscore.matrix <- data.frame(pscore.os, pscore.pfs, pscore.lc, pscore.dc)
rownames(pscore.matrix) <- treatments
colnames(pscore.matrix) <- outcomes
pscore.matrix
#
po <- netposet(pscore.matrix)
po12 <- netposet(pscore.matrix[, 1:2])
po
```

```
po12
#
hasse(po)
hasse(po12)
#
oldpar <- par(pty = "s")
plot(po12)
par(oldpar)
```

---

netrank *Frequentist method to rank treatments in network*

Description

Ranking treatments in frequentist network meta-analysis with and without resampling methods.

Usage

```
netrank(
  x,
  small.values = x$small.values,
  method,
  nsim,
  common = x$common,
  random = x$random,
  warn.deprecated = gs("warn.deprecated"),
  ...
)

## S3 method for class 'netrank'
print(
  x,
  common = x$common,
  random = x$random,
  sort = TRUE,
  digits = gs("digits.prop"),
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netmeta or rankogram.
small.values	A character string specifying whether small treatment effects indicate a beneficial ("good") or harmful ("bad") effect, can be abbreviated.
method	A character string specifying whether the "P-score" or "SUCRA" ranking metric will be calculated.

nsim	Number of simulations to calculate SUCRAs.
common	A logical indicating whether to print P-scores or SUCRAs for the common effects model.
random	A logical indicating whether to print P-scores or SUCRAs for the random effects model.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments passed on to print.data.frame function (used internally).
sort	A logical indicating whether printout should be sorted by decreasing P-score.
digits	Minimal number of significant digits, see print.default.

Details

Treatments are ranked based on a network meta-analysis. Ranking is performed by a ranking metric: P-score or SUCRA.

P-scores are based solely on the point estimates and standard errors of the network estimates. They measure the extent of certainty that a treatment is better than another treatment, averaged over all competing treatments (Rücker and Schwarzer 2015).

The Surface Under the Cumulative RAnking curve (SUCRA) is the rank of treatment *i* within the range of treatments, measured on a scale from 0 (worst) to 1 (best) (Salanti et al. 2011). A resampling method is used to calculate SUCRAs for frequentist network meta-analysis. The number of simulations is determine by argument nsim.

The interpretation of P-scores and SUCRAs is comparable.

The P-score of treatment *i* is defined as the mean of all 1 - P[*j*] where P[*j*] denotes the one-sided P-value of accepting the alternative hypothesis that treatment *i* is better than one of the competing treatments *j*. Thus, if treatment *i* is better than many other treatments, many of these P-values will be small and the P-score will be large. Vice versa, if treatment *i* is worse than most other treatments, the P-score is small.

The P-score of treatment *i* can be interpreted as the mean extent of certainty that treatment *i* is better than another treatment.

Value

An object of class netrank with corresponding print function. The object is a list containing the following components:

ranking.common	A named numeric vector with P-scores or SUCRAs for the common effects model.
Pmatrix.common	Numeric matrix based on pairwise one-sided p-values for the common effects model.
ranking.random	A named numeric vector with P-scores or SUCRAs for the random effects model.
Pmatrix.random	Numeric matrix based on pairwise one-sided p-values of the random effects model.

`small.values, method, x`
: As defined above.

`version`
: Version of R package netmeta used to create object.

Author(s)

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References

Rücker G, Schwarzer G (2017): Resolve conflicting rankings of outcomes in network meta-analysis: Partial ordering of treatments. *Research Synthesis Methods*, **8**, 526–36

Salanti G, Ades AE, Ioannidis JP (2011): Graphical methods and numerical summaries for presenting results from multiple-treatment meta-analysis: an overview and tutorial. *Journal of Clinical Epidemiology*, **64**, 163–71

See Also

netmeta, rankogram

Examples

```
data(Senn2013)

net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", random = FALSE)

nr1 <- netrank(net1)
nr1
print(nr1, sort = FALSE)

## Not run:
net2 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD")

nr2 <- netrank(net2)
nr2
print(nr2, sort = "common")
print(nr2, sort = FALSE)

## End(Not run)

## Not run:
net3 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD")

nr3 <- netrank(net3, method = "SUCRA", nsim = 100)
nr3
print(nr3, sort = "common")
print(nr3, sort = FALSE)
```

```
## End(Not run)
```

---

netsplit *Split direct and indirect evidence in network meta-analysis*

Description

Methods to split network estimates into the contribution of direct and indirect evidence and to test for local inconsistency in network meta-analysis.

Usage

```
netsplit(
  x,
  method,
  upper = TRUE,
  reference.group = x$reference.group,
  baseline.reference = x$baseline.reference,
  order = NULL,
  sep.trts = x$sep.trts,
  quote.trts = "",
  tol.direct = 5e-04,
  common = x$common,
  random = x$random,
  backtransf = x$backtransf,
  warn = FALSE,
  warn.deprecated = gs("warn.deprecated"),
  verbose = FALSE,
  ...
)

## S3 method for class 'netsplit'
print(
  x,
  common = x$x$common,
  random = x$x$random,
  show = "all",
  overall = TRUE,
  ci = FALSE,
  test = show %in% c("all", "with.direct", "both"),
  only.reference = FALSE,
  sortvar = NULL,
  nchar.trts = x$nchar.trts,
  digits = gs("digits"),
  digits.stat = gs("digits.stat"),
```

```
  digits.pval = gs("digits.pval"),
  digits.prop = max(gs("digits.pval") - 2, 2),
  text.NA = ".",
  backtransf = x$backtransf,
  scientific.pval = gs("scientific.pval"),
  big.mark = gs("big.mark"),
  legend = TRUE,
  indent = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netmeta or netsplit.
method	A character string indicating which method to split direct and indirect evidence is to be used. Either "Back-calculation" or "SIDDE", can be abbreviated. See Details.
upper	A logical indicating whether treatment comparisons should be selected from the lower or upper triangle of the treatment effect matrices (see list elements TE.common and TE.random in the netmeta object). Ignored if argument order is provided.
reference.group	Reference treatment. Ignored if argument order is provided.
baseline.reference	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment or vice versa. This argument is only considered if reference.group is not equal to "" and argumentorder is not provided.
order	A optional character or numerical vector specifying the order of treatments in comparisons.
sep.trts	A character string used in comparison names as separator between treatment labels, e.g., " vs ".
quote.trts	A character used to print around treatment labels.
tol.direct	A numeric defining the maximum deviation of the direct evidence proportion from 0 or 1 to classify a comparison as providing only indirect or direct evidence, respectively.
common	A logical indicating whether results for the common effects network meta-analysis should be printed.
random	A logical indicating whether results for the random effects network meta-analysis should be printed.
backtransf	A logical indicating whether printed results should be back transformed. For example, if backtransf = TRUE, results for sm = "OR" are printed as odds ratios rather than log odds ratios.
warn	A logical indicating whether warnings should be printed.

warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
verbose	A logical indicating whether progress information should be printed.
...	Additional arguments.
show	A character string indicating which comparisons should be printed (see Details).
overall	A logical indicating whether estimates from network meta-analyis should be printed in addition to direct and indirect estimates.
ci	A logical indicating whether confidence intervals should be printed in addition to treatment estimates.
test	A logical indicating whether results of a test comparing direct and indirect estimates should be printed.
only.reference	A logical indicating whether only comparisons with the reference group should be printed.
sortvar	An optional vector used to sort comparisons (must be of same length as the total number of comparisons).
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
digits	Minimal number of significant digits, see `print.default`.
digits.stat	Minimal number of significant digits for z-value of test of agreement between direct and indirect evidence, see `print.default`.
digits.pval	Minimal number of significant digits for p-value of test of agreement between direct and indirect evidence, see `print.default`.
digits.prop	Minimal number of significant digits for direct evidence proportions, see `print.default`.
text.NA	A character string specifying text printed for missing values.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
big.mark	A character used as thousands separator.
legend	A logical indicating whether a legend should be printed.
indent	A logical indicating whether items in the legend should be indented.

Details

A comparison of direct and indirect treatment estimates can serve as check for consistency of network meta-analysis (Dias et al., 2010).

This function provides two methods to derive indirect estimates:

- Separate Indirect from Direct Evidence (SIDE) using a back-calculation method. The *direct evidence proportion* as described in König et al. (2013) is used in the calculation of the indirect evidence;
- Separate Indirect from Direct Design Evidence (SIDDE) as described in Efthimiou et al. (2019).

Note, for the back-calculation method, indirect treatment estimates are already calculated in netmeta and this function combines and prints these estimates in a user-friendly way. Furthermore, this method is not available for the Mantel-Haenszel and non-central hypergeometric distribution approach implemented in netmetabin.

For the random-effects model, the direct treatment estimates are based on the common between-study variance τ^2 from the network meta-analysis, i.e. the square of list element x$tau.

Argument show determines which comparisons are printed:

"all"	All comparisons
"both"	Only comparisons contributing both direct and indirect evidence
"with.direct"	Comparisons providing direct evidence
"direct.only"	Comparisons providing only direct evidence
"indirect.only"	Comparisons providing only indirect evidence

The SIDDE approach can be compute-intensive in large networks. Crude information on the computation progress is printed for SIDDE if argument verbose is TRUE. In addition, computation times are printed if R package **tictoc** is installed.

Value

An object of class netsplit with corresponding print and forest functions. The object is a list containing the following components:

common, random	As defined above.
comparison	A vector with treatment comparisons.
prop.common, prop.random	A vector with direct evidence proportions (common / random effects model).
common, random	Results of network meta-analysis (common / random effects model), i.e., data frame with columns comparison, TE, seTE, lower, upper, z, and p.
direct.common, direct.random	Network meta-analysis results based on direct evidence (common / random effects model), i.e., data frame with columns comparison, TE, seTE, lower, upper, z, and p.
indirect.common, indirect.random	Network meta-analysis results based on indirect evidence (common / random effects model), i.e., data frame with columns comparison, TE, seTE, lower, upper, z, and p.
compare.common, compare.random	Comparison of direct and indirect evidence in network meta-analysis (common / random effects model), i.e., data frame with columns comparison, TE, seTE, lower, upper, z, and p.
sm	A character string indicating underlying summary measure
level.ma	The level used to calculate confidence intervals for pooled estimates.
tictoc	Computation times for SIDDE approach (if R package **tictoc** is installed).
version	Version of R package netmeta used to create object.

Author(s)

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References

Dias S, Welton NJ, Caldwell DM, Ades AE (2010): Checking consistency in mixed treatment comparison meta-analysis. *Statistics in Medicine*, **29**, 932–44

Efthimiou O, Rücker G, Schwarzer G, Higgins J, Egger M, Salanti G (2019): A Mantel-Haenszel model for network meta-analysis of rare events. *Statistics in Medicine*, 1–21, https://doi.org/10.1002/sim.8158

König J, Krahn U, Binder H (2013): Visualizing the flow of evidence in network meta-analysis and characterizing mixed treatment comparisons. *Statistics in Medicine*, **32**, 5414–29

Puhan MA, Schünemann HJ, Murad MH, et al. (2014): A GRADE working group approach for rating the quality of treatment effect estimates from network meta-analysis. *British Medical Journal*, **349**, g5630

See Also

forest.netsplit, netmeta, netmetabin, netmeasures

Examples

```
data(Woods2010)
#
p1 <- pairwise(treatment, event = r, n = N,
  studlab = author, data = Woods2010, sm = "OR")
#
net1 <- netmeta(p1)
#
print(netsplit(net1), digits = 2)

## Not run:
print(netsplit(net1), digits = 2,
  backtransf = FALSE, common = FALSE)

# Sort by increasing number of studies in direct comparisons
print(netsplit(net1), digits = 2, sortvar = k)
# Sort by decreasing number of studies in direct comparisons
print(netsplit(net1), digits = 2, sortvar = -k)

# Sort by increasing evidence proportion under common effects model
print(netsplit(net1), digits = 2, sortvar = prop.common)
# Sort by decreasing evidence proportion under common effects model
print(netsplit(net1), digits = 2, sortvar = -prop.common)

# Sort by decreasing evidence proportion under common effects model
# and number of studies
print(netsplit(net1), digits = 2, sortvar = cbind(-prop.common, -k))

data(Senn2013)
```

```
#
net2 <- netmeta(TE, seTE, treat1.long, treat2.long,
  studlab, data = Senn2013)
#
print(netsplit(net2), digits = 2)
# Layout of Puhan et al. (2014), Table 1
print(netsplit(net2), digits = 2, ci = TRUE, test = FALSE)

data(Dong2013)
p3 <- pairwise(treatment, death, randomized, studlab = id,
  data = Dong2013, sm = "OR")
net3 <- netmetabin(p3)
netsplit(net3)

## End(Not run)
```

---

nettable *Table with network meta-analysis results*

Description

Construct a table with network, direct and indirect estimates from one or more network meta-analyses.

Usage

```
nettable(
  ...,
  name = NULL,
  method = NULL,
  order = NULL,
  common,
  random,
  upper = TRUE,
  reference.group = NULL,
  baseline.reference = NULL,
  backtransf = NULL,
  digits = gs("digits"),
  digits.I2 = gs("digits.I2"),
  digits.pval = gs("digits.pval"),
  scientific.pval = gs("scientific.pval"),
  zero.pval = gs("zero.pval"),
  JAMA.pval = gs("JAMA.pval"),
  big.mark = gs("big.mark"),
  text.NA = ".",
  bracket = gs("CIbracket"),
  separator = gs("CIseparator"),
```

```
  lower.blank = gs("CIlower.blank"),
  upper.blank = gs("CIupper.blank"),
  tol.direct = 5e-04,
  writexl = !missing(path),
  path = "nettable.xlsx",
  overwrite = FALSE,
  warn = FALSE,
  verbose = FALSE
)

## S3 method for class 'nettable'
print(x, common = x$x$common, random = x$x$random, ...)
```

Arguments

...	Any number of network meta-analysis objects or a single list with network meta-analyses.
name	An optional character vector providing descriptive names for network meta-analysis objects.
method	A character string indicating which method to split direct and indirect evidence is to be used. Either "Back-calculation" or "SIDDE", can be abbreviated. See Details.
order	A optional character or numerical vector specifying the order of treatments in comparisons.
common	A logical indicating whether table for the common effects network meta-analysis should be printed.
random	A logical indicating whether table for the random effects network meta-analysis should be printed.
upper	A logical indicating whether treatment comparisons should be selected from the lower or upper triangle of the treatment effect matrices (see list elements TE.common and TE.random in the netmeta object). Ignored if argument order is provided.
reference.group	Reference treatment. Ignored if argument order is provided.
baseline.reference	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment or vice versa. This argument is only considered if reference.group is not equal to "" and argumentorder is not provided.
backtransf	A logical indicating whether printed results should be back transformed. For example, if backtransf = TRUE, results for sm = "OR" are printed as odds ratios rather than log odds ratios.
digits	Minimal number of significant digits, see print.default.
digits.I2	Minimal number of significant digits for I-squared statistic, see print.default.
digits.pval	Minimal number of significant digits for p-value of test of agreement between direct and indirect evidence, see print.default.

`scientific.pval`
: A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.

`zero.pval`
: A logical specifying whether p-values should be printed with a leading zero.

`JAMA.pval`
: A logical specifying whether p-values for test of overall effect should be printed according to JAMA reporting standards.

`big.mark`
: A character used as thousands separator.

`text.NA`
: A character string specifying text printed for missing values.

`bracket`
: A character with bracket symbol to print lower confidence interval: "[", "(", "{", "".

`separator`
: A character string with information on separator between lower and upper confidence interval.

`lower.blank`
: A logical indicating whether blanks between left bracket and lower confidence limit should be printed.

`upper.blank`
: A logical indicating whether blanks between separator and upper confidence limit should be printed.

`tol.direct`
: A numeric defining the maximum deviation of the direct evidence proportion from 0 or 1 to classify a comparison as providing only indirect or direct evidence, respectively.

`writexl`
: A logical indicating whether an Excel file should be created (R package **writexl** must be available).

`path`
: A character string specifying the filename of the Excel file.

`overwrite`
: A logical indicating whether an existing Excel file should be overwritten.

`warn`
: A logical indicating whether warnings should be printed.

`verbose`
: A logical indicating whether progress information should be printed.

`x`
: An object of class `nettable`.

Details

Construct a table with network, direct and indirect estimates from one or more network meta-analyses. The table looks very similar to the statistical part of a GRADE table for a network meta-analysis (Puhan et al., 2014).

By default, an R object with the network tables is generated. Alternatively, an Excel file is created if argument `writexl = TRUE`.

Two methods to derive indirect estimates are available:

- Separate Indirect from Direct Evidence (SIDE) using a back-calculation method. The *direct evidence proportion* as described in König et al. (2013) is used in the calculation of the indirect evidence;
- Separate Indirect from Direct Design Evidence (SIDDE) as described in Efthimiou et al. (2019).

Note, for the back-calculation method, indirect treatment estimates are already calculated in netmeta and this function combines and prints these estimates in a user-friendly way. Furthermore, this method is not available for the Mantel-Haenszel and non-central hypergeometric distribution approach implemented in netmetabin.

For the random-effects model, the direct treatment estimates are based on the common between-study variance τ^2 from the network meta-analysis, i.e. the square of list element x$tau.

The SIDDE approach can be compute-intensive in large networks. Crude information on the computation progress is printed for SIDDE if argument verbose is TRUE.

Value

An object of class nettable with corresponding print function if argument writexl = FALSE. The object is a list containing the network tables in list elements 'common' and 'random'. An Excel file is created if writexl = TRUE. In this case, NULL is returned in R.

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References

Efthimiou O, Rücker G, Schwarzer G, Higgins J, Egger M, Salanti G (2019): A Mantel-Haenszel model for network meta-analysis of rare events. *Statistics in Medicine*, 1–21, https://doi.org/10.1002/sim.8158

König J, Krahn U, Binder H (2013): Visualizing the flow of evidence in network meta-analysis and characterizing mixed treatment comparisons. *Statistics in Medicine*, **32**, 5414–29

Puhan MA, Schünemann HJ, Murad MH, et al. (2014): A GRADE working group approach for rating the quality of treatment effect estimates from network meta-analysis. *British Medical Journal*, **349**, g5630

See Also

netsplit, netmeta, netmetabin, netmeasures

Examples

```
data(Woods2010)
#
p1 <- pairwise(treatment, event = r, n = N,
  studlab = author, data = Woods2010, sm = "OR")
#
net1 <- netmeta(p1)
#
nt1 <- nettable(net1, digits = 2)
nt1
print(nt1, common = FALSE)
print(nt1, random = FALSE)

## Not run:
# Create a CSV file with network table from random effects model
#
```

```
table1 <- nettable(net1, digits = 2, bracket = "(", separator = " to ")
#
write.table(table1$random, file = "table1-random.csv",
  row.names = FALSE, col.names = TRUE, sep = ",")
#
# Create Excel files with network tables
# (if R package writexl is available)
#
nettable(net1, digits = 2, bracket = "(", separator = " to ",
         path = tempfile(fileext = ".xlsx"))

## End(Not run)
```

`pairwise` *Transform meta-analysis data from two arm-based formats into contrast-based format*

Description

This function transforms data that are given in wide or long arm-based format (e.g. input format for WinBUGS) to a contrast-based format that is needed as input to R function `netmeta`. The function can transform data with binary, continuous, or generic outcomes as well as incidence rates from arm-based to contrast-based format.

Usage

```
pairwise(
  treat,
  event,
  n,
  mean,
  sd,
  TE,
  seTE,
  time,
  data = NULL,
  studlab,
  incr = gs("incr"),
  allincr = gs("allincr"),
  addincr = gs("addincr"),
  allstudies = gs("allstudies"),
  reference.group,
  keep.all.comparisons,
  warn = FALSE,
  ...
)
```

Arguments

treat	A list or vector with treatment information for individual treatment arms (see Details).
event	A list or vector with information on number of events for individual treatment arms (see Details).
n	A list or vector with information on number of observations for individual treatment arms (see Details).
mean	A list or vector with estimated means for individual treatment arms (see Details).
sd	A list or vector with information on the standard deviation for individual treatment arms (see Details).
TE	A list or vector with estimated treatment effects for individual treatment arms (see Details).
seTE	A list or vector with standard errors of estimated treatment effect for individual treatment arms (see Details).
time	A list or vector with information on person time at risk for individual treatment arms (see Details).
data	An optional data frame containing the study information.
studlab	A vector with study labels (optional).
incr	A numerical value which is added to cell frequencies for studies with a zero cell count, see Details.
allincr	A logical indicating if incr is added to cell frequencies of all studies if at least one study has a zero cell count. If FALSE (default), incr is added only to cell frequencies of studies with a zero cell count.
addincr	A logical indicating if incr is added to cell frequencies of all studies irrespective of zero cell counts.
allstudies	A logical indicating if studies with zero or all events in two treatment arms are to be included in the meta-analysis (applies only if sm is equal to "RR" or "OR").
reference.group	Reference treatment (first treatment is used if argument is missing).
keep.all.comparisons	A logical indicating whether all pairwise comparisons or only comparisons with the study-specific reference group should be kept ('basic parameters').
warn	A logical indicating whether warnings should be printed (e.g., if studies are excluded due to only providing a single treatment arm).
...	Additional arguments passed-through to the functions to calculate effects.

Details

R function netmeta expects data in a **contrast-based format**, where each row corresponds to a comparison of two treatments and contains a measure of the treatment effect comparing two treatments with standard error, labels for the two treatments and an optional study label. In contrast-based format, a three-arm study contributes three rows with treatment comparison and corresponding standard error for pairwise comparison *A* vs *B*, *A* vs *C*, and *B* vs *C* whereas a four-arm study contributes six rows / pairwise comparisons: *A* vs *B*, *A* vs *C*, ..., *C* vs *D*.

Other programs for network meta-analysis in WinBUGS and Stata require data in an *arm-based* format, i.e. treatment estimate for each treatment arm instead of a difference of two treatments. A common **(wide) arm-based format** consists of one data row per study, containing treatment and other necessary information for all study arms. For example, a four-arm study contributes one row with four treatment estimates and corresponding standard errors for treatments *A*, *B*, *C*, and *D*. Another possible arm-based format is a long format where each row corresponds to a single study arm. Accordingly, in the **long arm-based format** a study contributes as many rows as treatments considered in the study.

The pairwise function transforms data given in (wide or long) arm-based format into the contrast-based format which consists of *pairwise* comparisons and is needed as input to the `netmeta` function.

The pairwise function can transform data with binary outcomes (using the `metabin` function from R package meta), continuous outcomes (`metacont` function), incidence rates (`metainc` function), and generic outcomes (`metagen` function). Depending on the outcome, the following arguments are mandatory:

- treat, event, n (see `metabin`);
- treat, n, mean, sd (see `metacont`);
- treat, event, time (see `metainc`);
- treat, TE, seTE (see `metagen`).

Argument `treat` is mandatory to identify the individual treatments. The other arguments contain outcome specific data. These arguments must be either lists (wide arm-based format, i.e., one row per study) or vectors (long arm-based format, i.e., multiple rows per study) of the same length.

For the wide arm-based format, each list consists of as many vectors of the same length as the multi-arm study with the largest number of treatments. If a single multi-arm study has five arms, five vectors have to be provided for each lists. Two-arm studies have entries with `NA` for the third and subsequent vectors. Each list entry is a vector with information for each individual study; i.e., the length of this vector corresponds to the total number of studies incorporated in the network meta-analysis. Typically, list elements are part of a data frame (argument `data`, optional); see Examples. An optional vector with study labels can be provided which can be part of the data frame.

In the long arm-based format, argument `studlab` is mandatory to identify rows contributing to individual studies.

Additional arguments for meta-analysis functions can be provided using argument '...'. The most important argument is `sm` defining the summary measure. More information on this and other arguments is given in the help pages of R functions `metabin`, `metacont`, `metainc`, and `metagen`, respectively.

For standardised mean differences (argument `sm = "SMD"`), equations (4) and (5) in Crippa & Orsini (2016) are used to calculated SMDs and standard errors. These equations guarantee consistent SMDs and standard errors for multi-arm studies. Note, the summary measure is actually Cohen's d as Hedges' g is not consistent in multi-arm studies.

For binary outcomes, 0.5 is added to all cell frequencies (odds ratio) or only the number of events (risk ratio) for studies with a zero cell count. For odds ratio and risk ratio, treatment estimates and standard errors are only calculated for studies with zero or all events in both groups if `allstudies` is `TRUE`. This continuity correction is used both to calculate individual study results with confidence

limits and to conduct meta-analysis based on the inverse variance method. For the risk difference, 0.5 is only added to all cell frequencies to calculate the standard error.

For incidence rates, 0.5 is added to all cell frequencies for the incidence rate ratio as summary measure. For the incidence risk difference, 0.5 is only added to all cell frequencies to calculate the standard error.

The value of pairwise is a data frame with as many rows as there are pairwise comparisons. For each study with *p* treatments, *p*(p-1) / 2* contrasts are generated. Each row contains the treatment effect (`TE`), its standard error (`seTE`), the treatments compared ((`treat1`), (`treat2`)) and the study label ((`studlab`)). Further columns are added according to type of data.

All variables from the original dataset are also part of the output dataset. If data are provided in the long arm-based format, the value of a variable can differ between treatment arms; for example, the mean age or percentage of women in the treatment arm. In this situation, two variables instead of one variable will be included in the output dataset. The values "1" and "2" are added to the names for these variables, e.g. "mean.age1" and "mean.age2" for the mean age.

In general, any variable names in the original dataset that are identical to the main variable names (i.e., "TE", "seTE", ...) will be renamed to variable names with ending ".orig".

A reduced data set with basic comparisons (Rücker & Schwarzer, 2014) can be generated using argument `keep.all.comparisons = FALSE`. Furthermore, the reference group for the basic comparisons can be specified with argument `reference.group`.

Value

A data frame with the following columns:

`TE`	Treatment estimate comparing treatment 'treat1' and 'treat2'.
`seTE`	Standard error of treatment estimate.
`studlab`	Study labels.
`treat1`	First treatment in comparison.
`treat2`	Second treatment in comparison.
`event1`	Number of events for first treatment arm (for metabin and metainc).
`event2`	Number of events for second treatment arm (for metabin and metainc).
`n1`	Number of observations for first treatment arm (for metabin and metacont).
`n2`	Number of observations for second treatment arm (for metabin and metacont).
`mean1`	Estimated mean for first treatment arm (for metacont).
`mean2`	Estimated mean for second treatment arm (for metacont).
`sd1`	Standard deviation for first treatment arm (for metacont).
`sd2`	Standard deviation for second treatment arm (for metacont).
`TE1`	Estimated treatment effect for first treatment arm (for metagen).
`TE2`	Estimated treatment effect for second treatment arm (for metagen).
`seTE1`	Standard error of estimated treatment effect for first treatment arm (for metagen).
`seTE2`	Standard error of estimated treatment effect for second treatment arm (for metagen).

time1	Person time at risk for first treatment arm (for metainc).
time2	Person time at risk for second treatment arm (for metainc).

All variables from the original dataset are also part of the output dataset; see Details.

Note

This function must not be confused with netpairwise which can be used to conduct pairwise meta-analyses for all comparisons with direct evidence in a network meta-analysis.

Author(s)

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References

Crippa A, Orsini N (2016): Dose-response meta-analysis of differences in means. *BMC Medical Research Methodology*, **16**:91.

See Also

netmeta, metacont, metagen, metabin, metainc, netgraph.netmeta, pairwise

Examples

```
# Example using continuous outcomes (internal call of function
# metacont)
#
data(Franchini2012)
# Transform data from arm-based format to contrast-based format
p1 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = Franchini2012, studlab = Study)
p1

## Not run:
# Conduct network meta-analysis
#
net1 <- netmeta(p1)
net1

# Draw network graphs
#
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.5,
  thickness = "se.common")
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.5,
  plastic = TRUE, thickness = "se.common",
  iterate = TRUE)
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.5,
  plastic = TRUE, thickness = "se.common",
  iterate = TRUE, start = "eigen")
```

```
# Example using generic outcomes (internal call of function
# metagen)
#
# Calculate standard error for means y1, y2, y3
Franchini2012$se1 <- with(Franchini2012, sqrt(sd1^2 / n1))
Franchini2012$se2 <- with(Franchini2012, sqrt(sd2^2 / n2))
Franchini2012$se3 <- with(Franchini2012, sqrt(sd3^2 / n3))
# Transform data from arm-based format to contrast-based format
# using means and standard errors (note, argument 'sm' has to be
# used to specify that argument 'TE' is a mean difference)
p2 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  TE = list(y1, y2, y3), seTE = list(se1, se2, se3),
  n = list(n1, n2, n3),
  data = Franchini2012, studlab = Study,
  sm = "MD")
p2

# Compare pairwise objects p1 (based on continuous outcomes) and p2
# (based on generic outcomes)
#
all.equal(p1[, c("TE", "seTE", "studlab", "treat1", "treat2")],
  p2[, c("TE", "seTE", "studlab", "treat1", "treat2")])

# Same result as network meta-analysis based on continuous outcomes
# (object net1)
net2 <- netmeta(p2)
net2

# Example with binary data
#
data(smokingcessation)
# Transform data from arm-based format to contrast-based format
# (interal call of metabin function). Argument 'sm' has to be used
# for odds ratio as risk ratio (sm = "RR") is default of metabin
# function.
#
p3 <- pairwise(list(treat1, treat2, treat3),
  list(event1, event2, event3), list(n1, n2, n3),
  data = smokingcessation,
  sm = "OR")
p3

# Conduct network meta-analysis
#
net3 <- netmeta(p3)
net3

# Example with incidence rates
#
data(dietaryfat)

# Transform data from arm-based format to contrast-based format
```

```
#
p4 <- pairwise(list(treat1, treat2, treat3),
  list(d1, d2, d3), time = list(years1, years2, years3),
  studlab = ID,
  data = dietaryfat)
p4

# Conduct network meta-analysis using incidence rate ratios (sm =
# "IRR"). Note, the argument 'sm' is not necessary as this is the
# default in R function metainc called internally.
#
net4 <- netmeta(p4, sm = "IRR")
summary(net4)

# Example with long data format
#
data(Woods2010)

# Transform data from long arm-based format to contrast-based
# format Argument 'sm' has to be used for odds ratio as summary
# measure; by default the risk ratio is used in the metabin
# function called internally.
#
p5 <- pairwise(treatment, event = r, n = N,
  studlab = author, data = Woods2010, sm = "OR")
p5

# Conduct network meta-analysis
net5 <- netmeta(p5)
net5

## End(Not run)
```

`plot.netposet` *Scatter plot or biplot showing partially order of treatment ranks*

Description

This function generates a scatter plot or biplot of P-scores with an overlay describing partial order of treatment ranks.

Usage

```
## S3 method for class 'netposet'
plot(
  x,
  plottype = "scatter",
  pooled = ifelse(x$random, "random", "common"),
  dim = "2d",
```

```
  sel.x = 1,
  sel.y = 2,
  sel.z = 3,
  cex = 1,
  col = "black",
  cex.text = cex,
  col.text = col,
  adj.x = 0,
  adj.y = 1,
  offset.x = 0.005,
  offset.y = -0.005,
  pch = NULL,
  cex.points = cex,
  col.points = col,
  col.lines = "black",
  lty.lines = 1,
  lwd.lines = 1,
  arrows = FALSE,
  length = 0.05,
  grid = TRUE,
  col.grid = "gray",
  lty.grid = 2,
  lwd.grid = 1,
  ...
)
```

Arguments

x	An object of class netmeta (mandatory).
plottype	A character string indicating whether a scatter plot or biplot should be produced, either "scatter" or "biplot". Can be abbreviated.
pooled	A character string indicating whether scatter plot should be drawn for common ("common") or random effects model ("random"). Can be abbreviated.
dim	A character string indicating whether a 2- or 3-dimensional plot should be produced, either "2d" or "3d". Can be abbreviated.
sel.x	A numeric specifying number of outcome to use for the x-axis in a scatterplot (argument is not considered for a biplot).
sel.y	A numeric specifying number of outcome to use for the y-axis in a scatterplot (argument is not considered for a biplot).
sel.z	A numeric specifying number of outcome to use for the z-axis in a scatterplot (argument is not considered for a biplot).
cex	The magnification to be used for treatment labels and points.
col	Colour(s) of treatment labels and points.
cex.text	The magnification to be used for treatment labels.
col.text	Colour(s) of treatment labels.

adj.x	Value(s) in [0, 1] to specify adjustment of treatment labels on x-axis (only considered in 2-D plots); see text.
adj.y	Value(s) in [0, 1] to specify adjustment of treatment labels on y-axis (only considered in 2-D plots); see text.
offset.x	Offset(s) of treatment labels on x-axis (only considered in 2-D plots).
offset.y	Offset(s) of treatment labels on y-axis (only considered in 2-D plots).
pch	Plot symbol(s) for points; no points printed if equal to NULL.
cex.points	Magnification(s) to be used for points.
col.points	Colour(s) of points.
col.lines	Line colour.
lty.lines	Line type.
lwd.lines	Line width.
arrows	A logical indicating whether arrows should be printed (only considered in 2-D plots).
length	Length of arrows; see arrows.
grid	A logical indicating whether grid lines should be added to plot.
col.grid	Colour of grid lines.
lty.grid	Line type of grid lines.
lwd.grid	Line width of grid lines.
...	Additional graphical arguments.

Details

By default (arguments plottype = "scatter" and dim = "2d"), a scatter plot is created showing P-scores (see netrank) for the first two outcomes considered in the generation of a partially ordered set of treatment ranks (using netposet). In addition to the P-scores, the partially order of treatment ranks is shown as lines connecting treatments which is analogous to a Hasse diagram. If argument dim = "3d"), a 3-D scatter plot is generated showing P-scores for the first three outcomes.

To overcome the restriction of two or three dimension, a biplot (Gabriel, 1971) can be generated using argument plottype = "biplot". This is essentially a scatter plot using the first two (dim = "2d") or three (dim = "3d") components in a principal components analysis (using prcomp). Note, if only two / three outcomes are considered in a netposet object, a 2-D / 3-D scatter plot is generated instead of a biplot as a principal component analysis is superfluous in such a situation.

Arguments sel.x and sel.y can be used to select different outcomes to show on x- and y-axis in a 2-D scatter plot; argument sel.z can be used accordingly in a 3-D scatter plot. These arguments are ignored for a biplot.

Note, in order to generate 3-D plots (argument dim = "3d"), R package **rgl** is necessary. Note, under macOS the X.Org X Window System must be available (see https://www.xquartz.org).

Author(s)

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References

Carlsen L, Bruggemann R (2014): Partial order methodology: a valuable tool in chemometrics. *Journal of Chemometrics*, **28**, 226–34

Gabriel KR (1971): The biplot graphic display of matrices with application to principal component analysis. *Biometrika*, **58**, 453–67

See Also

netposet, hasse, netrank, netmeta

Examples

```
## Not run:
data(Linde2015)

# Define order of treatments
#
trts <- c("TCA", "SSRI", "SNRI", "NRI",
  "Low-dose SARI", "NaSSa", "rMAO-A", "Hypericum", "Placebo")
#
# Outcome labels
#
outcomes <- c("Early response", "Early remission")

# (1) Early response
#
p1 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(resp1, resp2, resp3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net1 <- netmeta(p1, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "bad")

# (2) Early remission
#
p2 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(remi1, remi2, remi3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net2 <- netmeta(p2, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "bad")

# Partial order of treatment rankings
#
po2 <- netposet(netrank(net1), netrank(net2), outcomes = outcomes)

# Scatter plot
#
plot(po2)

# Same scatter plot as only two outcomes considered in netposet()
```

```
#
plot(po2, "biplot")


# Consider three outcomes
#
# Outcome labels
#
outcomes <- c("Early response", "Early remission", "Lost to follow-up")

# (3) Loss to follow-up
#
p3 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(loss1, loss2, loss3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net3 <- netmeta(p3, common = FALSE,
  seq = trts, ref = "Placebo", small.values = "good")

# Partial order of treatment rankings (with three outcomes)
#
po3 <- netposet(netrank(net1), netrank(net2), netrank(net3),
  outcomes = outcomes)

# Hasse diagram
#
hasse(po3)

# Scatter plot
#
plot(po3)

# Biplot (reverse limits of y-axis as biplot is upside down)
#
plot(po3, "bi", xlim = c(-1, 1.7), ylim = c(2.5, -2.5))

## End(Not run)
```

`plot.netrank` *Plot treatment ranking(s) of network meta-analyses*

Description

Produce an image plot of treatment ranking(s) generated with R function `netrank`.

Usage

```
## S3 method for class 'netrank'
plot(
```

```
  ...,
  name,
  common,
  random,
  seq,
  low = "red",
  mid = "yellow",
  high = "green",
  col = "black",
  main,
  main.size = 14,
  main.col = col,
  main.face = "bold",
  legend = TRUE,
  axis.size = 12,
  axis.col = col,
  axis.face = "plain",
  na.value = "grey50",
  angle = 45,
  hjust.x = 1,
  vjust.x = 1,
  hjust.y = 1,
  vjust.y = 0,
  nchar.trts,
  digits = 3,
  fixed,
  comb.fixed,
  comb.random,
  warn.deprecated = gs("warn.deprecated")
)
```

Arguments

...	A single netrank object or a list of netrank objects.
name	An optional character vector providing descriptive names for the network meta-analysis objects.
common	A logical indicating whether results for the common effects model should be plotted.
random	A logical indicating whether results for the random effects model should be plotted.
seq	A character or numerical vector specifying the sequence of treatments on the x-axis.
low	A character string defining the colour for a P-score of 0, see scale_fill_gradient2.
mid	A character string defining the colour for a P-score of 0.5, see scale_fill_gradient2.
high	A character string defining the colour for a P-score of 1, see scale_fill_gradient2.
col	Colour of text.

main	Title.
main.size	Font size of title, see element_text.
main.col	Colour of title, see element_text.
main.face	Font face of title, see element_text.
legend	A logical indicating whether a legend should be printed.
axis.size	Font size of axis text, see element_text.
axis.col	Colour of axis text, see element_text.
axis.face	Font face of axis text, see element_text.
na.value	Colour for missing values, see scale_fill_gradient2.
angle	Angle for text on x-axis, see element_text.
hjust.x	A numeric between 0 and 1 with horizontal justification of text on x-axis, see element_text.
vjust.x	A numeric between 0 and 1 with vertical justification of text on x-axis, see element_text.
hjust.y	A numeric between 0 and 1 with horizontal justification of text on y-axis, see element_text.
vjust.y	A numeric between 0 and 1 with vertical justification of text on y-axis, see element_text.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
digits	Minimal number of significant digits, see print.default.
fixed	Deprecated argument (replaced by 'common').
comb.fixed	Deprecated argument (replaced by 'common').
comb.random	Deprecated argument (replaced by 'random').
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.

Details

This function produces an image plot of network rankings (Palpacuer et al., 2018, Figure 4). Note, a scatter plot of two network rankings can be generated with plot.netposet.

By default, treatments are ordered by decreasing P-scores of the first network meta-analysis object. Argument seq can be used to specify a differenct treatment order.

Value

A ggplot2 object or NULL if no ranking was conducted.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Clément Palpacuer <clementpalpacuer@gmail.com>

References

Palpacuer C, Duprez R, Huneau A, Locher C, Boussageon R, Laviolle B, et al. (2018): Pharmacologically controlled drinking in the treatment of alcohol dependence or alcohol use disorders: a systematic review with direct and network meta-analyses on nalmefene, naltrexone, acamprosate, baclofen and topiramate. *Addiction*, **113**, 220–37

See Also

netrank, netmeta, netposet, hasse

Examples

```
## Not run:
# Use depression dataset
#
data(Linde2015)

# Define order of treatments
#
trts <- c("TCA", "SSRI", "SNRI", "NRI",
  "Low-dose SARI", "NaSSa", "rMAO-A", "Hypericum", "Placebo")

# Outcome labels
#
outcomes <- c("Early response", "Early remission")

# (1) Early response
#
p1 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(resp1, resp2, resp3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net1 <- netmeta(p1, common = FALSE,
  seq = trts, ref = "Placebo")

# (2) Early remission
#
p2 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(remi1, remi2, remi3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net2 <- netmeta(p2, common = FALSE,
  seq = trts, ref = "Placebo")

# Image plot of treatment rankings (two outcomes)
#
plot(netrank(net1, small.values = "bad"),
  netrank(net2, small.values = "bad"),
  name = outcomes, digits = 2)


# Outcome labels
```

```
#
outcomes <- c("Early response", "Early remission",
  "Lost to follow-up", "Lost to follow-up due to AEs",
  "Adverse events (AEs)")

# (3) Loss to follow-up
#
p3 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(loss1, loss2, loss3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net3 <- netmeta(p3, common = FALSE, seq = trts, ref = "Placebo")

# (4) Loss to follow-up due to adverse events
#
p4 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(loss.ae1, loss.ae2, loss.ae3), n = list(n1, n2, n3),
  studlab = id, data = subset(Linde2015, id != 55), sm = "OR")
#
net4 <- netmeta(p4, common = FALSE, seq = trts, ref = "Placebo")

# (5) Adverse events
#
p5 <- pairwise(treat = list(treatment1, treatment2, treatment3),
  event = list(ae1, ae2, ae3), n = list(n1, n2, n3),
  studlab = id, data = Linde2015, sm = "OR")
#
net5 <- netmeta(p5, common = FALSE, seq = trts, ref = "Placebo")

# Image plot of treatment rankings (two outcomes)
#
plot(netrank(net1, small.values = "bad"),
  netrank(net2, small.values = "bad"),
  netrank(net3, small.values = "good"),
  netrank(net4, small.values = "good"),
  netrank(net5, small.values = "good"),
  name = outcomes, digits = 2)

## End(Not run)
```

`plot.rankogram` *Plot rankograms*

Description

This function produces a rankogram, i.e., an image plot of ranking probabilities for all treatments.

Usage

```
## S3 method for class 'rankogram'
plot(
  x,
  type = if (cumulative.rankprob) "step" else "bar",
  pooled = ifelse(x$random, "random", "common"),
  sort = TRUE,
  trts,
  cumulative.rankprob = x$cumulative.rankprob,
  ylim,
  ylab,
  nchar.trts = x$nchar.trts,
  ...
)
```

Arguments

x	An object of class rankogram.
type	A character string specifying whether a "bar" chart, a "line" graph, or "step" functions should be drawn. Can be abbreviated.
pooled	A character string indicating whether results for the common ("common") or random effects model ("random") should be plotted. Can be abbreviated.
sort	A logical indicating whether treatments should be sorted by decreasing SUCRAs.
trts	Treatment(s) to show in rankogram.
cumulative.rankprob	A logical indicating whether cumulative ranking probabilites should be shown.
ylim	The y limits (min, max) of the plot.
ylab	A label for the y-axis.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
...	Additional graphical arguments (ignored at the moment).

Details

This function produces an image plot of (cumulative) ranking probabilities for all treatments as a bar graph, a line graph or as step functions (argument type).

By default (argument pooled), results for the random effects model are shown if a network meta-analysis was conducted for both the common and random effects model.

Treatments are sorted according to their mean effects if argument sort = TRUE (default). A subset of treatments can be specified using argument trts.

Cumulative ranking probabilites are shown if cumulative.rankprob = TRUE. By default, step functions are shown for cumulative ranking probabilites.

Author(s)

Theodoros Papakonstantinou <dev@tpapak.com>, Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Salanti G, Ades AE, Ioannidis JP (2011): Graphical methods and numerical summaries for presenting results from multiple-treatment meta-analysis: an overview and tutorial. *Journal of Clinical Epidemiology*, **64**, 163–71

See Also

rankogram

Examples

```
data(Woods2010)
p1 <- pairwise(treatment, event = r, n = N, studlab = author,
  data = Woods2010, sm = "OR")
net1 <- netmeta(p1, small.values = "good")

ran1 <- rankogram(net1, nsim = 100)
ran1

plot(ran1)
plot(ran1, type = "l")
plot(ran1, cumulative.rankprob = TRUE)
```

---

print.decomp.design *Print method for objects of class decomp.design*

Description

Print method for objects of class decomp.design.

Usage

```
## S3 method for class 'decomp.design'
print(
  x,
  digits.Q = gs("digits.Q"),
  showall = FALSE,
  digits.pval.Q = gs("digits.pval.Q"),
  digits.tau2 = gs("digits.tau2"),
  scientific.pval = gs("scientific.pval"),
  big.mark = gs("big.mark"),
  nchar.trts = x$nchar.trts,
  legend = TRUE,
  ...
)
```

Arguments

x	An object of class decomp.design.
digits.Q	Minimal number of significant digits for Q statistics, see print.default.
showall	A logical indicating whether results should be shown for all designs or only designs contributing to chi-squared statistics (default).
digits.pval.Q	Minimal number of significant digits for p-value of heterogeneity tests, see print.default.
digits.tau2	Minimal number of significant digits for between-study variance, see print.default.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
big.mark	A character used as thousands separator.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
legend	A logical indicating whether a legend should be printed.
...	Additional arguments (ignored at the moment).

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>, Ulrike Krahn <ulrike.krahn@bayer.com>

See Also

decomp.design

Examples

```
data(Senn2013)

# Only consider first five studies (to reduce runtime of example)
#
studies <- unique(Senn2013$studlab)
Senn2013.5 <- subset(Senn2013, studlab %in% studies[1:5])

# Conduct network meta-analysis with placebo as reference treatment
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013.5, sm = "MD", reference = "plac")

# Decomposition of Cochran's Q
#
decomp.design(net1)
```

print.netbind *Print method for objects of class netbind*

Description

Print method for objects of class netbind.

Usage

```
## S3 method for class 'netbind'
print(
  x,
  common = x$x$common,
  random = x$x$random,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netbind or summary.netbind.
common	A logical indicating whether results for the common effects model should be printed.
random	A logical indicating whether results for the random effects model should be printed.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netbind

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))
```

```
# Standard random effects NMA model (with placebo as reference
# treatment)
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, reference.group = "placebo",
  sm = "OR", common = FALSE)

# Additive CNMA model with placebo as inactive component and
# reference
#
nc1 <- netcomb(net1, inactive = "placebo")

# Combine results of standard NMA and CNMA
#
nb1 <- netbind(nc1, net1,
  name = c("Additive CNMA", "Standard NMA"),
  col.study = c("red", "black"), col.square = c("red", "black"))

nb1
print(nb1, common = TRUE)
```

`print.netcomb` *Print method for objects of class netcomb*

Description

Print method for objects of class `netcomb`.

Usage

```
## S3 method for class 'netcomb'
print(
  x,
  common = x$common,
  random = x$random,
  backtransf = x$backtransf,
  nchar.comps = x$nchar.comps,
  digits = gs("digits"),
  digits.stat = gs("digits.stat"),
  digits.pval = gs("digits.pval"),
  digits.pval.Q = max(gs("digits.pval.Q"), 2),
  digits.Q = gs("digits.Q"),
  digits.tau2 = gs("digits.tau2"),
  digits.tau = gs("digits.tau"),
  digits.I2 = gs("digits.I2"),
  scientific.pval = gs("scientific.pval"),
  zero.pval = gs("zero.pval"),
  JAMA.pval = gs("JAMA.pval"),
```

```
  big.mark = gs("big.mark"),
  text.tau2 = gs("text.tau2"),
  text.tau = gs("text.tau"),
  text.I2 = gs("text.I2"),
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netcomb or summary.netcomb.
common	A logical indicating whether results for the common effects model should be printed.
random	A logical indicating whether results for the random effects model should be printed.
backtransf	A logical indicating whether results should be back transformed in printouts and forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
nchar.comps	A numeric defining the minimum number of characters used to create unique names for components.
digits	Minimal number of significant digits, see print.default.
digits.stat	Minimal number of significant digits for z- or t-value, see print.default.
digits.pval	Minimal number of significant digits for p-value of overall treatment effect, see print.default.
digits.pval.Q	Minimal number of significant digits for p-value of heterogeneity tests, see print.default.
digits.Q	Minimal number of significant digits for heterogeneity statistics, see print.default.
digits.tau2	Minimal number of significant digits for between-study variance, see print.default.
digits.tau	Minimal number of significant digits for square root of between-study variance, see print.default.
digits.I2	Minimal number of significant digits for I-squared statistic, see print.default.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
zero.pval	A logical specifying whether p-values should be printed with a leading zero.
JAMA.pval	A logical specifying whether p-values for test of component or combination effect should be printed according to JAMA reporting standards.
big.mark	A character used as thousands separator.
text.tau2	Text printed to identify between-study variance τ^2.
text.tau	Text printed to identify τ, the square root of the between-study variance τ^2.
text.I2	Text printed to identify heterogeneity statistic I^2.
legend	A logical indicating whether a legend should be printed.

warn.deprecated
: A logical indicating whether warnings should be printed if deprecated arguments are used.

... : Additional arguments (to catch deprecated arguments).

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netcomb, discomb

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, reference.group = "placebo",
  sm = "OR", common = FALSE)

# Additive model for treatment components
#
nc1 <- netcomb(net1)
nc1
print(nc1, digits = 2, digits.stat = 3)

## Not run:
# Conduct random effects network meta-analysis
#
net2 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = Linde2016, reference.group = "placebo",
  sm = "OR", common = FALSE)

# Additive model for treatment components
#
nc2 <- netcomb(net2)
nc2
print(nc2, digits = 2, digits.stat = 3)

## End(Not run)
```

print.netimpact *Print method for objects of class netimpact*

Description

Print method for objects of class netimpact.

Usage

```
## S3 method for class 'netimpact'
print(
  x,
  common = x$x$common,
  random = x$x$random,
  digits = gs("digits.prop"),
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class netimpact.
common	A logical indicating whether results for the common effects model should be printed.
random	A logical indicating whether results for the random effects model should be printed.
digits	Minimal number of significant digits.
legend	A logical indicating whether a legend should be printed.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netimpact

Examples

```
data(Franchini2012)

# Only consider first two studies (to reduce runtime of example)
#
studies <- unique(Franchini2012$Study)
p1 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = subset(Franchini2012, Study %in% studies[1:2]),
  studlab = Study)

net1 <- netmeta(p1)
ni <- netimpact(net1, verbose = TRUE)
ni
```

---

print.summary.netcomb *Print detailed information for component network meta-analysis*

Description

Print detailed information for component network meta-analysis.

Usage

```
## S3 method for class 'summary.netcomb'
print(
  x,
  common = x$x$common,
  random = x$x$random,
  backtransf = x$backtransf,
  nchar.comps = x$nchar.comps,
  digits = gs("digits"),
  digits.stat = gs("digits.stat"),
  digits.pval = gs("digits.pval"),
  digits.pval.Q = max(gs("digits.pval.Q"), 2),
  digits.Q = gs("digits.Q"),
  scientific.pval = gs("scientific.pval"),
  zero.pval = gs("zero.pval"),
  JAMA.pval = gs("JAMA.pval"),
  big.mark = gs("big.mark"),
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class summary.netcomb
common	A logical indicating whether results for the common effects model should be printed.
random	A logical indicating whether results for the random effects model should be printed.
backtransf	A logical indicating whether results should be back transformed in printouts and forest plots. If backtransf=TRUE, results for sm="OR" are presented as odds ratios rather than log odds ratios, for example.
nchar.comps	A numeric defining the minimum number of characters used to create unique component names.
digits	Minimal number of significant digits, see print.default.
digits.stat	Minimal number of significant digits for z- or t-value, see print.default.
digits.pval	Minimal number of significant digits for p-value of overall treatment effect, see print.default.
digits.pval.Q	Minimal number of significant digits for p-value of heterogeneity tests, see print.default.
digits.Q	Minimal number of significant digits for heterogeneity statistics, see print.default.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
zero.pval	A logical specifying whether p-values should be printed with a leading zero.
JAMA.pval	A logical specifying whether p-values for test of effects should be printed according to JAMA reporting standards.
big.mark	A character used as thousands separator.
legend	A logical indicating whether a legend should be printed.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netcomb, discomb, summary.netcomb

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, reference.group = "placebo",
  sm = "OR", common = FALSE)

# Additive model for treatment components
#
nc1 <- netcomb(net1)
print(summary(nc1), digits = 2)
```

---

print.summary.netmeta *Print detailed results of network meta-analysis*

Description

Print method for objects of class summary.netmeta.

Usage

```
## S3 method for class 'summary.netmeta'
print(
  x,
  sortvar,
  common = x$x$common,
  random = x$x$random,
  prediction = x$prediction,
  reference.group = x$reference.group,
  baseline.reference = x$baseline.reference,
  all.treatments = x$all.treatments,
  details = TRUE,
  nma = TRUE,
  backtransf = x$backtransf,
  nchar.trts = x$nchar.trts,
  nchar.studlab = x$nchar.studlab,
  digits = gs("digits"),
  digits.se = gs("digits.se"),
  digits.pval.Q = max(gs("digits.pval.Q"), 2),
  digits.Q = gs("digits.Q"),
```

```
  digits.tau2 = gs("digits.tau2"),
  digits.I2 = gs("digits.I2"),
  scientific.pval = gs("scientific.pval"),
  big.mark = gs("big.mark"),
  truncate,
  text.truncate = "*** Output truncated ***",
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

x	An object of class summary.netmeta.
sortvar	An optional vector used to sort individual studies (must be of same length as x$TE).
common	A logical indicating whether results for the common effects model should be printed.
random	A logical indicating whether results for the random effects model should be printed.
prediction	A logical indicating whether prediction intervals should be printed.
reference.group	Reference treatment.
baseline.reference	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa. This argument is only considered if reference.group has been specified.
all.treatments	A logical or "NULL". If TRUE, matrices with all treatment effects, and confidence limits will be printed.
details	A logical indicating whether further details for individual studies should be printed.
nma	A logical indicating whether summary results of network meta-analysis should be printed.
backtransf	A logical indicating whether results should be back transformed in printouts and forest plots. If backtransf = TRUE, results for sm = "OR" are presented as odds ratios rather than log odds ratios, for example.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
nchar.studlab	A numeric defining the minimum number of characters used to create unique study labels.
digits	Minimal number of significant digits, see print.default.
digits.se	Minimal number of significant digits for standard deviations and standard errors, see print.default.
digits.pval.Q	Minimal number of significant digits for p-value of heterogeneity tests, see print.default.

digits.Q	Minimal number of significant digits for heterogeneity statistics, see print.default.
digits.tau2	Minimal number of significant digits for between-study variance, see print.default.
digits.I2	Minimal number of significant digits for I-squared statistic, see print.default.
scientific.pval	A logical specifying whether p-values should be printed in scientific notation, e.g., 1.2345e-01 instead of 0.12345.
big.mark	A character used as thousands separator.
truncate	An optional vector used to truncate the printout of results for individual studies (must be a logical vector of length corresponding to the number of pairwise comparisons x$TE or contain numerical values).
text.truncate	A character string printed if study results were truncated from the printout.
legend	A logical indicating whether a legend should be printed.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netmeta, summary.netmeta

Examples

```
data(Senn2013)

# Conduct common effects network meta-analysis
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", random = FALSE, ref = "plac")
snet1 <- summary(net1)
print(snet1, digits = 3)

# Only show individual study results for multi-arm studies
#
print(snet1, digits = 3, truncate = multiarm)

# Only show first three individual study results
#
print(snet1, digits = 3, truncate = 1:3)

# Only show individual study results for Kim2007 and Willms1999
#
print(snet1, digits = 3, truncate = c("Kim2007", "Willms1999"))
```

```
# Only show individual study results for studies starting with the
# letter "W"
#
print(snet1, ref = "plac", digits = 3,
  truncate = substring(studlab, 1, 1) == "W")

## Not run:
# Conduct random effects network meta-analysis
#
net2 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", common = FALSE, ref = "plac")
print(summary(net2), digits = 3)

## End(Not run)
```

---

rankogram *Calculate rankogram*

Description

This function calculates the probabilities of each treatment being at each possible rank and the SUCRAs (Surface Under the Cumulative RAnking curve) in frequentist network meta-analysis.

Usage

```
rankogram(
  x,
  nsim = 1000,
  common = x$common,
  random = x$random,
  small.values = x$small.values,
  cumulative.rankprob = FALSE,
  nchar.trts = x$nchar.trts,
  warn.deprecated = gs("warn.deprecated"),
  ...
)

## S3 method for class 'rankogram'
print(
  x,
  common = x$common,
  random = x$random,
  cumulative.rankprob = x$cumulative.rankprob,
  nchar.trts = x$nchar.trts,
  digits = gs("digits.prop"),
  legend = TRUE,
  warn.deprecated = gs("warn.deprecated"),
```

```
  ...
)
```

Arguments

x	An object of class netmeta.
nsim	Number of simulations.
common	A logical indicating to compute ranking probabilities and SUCRAs for the common effects model.
random	A logical indicating to compute ranking probabilities and SUCRAs for the random effects model.
small.values	A character string specifying whether small treatment effects indicate a beneficial ("good") or harmful ("bad") effect, can be abbreviated.
cumulative.rankprob	A logical indicating whether cumulative ranking probabilites should be printed.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments for printing.
digits	Minimal number of significant digits, see print.default.
legend	A logical indicating whether a legend should be printed.

Details

We derive a matrix showing the probability of each treatment being at each possible rank. To this aim, we use resampling from a multivariate normal distribution with estimated network effects as means and corresponding estimated variance covariance matrix. We then summarise them using the ranking metric SUCRAs (Surface Under the Cumulative RAnking curve).

Value

An object of class rankogram with corresponding print and plot function. The object is a list containing the following components:

ranking.matrix.common	Numeric matrix giving the probability of each treatment being at each possible rank for the common effects model.
ranking.common	SUCRA values for the common effects model.
ranking.matrix.random	Numeric matrix giving the probability of each treatment being at each possible rank for the random effects model.
ranking.random	SUCRA values for the random effects model.
cumrank.matrix.common	Numeric matrix giving the cumulative ranking probability of each treatment for the common effects model.

`cumrank.matrix.random`
: Numeric matrix giving the cumulative ranking probability of each treatment for the random effects model.

`nsim, common, random`
: As defined above

,

`small.values, x`
: As defined above

,

Author(s)

Theodoros Papakonstantinou <dev@tpapak.com>, Guido Schwarzer <sc@imbi.uni-freiburg.de>

References

Salanti G, Ades AE, Ioannidis JP (2011): Graphical methods and numerical summaries for presenting results from multiple-treatment meta-analysis: an overview and tutorial. *Journal of Clinical Epidemiology*, **64**, 163–71

See Also

netmeta, netrank

Examples

```
data(Woods2010)
p1 <- pairwise(treatment, event = r, n = N, studlab = author,
               data = Woods2010, sm = "OR")
net1 <- netmeta(p1, small.values = "good")

ran1 <- rankogram(net1, nsim = 100)
ran1
print(ran1, cumulative.rankprob = TRUE)

plot(ran1)
```

Senn2013 *Network meta-analysis in diabetes*

Description

Network meta-analysis in diabetes comparing effects of a number of drugs on the HbA1c value.

These data are used as an example in Senn et al. (2013) and have been preprocessed for use in R package netmeta.

Format

A data frame with the following columns:

TE	treatment effect
seTE	standard error of treatment effect
treat1	treatment 1
treat2	treatment 2
treat1.long	treatment 1 (full treatment names)
treat2.long	treatment 2 (full treatment names)
studlab	Study label

Details

Treatment labels provided by columns `treat1` and `treat2` have been abbreviated:

- acar = Acarbose
- benf = Benfluorex
- metf = Metformin
- migl = Miglitol
- piog = Pioglitazone
- plac = Placebo
- rosi = Rosiglitazone
- sita = Sitagliptin
- sulf = Sulfonylurea
- vild = Vildagliptin

Full treatment names are available in columns `treat1.long` and `treat2.long`.

Source

Senn S, Gavini F, Magrez D, Scheen A (2013): Issues in performing a network meta-analysis. *Statistical Methods in Medical Research*, **22**, 169–89

See Also

netmeta

Examples

```
data(Senn2013)

# Common effects model
#
net1 <- netmeta(TE, seTE, treat1.long, treat2.long, studlab,
  data = Senn2013, sm = "MD", random = FALSE, nchar.trts = 4)
net1
net1$Q.decomp
```

```
# Forest plot
#
forest(net1, ref = "plac")

## Not run:
# Comparison with reference group
#
netmeta(TE, seTE, treat1.long, treat2.long,
  studlab, data = Senn2013, reference = "plac")

# Random effects model
#
net2 <- netmeta(TE, seTE, treat1.long, treat2.long, studlab,
  data = Senn2013, common = FALSE)
net2
forest(net2, ref = "plac")

## End(Not run)
```

`smokingcessation` *Network meta-analysis of interventions for smoking cessation*

Description

Network meta-analysis comparing the effects of a number of interventions for smoking cessation.

These data are used as an example in Dias et al. (2013), page 651.

Format

A data frame with the following columns:

event1	number of individuals with successful smoking cessation in arm 1
n1	number of individuals in arm 1
event2	number of individuals with successful smoking cessation in arm 2
n2	number of individuals in arm 2
event3	number of individuals with successful smoking cessation in arm 3
n3	number of individuals in arm 3
treat1	treatment 1
treat2	treatment 2
treat3	treatment 3

Source

Dias S, Welton NJ, Sutton AJ, Caldwell DM, Lu G and Ades AE (2013): Evidence Synthesis for Decision Making 4: Inconsistency in networks of evidence based on randomized controlled trials. *Medical Decision Making*, **33**, 641–56

See Also

pairwise, metabin, netmeta, netgraph.netmeta

Examples

```
data(smokingcessation)

# Transform data from arm-based format to contrast-based format
# Argument 'sm' has to be used for odds ratio as summary measure;
# by default the risk ratio is used in the metabin function called
# internally.
#
p1 <- pairwise(list(treat1, treat2, treat3),
  event = list(event1, event2, event3), n = list(n1, n2, n3),
  data = smokingcessation, sm = "OR")
p1

# Conduct network meta-analysis
#
net1 <- netmeta(p1)
net1

# Draw network graph
#
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.25)
tname <- c("No intervention", "Self-help",
  "Individual counselling", "Group counselling")
netgraph(net1, points = TRUE, cex.points = 3, cex = 1.25, labels = tname)
```

Stowe2010 *Network meta-analysis of adjuvant treatments to levodopa therapy for Parkinson's disease*

Description

This data set contains data from a Cochrane review assessing efficacy and safety of three drug classes as adjuvant treatment to levodopa therapy in patients with Parkinson's disease and motor complications (Stowe et al., 2010). The authors conducted three pairwise meta-analyses comparing dopamine agonists, catechol-O-methyl transferase inhibitors (COMTIs), and monoamine oxidase type B inhibitors (MAOBIs), respectively, with placebo.

The primary outcome was the mean reduction of the time spent in a relatively immobile 'off' phase (mean off-time), calculated in hours per day. Relative treatment effects were expressed as mean difference. Data on this outcome were available for 5,331 patients from 28 studies comparing an active treatment with placebo and one three-arm study comparing two active treatments with placebo.

Format

A data frame with the following columns:

study	study label
id	study id
t1	treatment 1
y1	treatment effect arm 1
sd1	Standard deviation arm 1
n1	Sample size arm 1
t2	treatment 2
y2	treatment effect arm 2
sd2	Standard deviation arm 2
n2	Sample size arm 2
t3	treatment 3
y3	treatment effect arm 3
sd3	Standard deviation arm 3
n3	Sample size arm 3

Source

Stowe R, Ives N, Clarke CE, Deane K, Hilten V, Wheatley K, et al. (2010): Evaluation of the efficacy and safety of adjuvant treatment to levodopa therapy in Parkinson's disease patients with motor complications. *Cochrane Database of Systematic Reviews*

See Also

pairwise, metacont, netmeta, netgraph.netmeta

Examples

```
data(Stowe2010)

# Transform data from arm-based format to contrast-based format
#
p1 <- pairwise(list(t1, t2, t3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = Stowe2010, studlab = study)
p1

# Conduct network meta-analysis
net1 <- netmeta(p1, ref = "plac")
net1
```

---

subset.pairwise *Return subset of pairwise object*

Description

The subset method returns a subset of a pairwise object.

Usage

```
## S3 method for class 'pairwise'
subset(x, subset, ...)
```

Arguments

x	An object of class pairwise.
subset	A logical expression indicating elements or rows to keep: missing values are taken as false.
...	Additional arguments.

Value

A pairwise object is returned.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

pairwise

Examples

```
# Transform data from arm-based format to contrast-based format
data(Franchini2012)
p1 <- pairwise(list(Treatment1, Treatment2, Treatment3),
  n = list(n1, n2, n3),
  mean = list(y1, y2, y3), sd = list(sd1, sd2, sd3),
  data = Franchini2012, studlab = Study)
p1[, 1:5]

# Subset without Lieberman studies
subset(p1, !grepl("Lieberman", studlab))[, 1:5]
```

---

summary.netcomb *Summary method for objects of class netcomb*

Description

Summary method for objects of class netcomb.

Usage

```
## S3 method for class 'netcomb'
summary(
  object,
  common = object$common,
  random = object$random,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

object	An object of class netcomb.
common	A logical indicating whether results for the common effects model should be printed.
random	A logical indicating whether results for the random effects model should be printed.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).

Value

A list is returned with the same elements as a netcomb object.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netcomb, discomb

Examples

```
data(Linde2016)

# Only consider studies including Face-to-face PST (to reduce
# runtime of example)
#
face <- subset(Linde2016, id %in% c(16, 24, 49, 118))

# Conduct random effects network meta-analysis
#
net1 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = face, reference.group = "placebo",
  sm = "OR", common = FALSE)
```

```
# Additive model for treatment components
#
nc1 <- netcomb(net1)
summary(nc1)
print(summary(nc1), digits = 2, digits.stat = 3)

## Not run:
# Conduct random effects network meta-analysis
#
net2 <- netmeta(lnOR, selnOR, treat1, treat2, id,
  data = Linde2016, reference.group = "placebo",
  sm = "OR", common = FALSE)

# Additive model for treatment components
#
nc2 <- netcomb(net2)
summary(nc2)
print(summary(nc2), digits = 2, digits.stat = 3)

## End(Not run)
```

`summary.netmeta` *Summary method for objects of class netmeta*

Description

Summary method for objects of class `netmeta`.

Usage

```
## S3 method for class 'netmeta'
summary(
  object,
  common = object$common,
  random = object$random,
  prediction = object$prediction,
  reference.group = object$reference.group,
  baseline.reference = object$baseline.reference,
  all.treatments = object$all.treatments,
  warn.deprecated = gs("warn.deprecated"),
  ...
)
```

Arguments

`object`	An object of class `netmeta`.
`common`	A logical indicating whether results for the common effects model should be printed.

random	A logical indicating whether results for the random effects model should be printed.
prediction	A logical indicating whether prediction intervals should be printed.
reference.group	Reference treatment.
baseline.reference	A logical indicating whether results should be expressed as comparisons of other treatments versus the reference treatment (default) or vice versa. This argument is only considered if reference.group has been specified.
all.treatments	A logical or "NULL". If TRUE, matrices with all treatment effects, and confidence limits will be printed.
warn.deprecated	A logical indicating whether warnings should be printed if deprecated arguments are used.
...	Additional arguments (to catch deprecated arguments).

Value

A list is returned with the following elements:

comparison	Results for pairwise comparisons (data frame with columns studlab, treat1, treat2, TE, seTE, lower, upper, z, p).
comparison.nma.common	Results for pairwise comparisons based on common effects model (data frame with columns studlab, treat1, treat2, TE, seTE, lower, upper, z, p, leverage).
comparison.nma.random	Results for pairwise comparisons based on random effects model (data frame with columns studlab, treat1, treat2, TE, seTE, lower, upper, z, p).
common	Results for common effects model (a list with elements TE, seTE, lower, upper, z, p).
random	Results for random effects model (a list with elements TE, seTE, lower, upper, z, p).
predict	Prediction intervals (a list with elements seTE, lower, upper).
studies	Study labels coerced into a factor with its levels sorted alphabetically.
narms	Number of arms for each study.
k	Total number of studies.
m	Total number of pairwise comparisons.
n	Total number of treatments.
d	Total number of designs (corresponding to the unique set of treatments compared within studies).
Q	Overall heterogeneity / inconsistency statistic.
df.Q	Degrees of freedom for test of heterogeneity / inconsistency.
pval.Q	P-value for test of heterogeneity / inconsistency.

`I2`, `lower.I2`, `upper.I2`
: I-squared, lower and upper confidence limits.

`tau`
: Square-root of between-study variance.

`Q.heterogeneity`
: Overall heterogeneity statistic.

`df.Q.heterogeneity`
: Degrees of freedom for test of overall heterogeneity.

`pval.Q.heterogeneity`
: P-value for test of overall heterogeneity.

`Q.inconsistency`
: Overall inconsistency statistic.

`df.Q.inconsistency`
: Degrees of freedom for test of overall inconsistency.

`pval.Q.inconsistency`
: P-value for test of overall inconsistency.

`sm`
: A character string indicating underlying summary measure.

`method`
: A character string indicating which method is to be used for pooling of studies.

`level`
: The level used to calculate confidence intervals for individual studies.

`level.ma`
: The level used to calculate confidence intervals for pooled estimates.

`common`, `random`
: As defined above.

`prediction`, `level.predict`
: As defined above.

`reference.group`, `baseline.reference`
: As defined above.

`all.treatments`, `backtransf`
: As defined above.

`ci.lab`
: Label for confidence interval.

`seq`
: A character specifying the sequence of treatments.

`tau.preset`
: An optional value for the square-root of the between-study variance τ^2.

`sep.trts`
: A character used in comparison names as separator between treatment labels.

`nchar.trts`
: A numeric defining the minimum number of characters used to create unique treatment names.

`title`
: Title of meta-analysis / systematic review.

`call`
: Function call.

`version`
: Version of R package netmeta used to create object.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

netmeta

Examples

```
data(Senn2013)

# Conduct common effects network meta-analysis
#
net1 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", random = FALSE)
print(net1, ref = "plac", digits = 3)
summary(net1)

## Not run:
# Conduct random effects network meta-analysis
#
net2 <- netmeta(TE, seTE, treat1, treat2, studlab,
  data = Senn2013, sm = "MD", common = FALSE)
print(net2, ref = "plac", digits = 3)
summary(net2)

## End(Not run)
```

---

treats *Abbreviate treatment names*

Description

Auxiliary functions to create uniquely abbreviated treatment names.

Usage

```
treats(x, nchar.trts = 8, row = TRUE)

comps(x, trts, sep.trts, nchar.trts = 8, row = TRUE)
```

Arguments

x	A vector with treatment or comparison names or a matrix with treatment or comparison names as row and / or column names.
nchar.trts	A numeric defining the minimum number of characters used to create unique treatment names.
row	A logical indicating whether row or column names should be used (only considered if argument x is a matrix).
trts	A character vector with treatment names.
sep.trts	A character used in comparison names as separator between treatment labels.

Details

These auxiliary functions can be used to create uniquely abbreviated treatment names (and are used internally in several R functions for this purpose).

In order to construct uniquely abbreviated treatment names, `treats` uses `substring` to extract the first `nchar.trts` characters. If these abbreviated treatment names are not unique, `abbreviate` with argument `minlength` = `nchar.trts` is used.

In order to construct comparisons with uniquely abbreviated treatment names, `comps` calls `treats` internally.

Author(s)

Guido Schwarzer <sc@imbi.uni-freiburg.de>

See Also

`netmeta`, `print.netmeta`, `print.summary.netmeta`

Examples

```
data(Senn2013)
#
net1 <- netmeta(TE, seTE, treat1.long, treat2.long, studlab,
  data = Senn2013)

# Full treatment names
#
net1$trts

# Treatment names with four characters
#
treats(net1$trts, nchar.trts = 4)

# With two characters
#
treats(net1$trts, nchar.trts = 2)

# With one character (if possible)
#
treats(net1$trts, nchar.trts = 1)

# Full comparison names
#
net1$comparisons

# Abbreviated comparison names
#
with(net1, comps(comparisons, trts, sep.trts, nchar = 4))
```

Woods2010 *Count statistics of survival data*

Description

Count mortality statistics in randomised controlled trials of treatments for chronic obstructive pulmonary disease (Woods et al. (2010), Table 1).

Format

A data frame with the following columns:

author	first author / study name
treatment	treatment
r	number of deaths in treatment arm
N	number of patients in treatment arm

Source

Woods BS, Hawkins N, Scott DA (2010): Network meta-analysis on the log-hazard scale, combining count and hazard ratio statistics accounting for multi-arm trials: A tutorial. *BMC Medical Research Methodology*, **10**, 54

See Also

pairwise, metabin, netmeta

Examples

```
data(Woods2010)

# Transform data from long arm-based format to contrast-based
# format Argument 'sm' has to be used for odds ratio as summary
# measure; by default the risk ratio is used in the metabin
# function called internally.
#
p1 <- pairwise(treatment, event = r, n = N,
  studlab = author, data = Woods2010, sm = "OR")
p1

# Conduct network meta-analysis
#
net1 <- netmeta(p1)
net1

## Not run:
# Show forest plot
#
forest(net1, ref = "Placebo", drop = TRUE,
```

```
  leftlabs = "Contrast to Placebo")

## End(Not run)
```

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